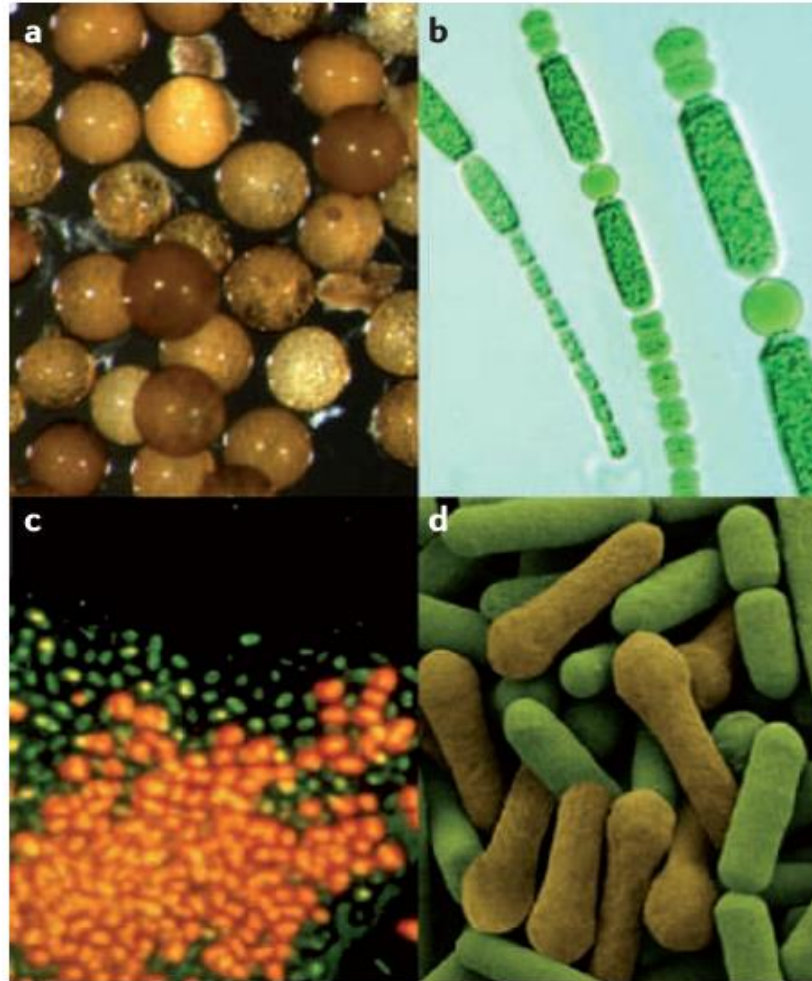


KITP, July 2014

Single-Cell Variability of Growth in Bacteria as an Optimization Process

Nathalie Q. Balaban
Racah Institute
The Hebrew University
Jerusalem, Israel

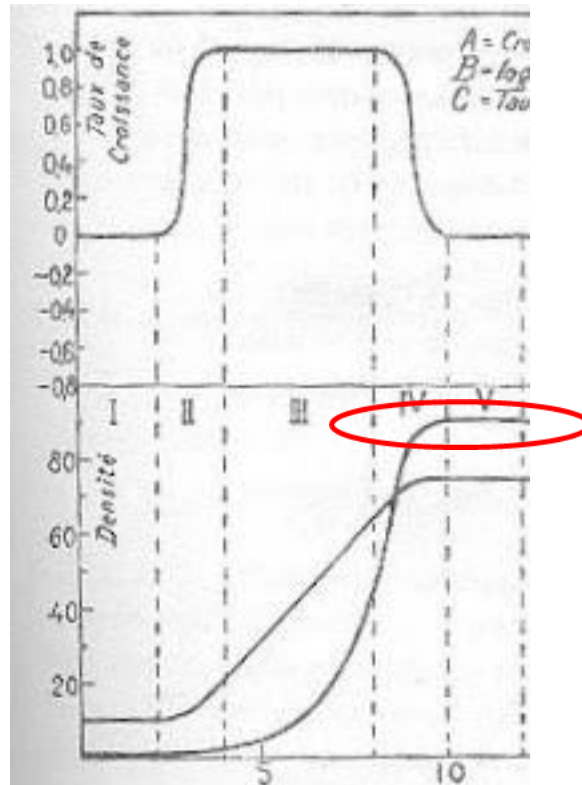
Growth arrest is the prevalent state of microorganisms in nature



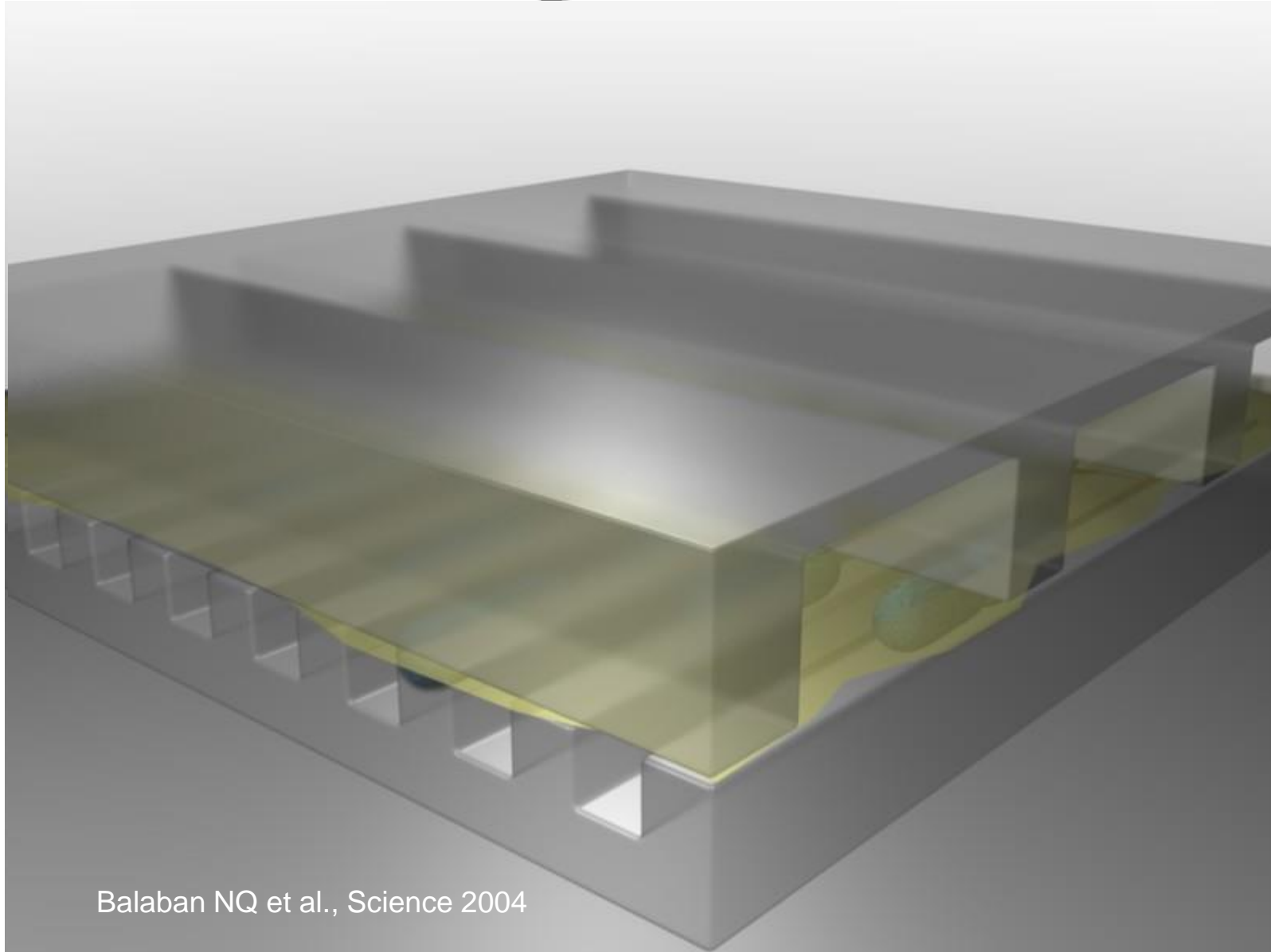
Examples of microbial dormancy.

Lennon JT & Jone SE, *Nat Rev Micro* (2011)

Typical bacterial growth curve



Microfluidic devices for following single-cells

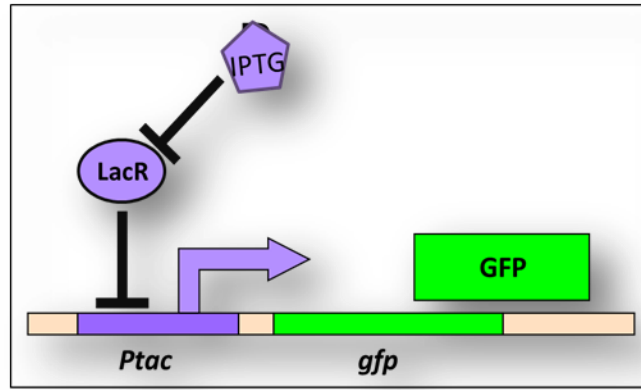


Balaban NQ et al., Science 2004

Growth arrest in microfluidic devices

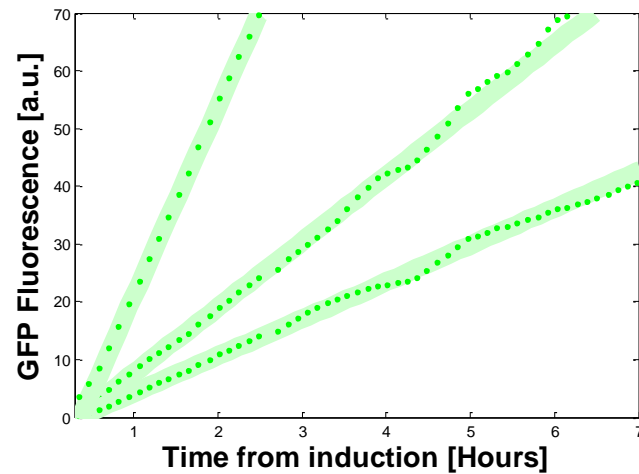
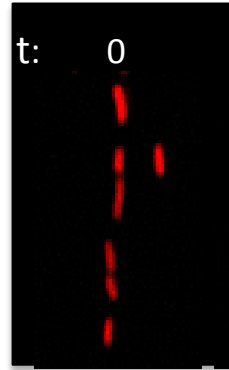


Measuring the activity of single growth arrested cells



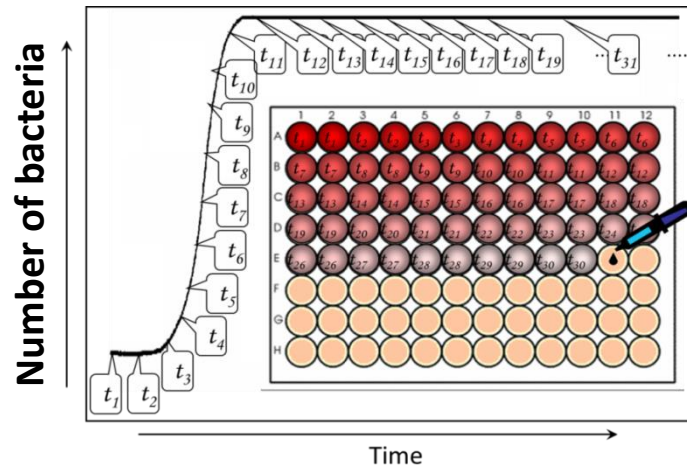
Do stationary phase bacteria respond to induction?

Fluorescence induction of growth arrested cells in microfluidic devices



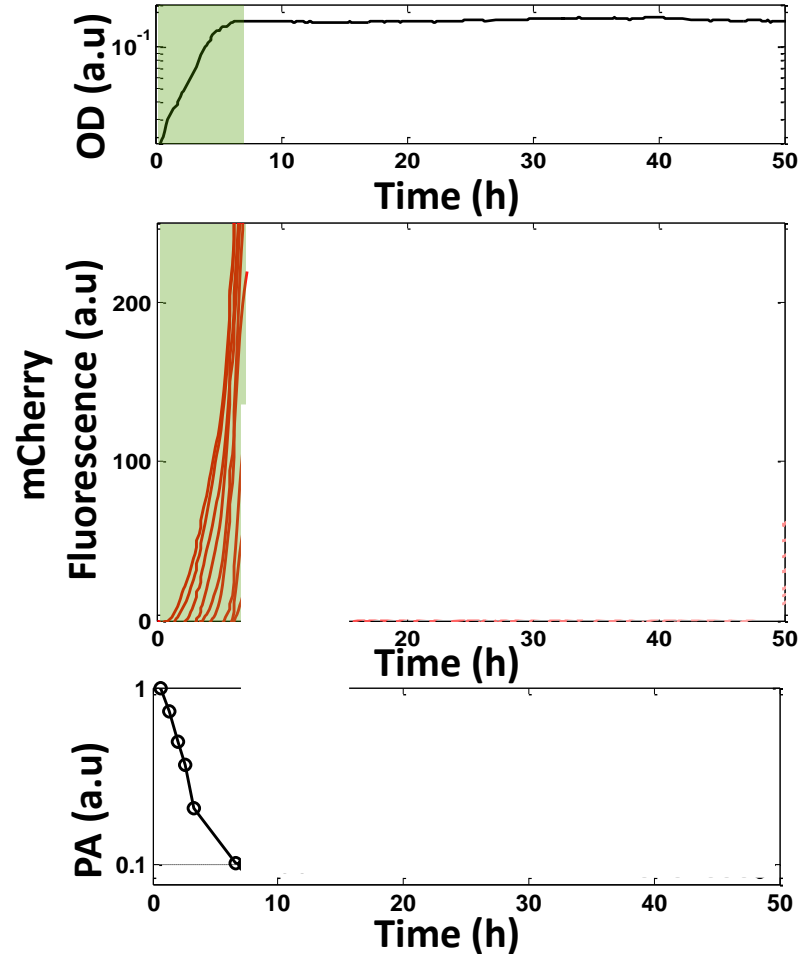
CASP : Constant Activity
Stationary Phase

CASP: Constant Activity Stationary Phase in batch cultures

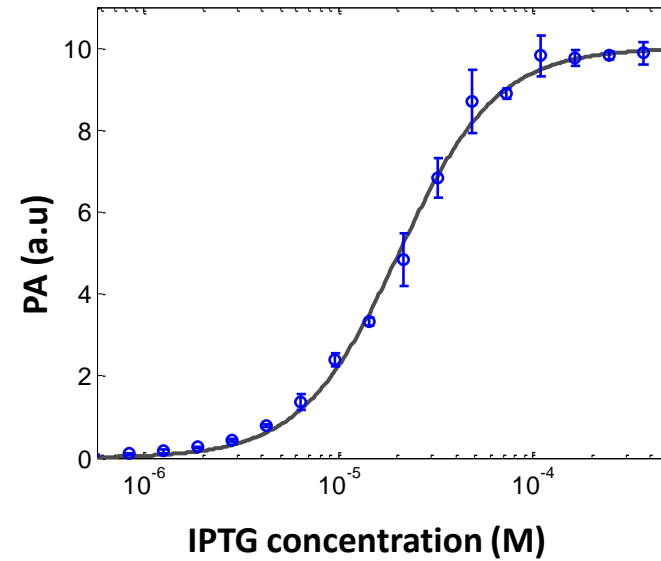
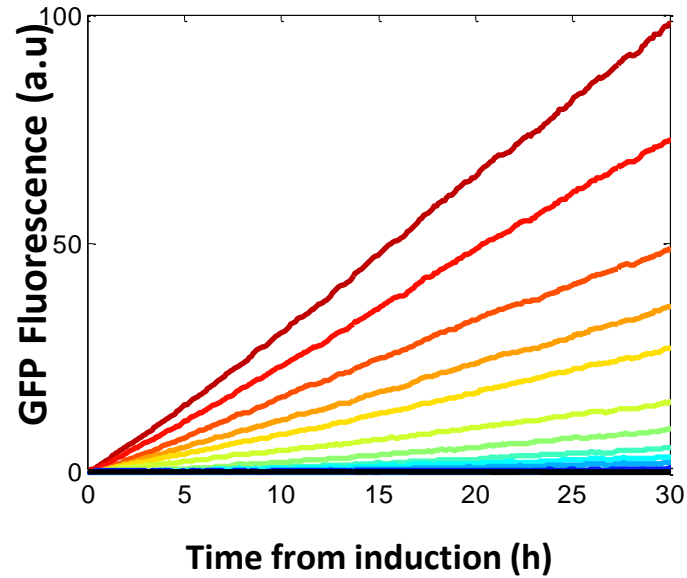


At CASP, cells produce fluorescence at a constant rate for several days, despite the absence of growth

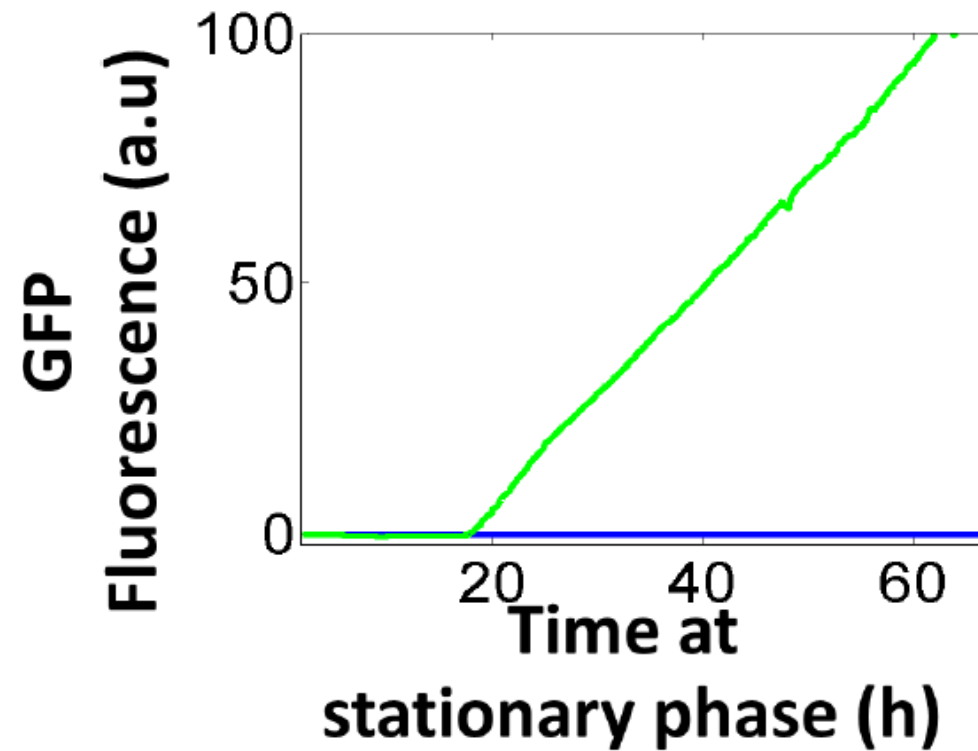
Gefen O. *et al.*, PNAS (2014)



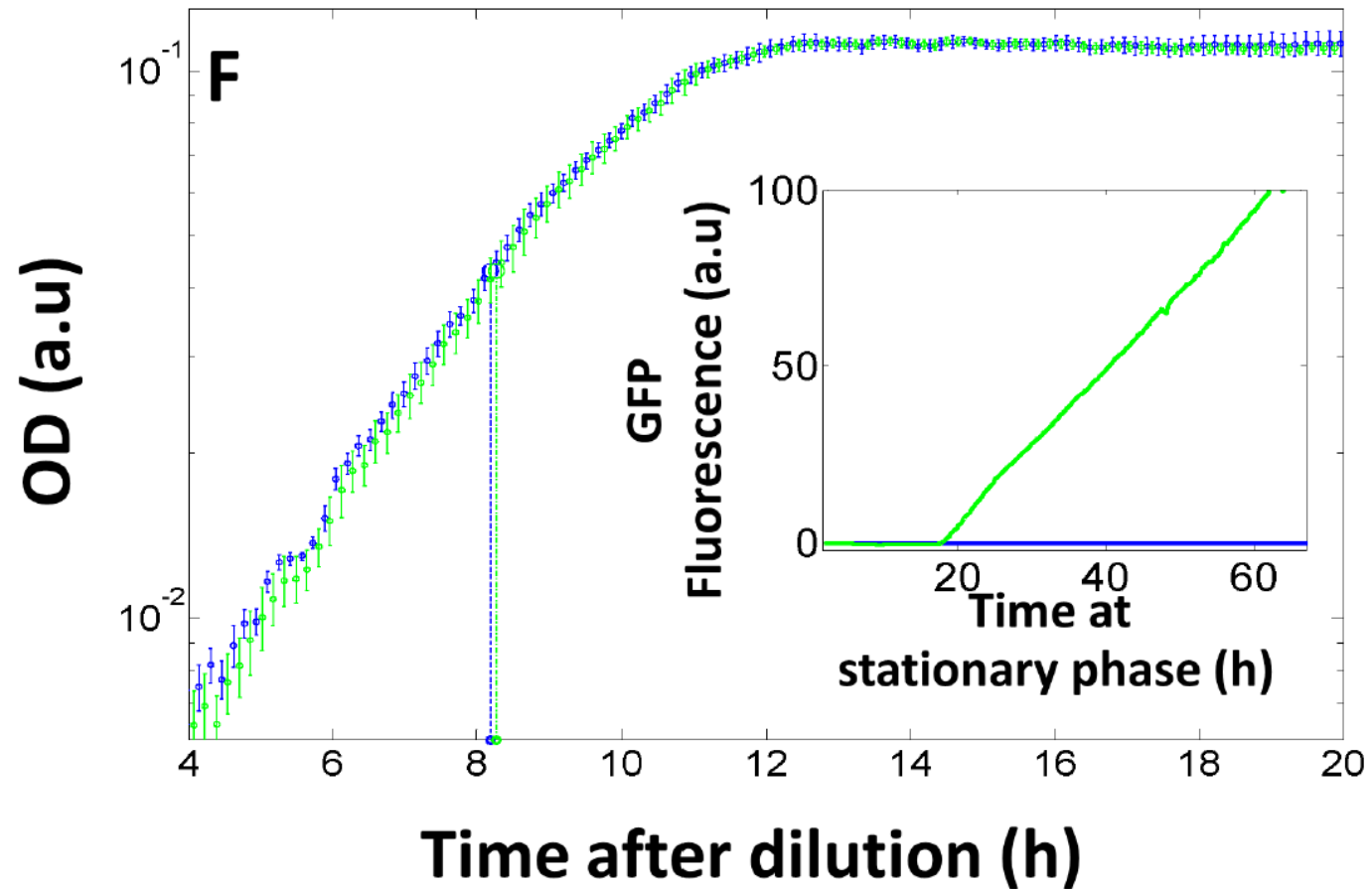
Precise promoter activity measurements at CASP



Fitness cost of protein production at CASP

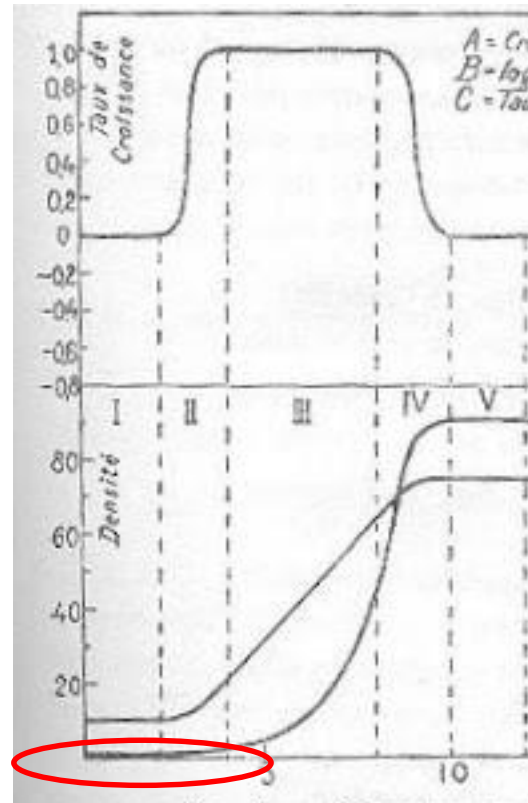


No detectable cost of protein production at CASP

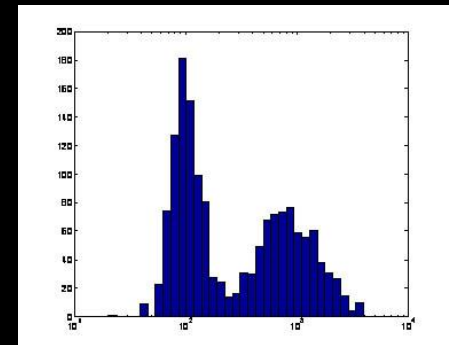
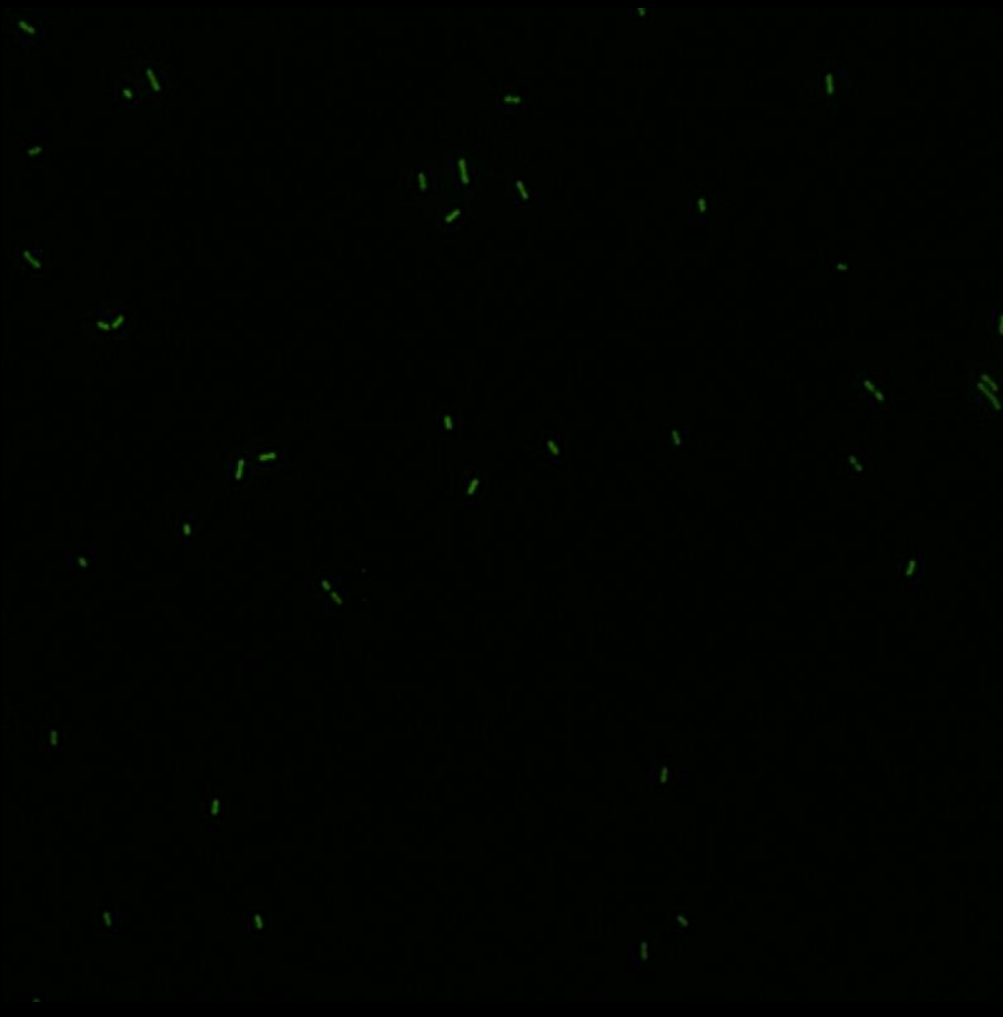




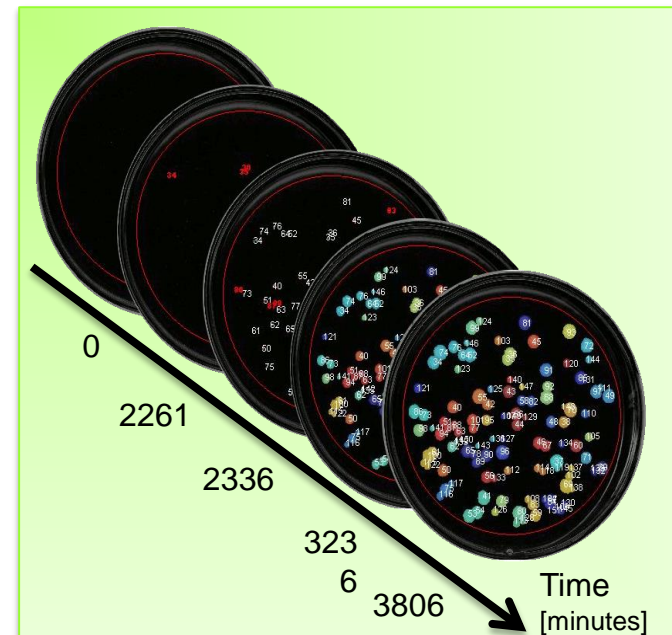
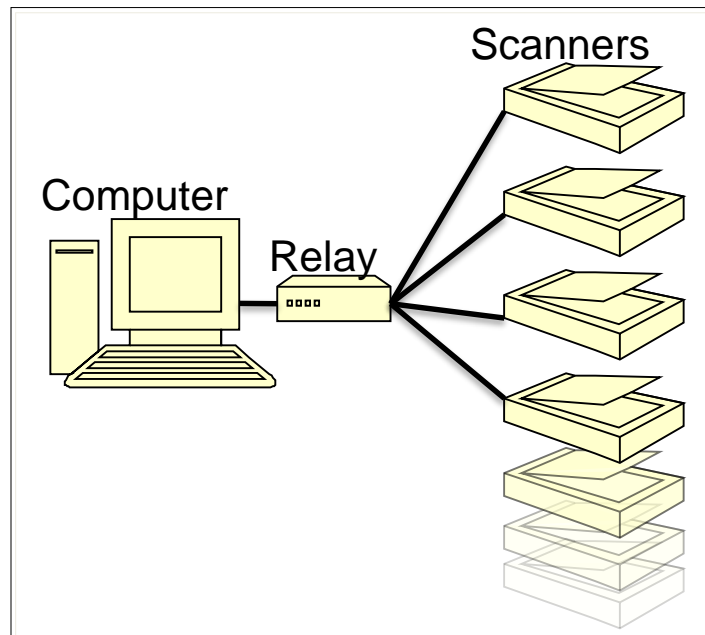
Typical bacterial growth curve



Growth arrest during the lag phase

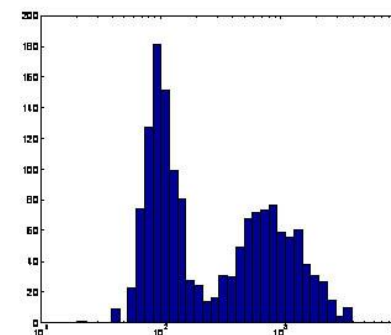


ScanLag: High throughput assay for lag variability quantification

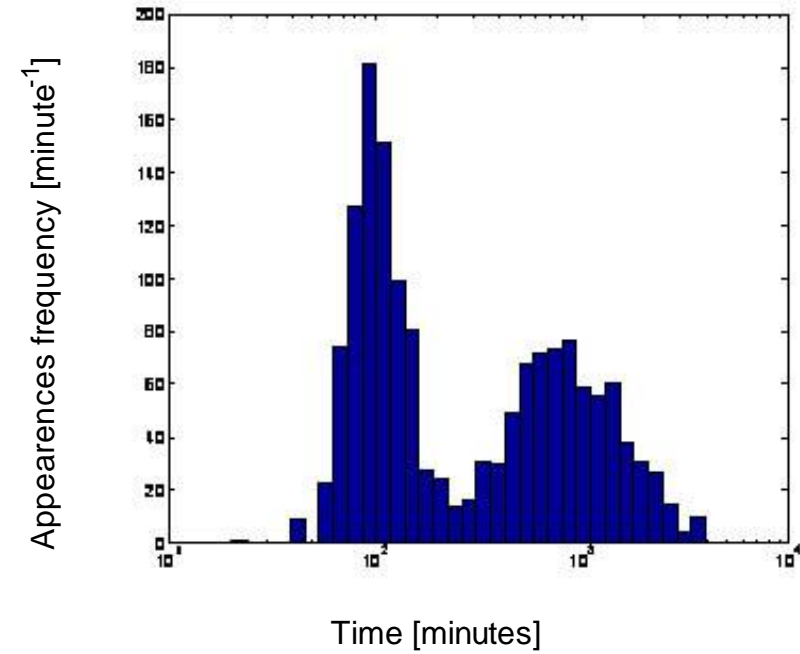
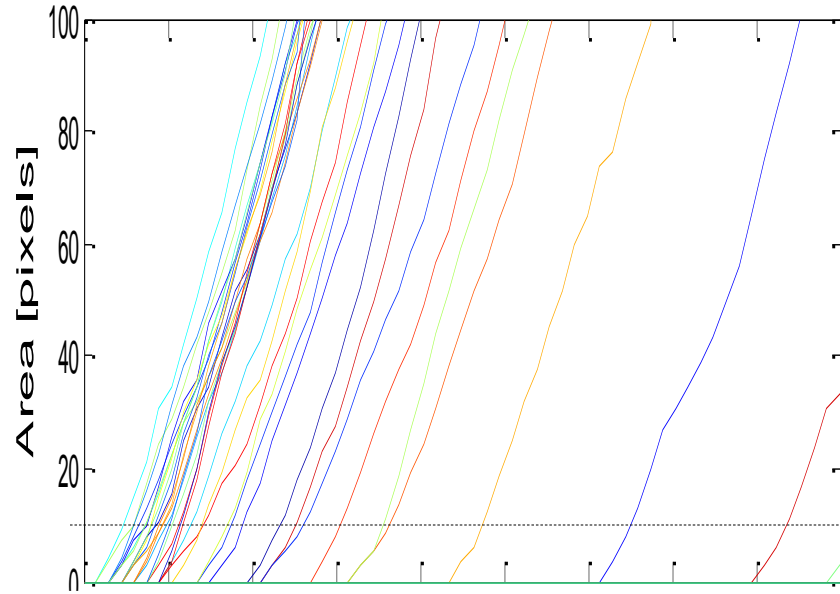


Irit Levin-Reisman *et al.* Nature Methods (2010)

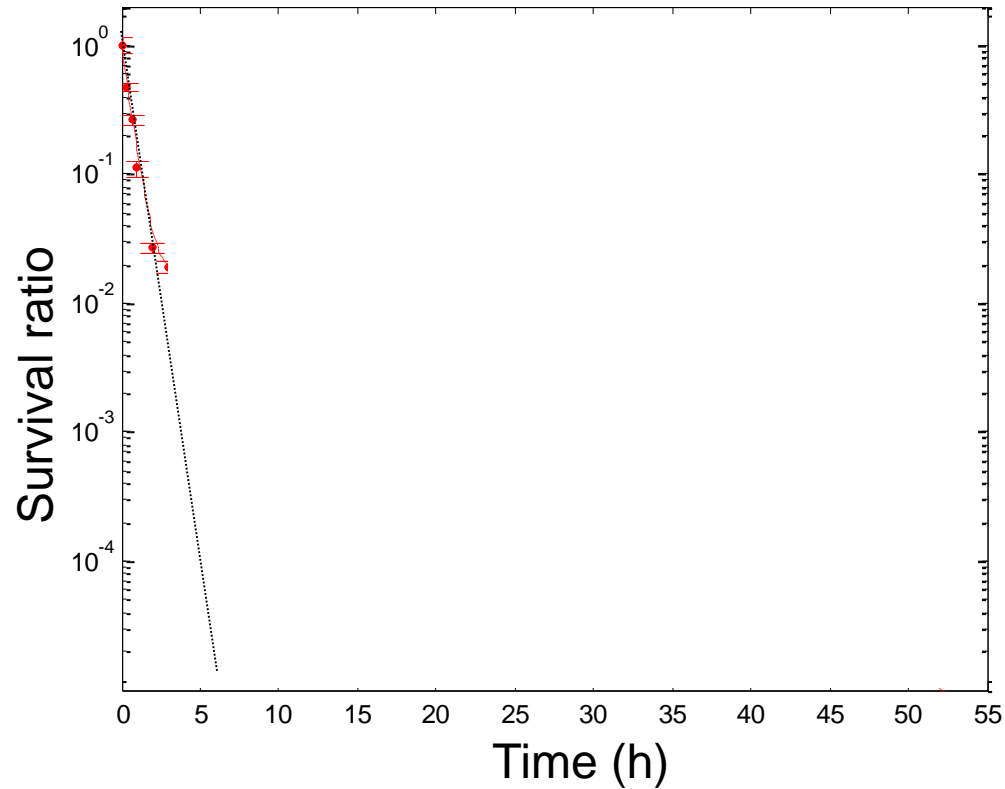
Irit Levin-Reisman *et al.* Jove (in press)



Appearance distribution can reflect growth arrest distribution



Persistence to antibiotics

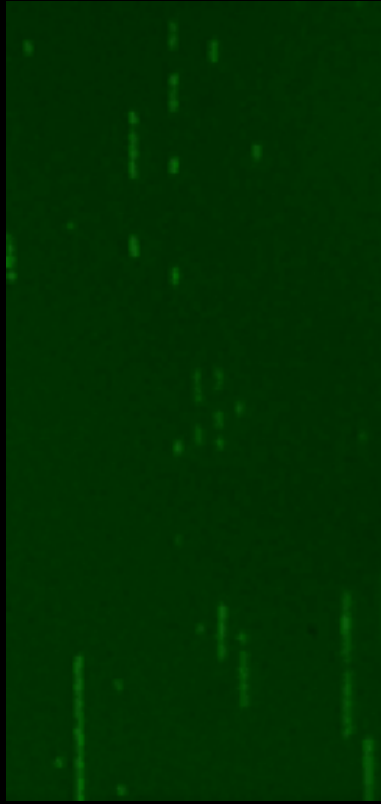


$$N(t) \equiv N_0 (\alpha e^{-\mu_1 t} + (1-\alpha)e^{-\mu_2 t})$$

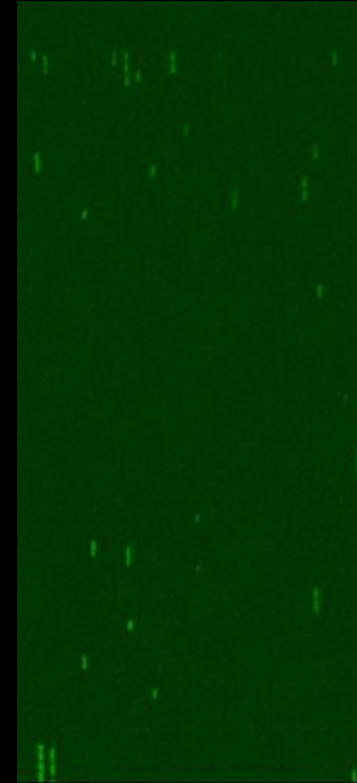
Balaban Curr Opi Gen & Dev 2011

Bigger, The Lancet 1944

Direct observation of the single cell response to antibiotics

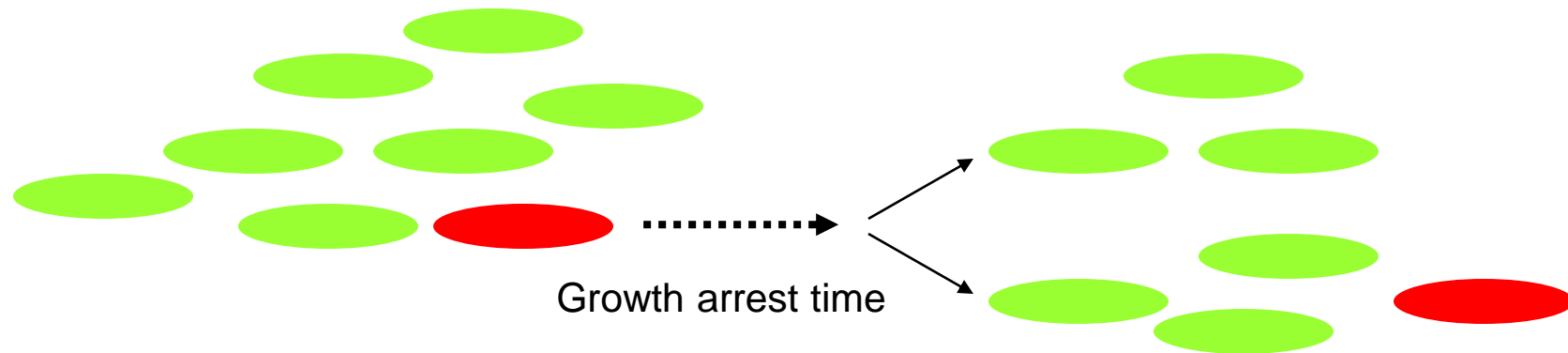


Persistence is not due to a mutation



High persistence is due to a sub-
population of growth arrested
bacteria

Extended lag variability as a bet-hedging strategy



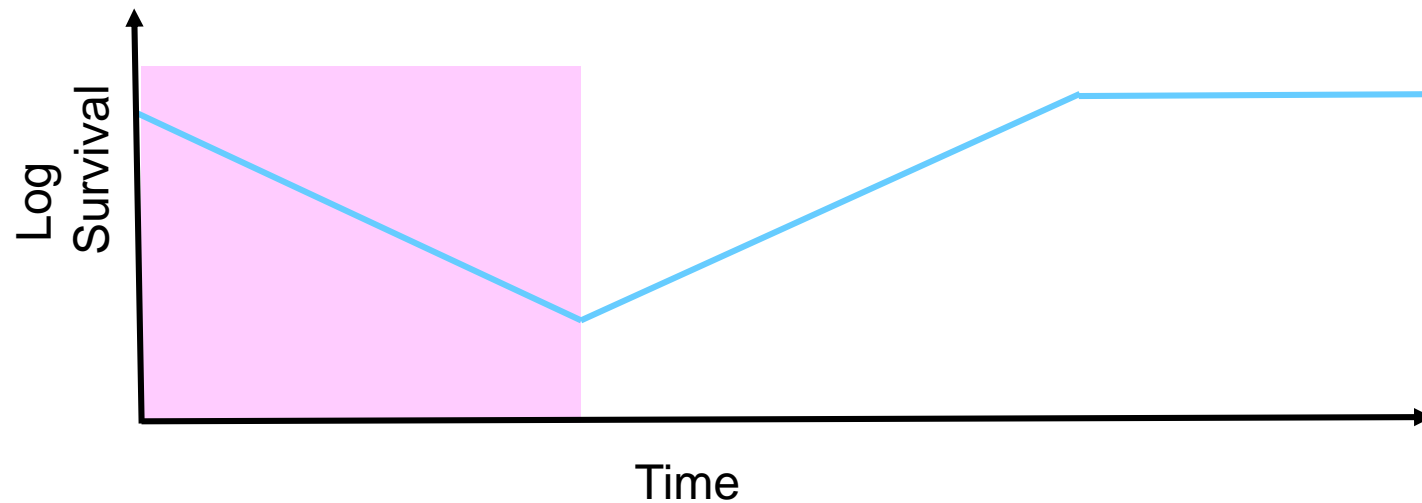
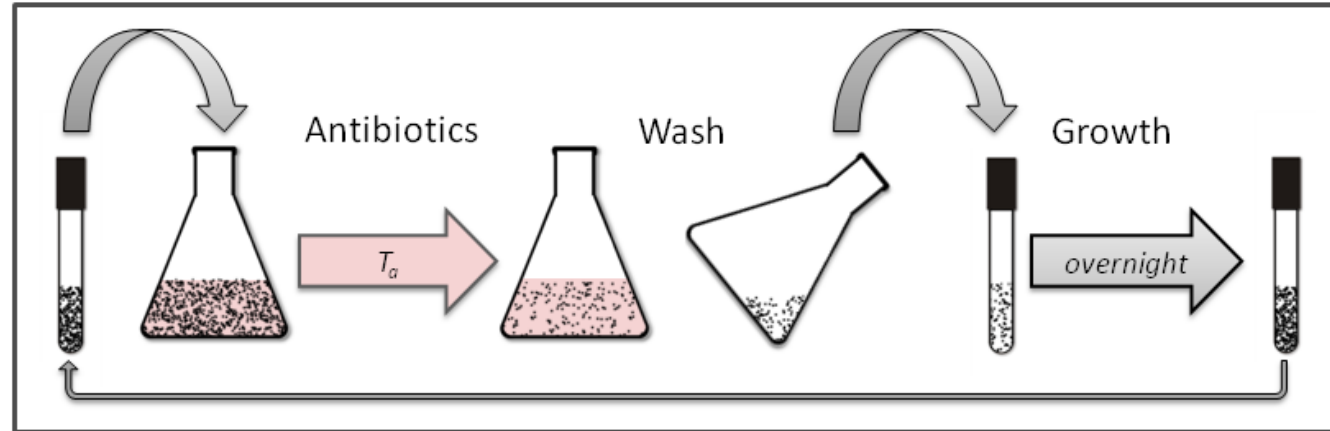
Main Questions

- **Molecular level:** What controls the duration of the lag?

Main Questions

- **Molecular level:** What controls the duration of the lag?
- **Cell level:** Is growth arrest at lag similar to stationary phase arrest?
- **Population level:** How can growth arrested and growing cells co-exist?
- **Evolutionary level:** Is lag extension a strategy selected by evolution or the by-product of defective cell-cycle?

Protocol of evolution under cyclic antibiotic exposure

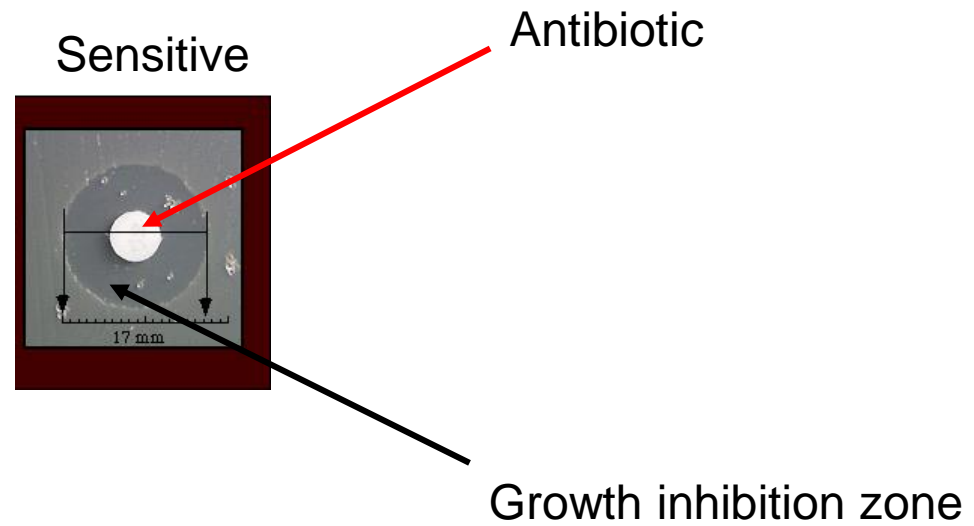


Ofer Fridman

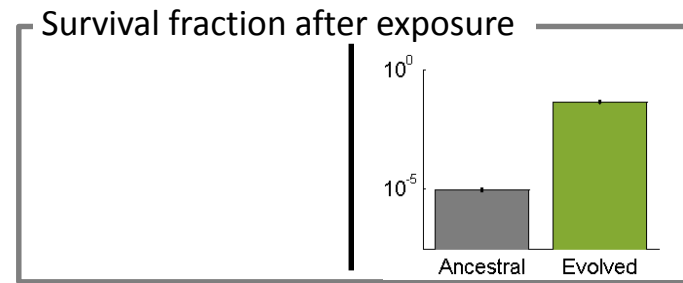
The evolved populations survive extensive antibiotic treatment



Failure of antibiotic treatments is typically attributed to resistance



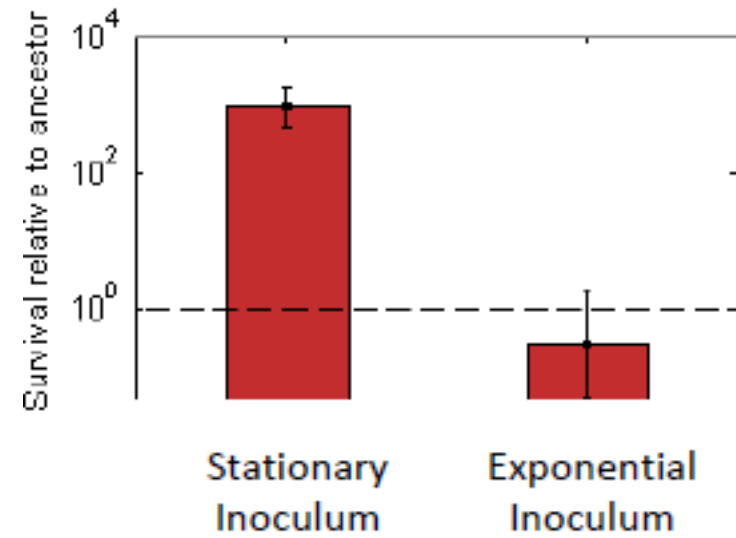
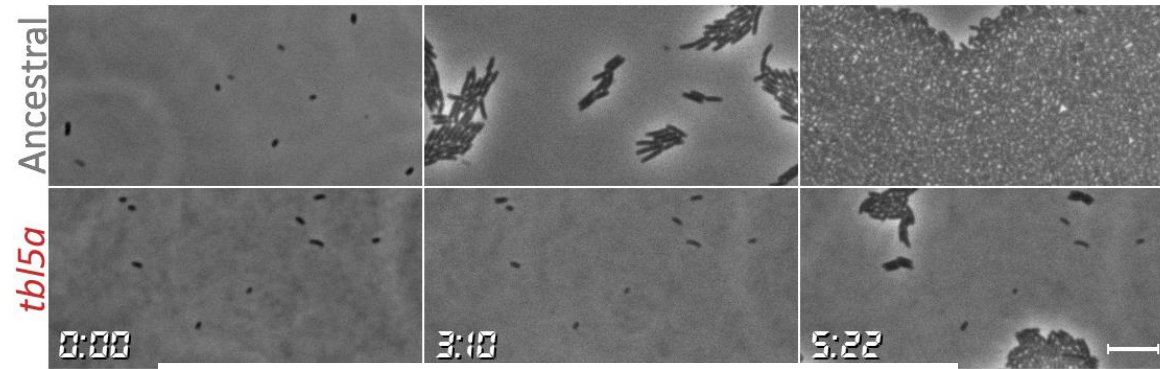
Evolution of tolerance



No difference in sensitivity to the antibiotic

No difference in growth rate

Tolerance is due to an extension of the lag time



Evolution of tolerance

T_a =duration of intermittent antibiotic treatment

$T_a=3h$

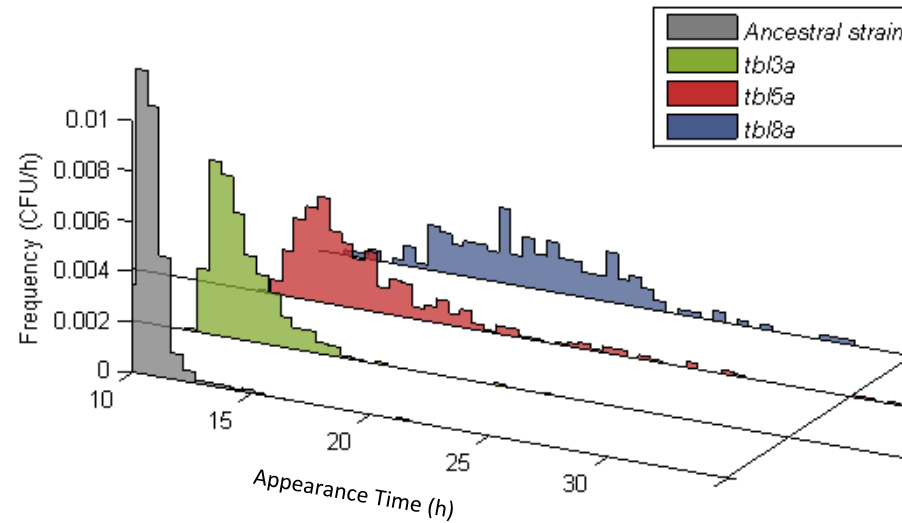
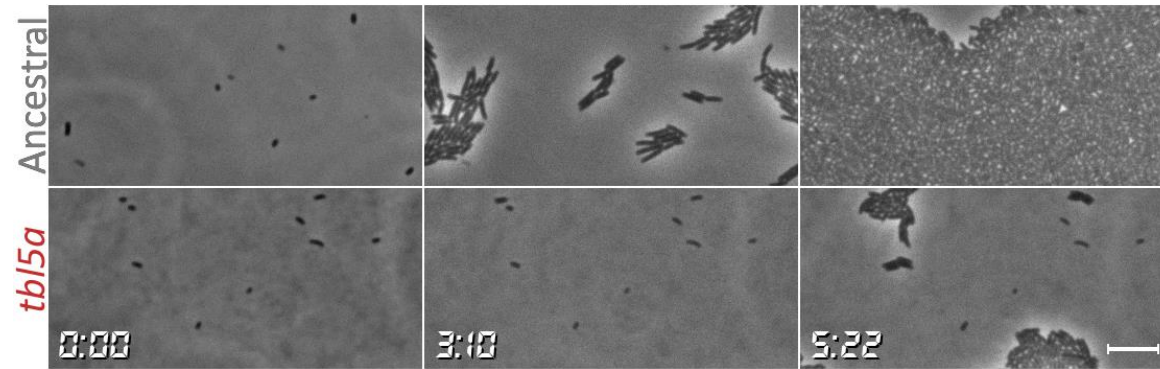
$T_a=5h$

$T_a=8h$

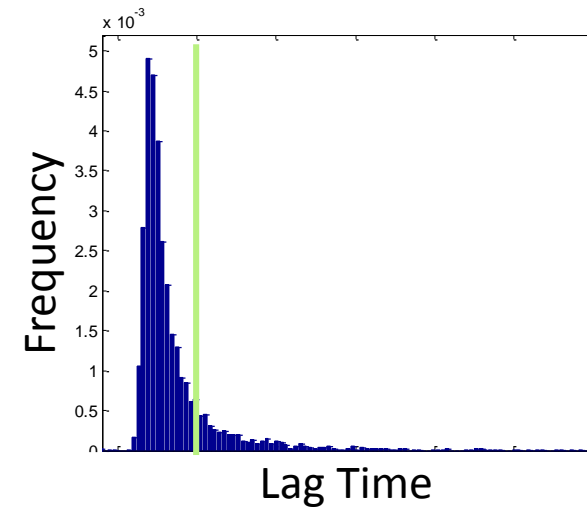
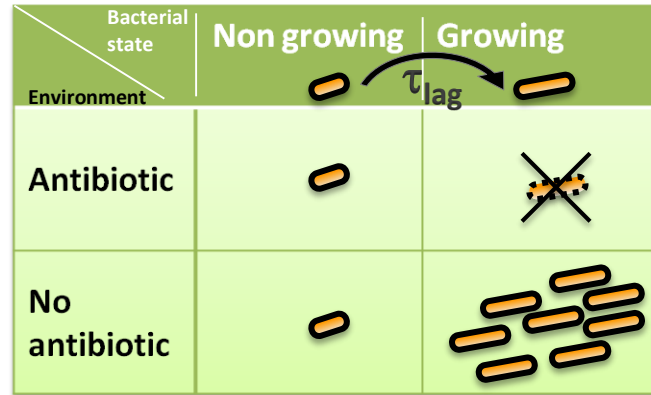
No difference in sensitivity to the antibiotic

No difference in growth rate

Tolerance is due to an extension of the lag time



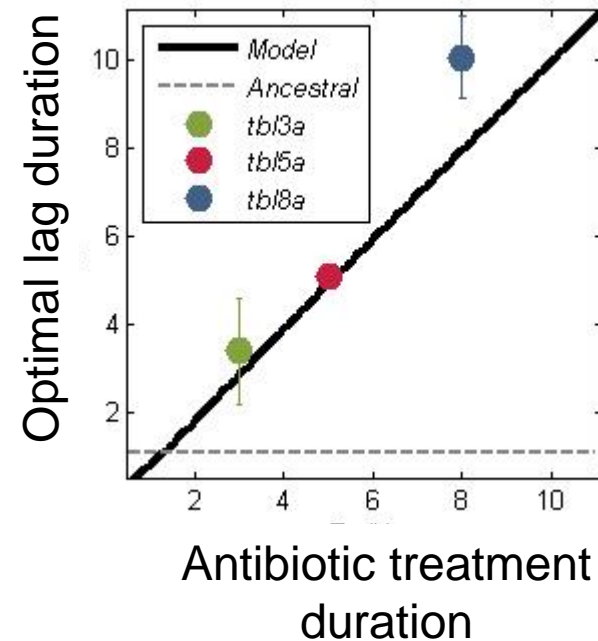
The evolved lag time as an optimization process



$$\dot{L} = -\frac{L}{\tau_{lag}}$$

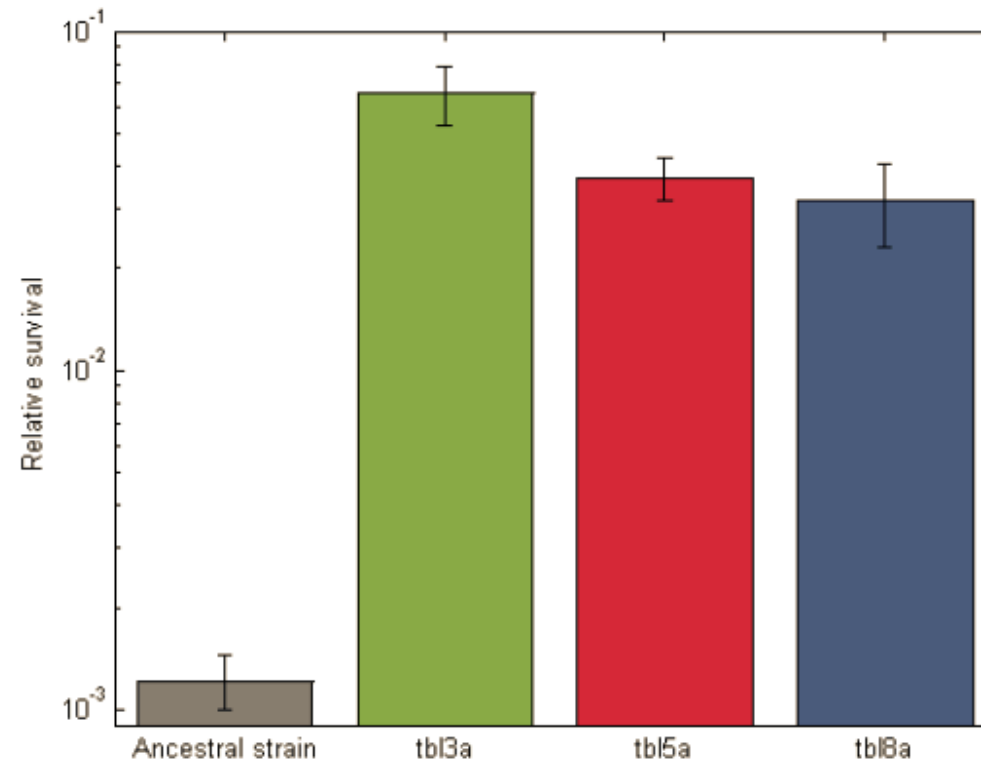
$$\dot{G} = \alpha \frac{G}{\tau_{grow}} + \frac{L}{\tau_{lag}}$$

The evolved lag time is optimized to the duration of the antibiotic exposure



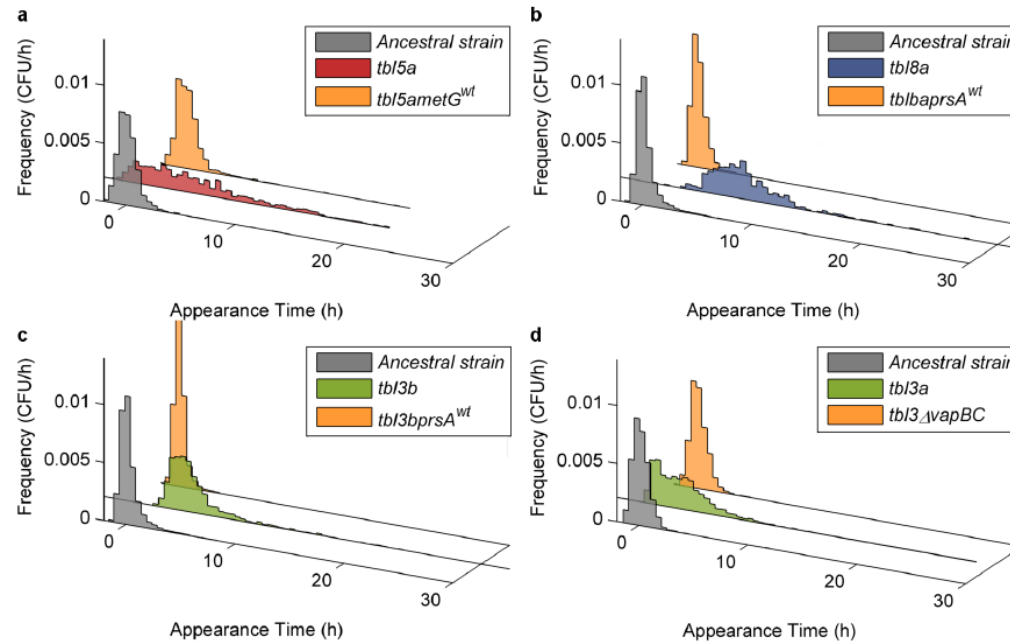
The tolerance-by-lag strains have adapted to the duration of the treatment, not to its precise chemical nature

Strains evolved under cell-wall targeting antibiotic are also tolerant to DNA gyrase inhibitors

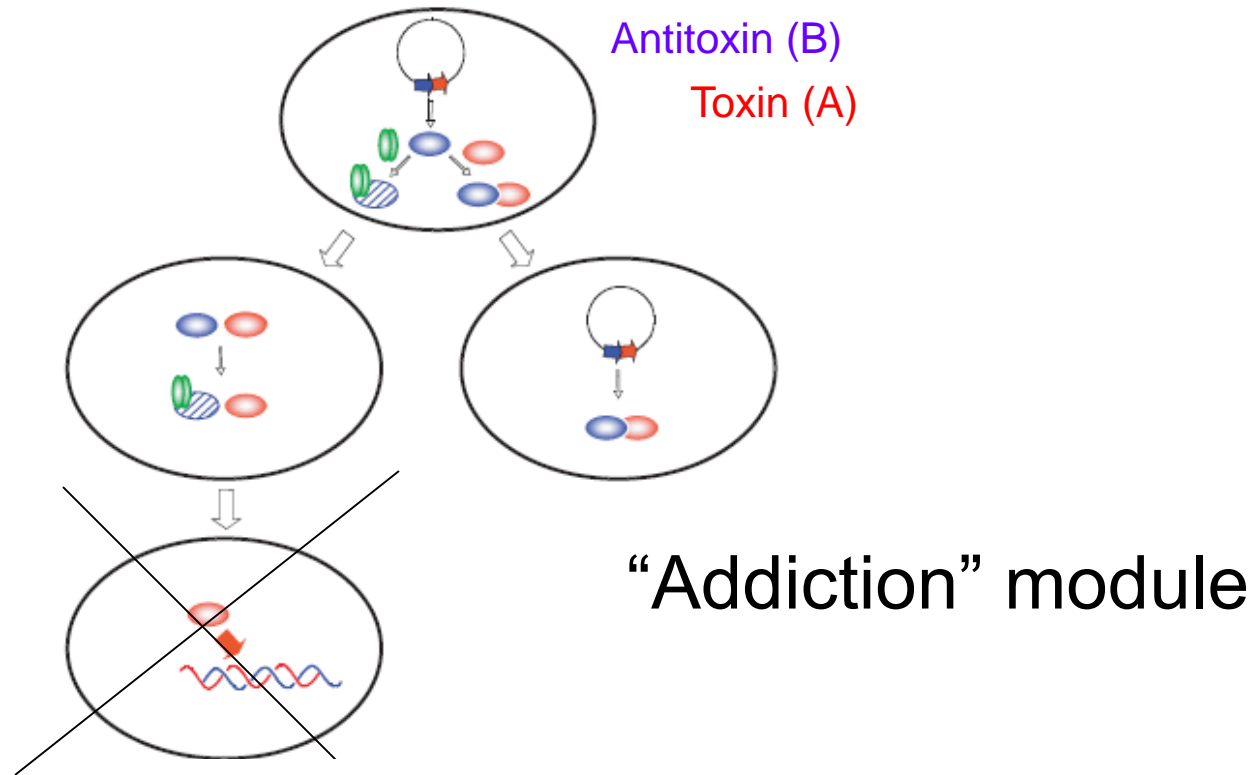


Tolerance-by-lag genes (*tbl*)

- Toxin-antitoxin module (*vapBC*-like)
- Methionyl-tRNA synthetase (*metG*)
- Ribose-phosphate diphosphokinase (*prsA*)



Toxin-Antitoxin Modules on Plasmids



Toxin-Antitoxin Chromosomal Modules (Aizenman E et al. PNAS 1996)

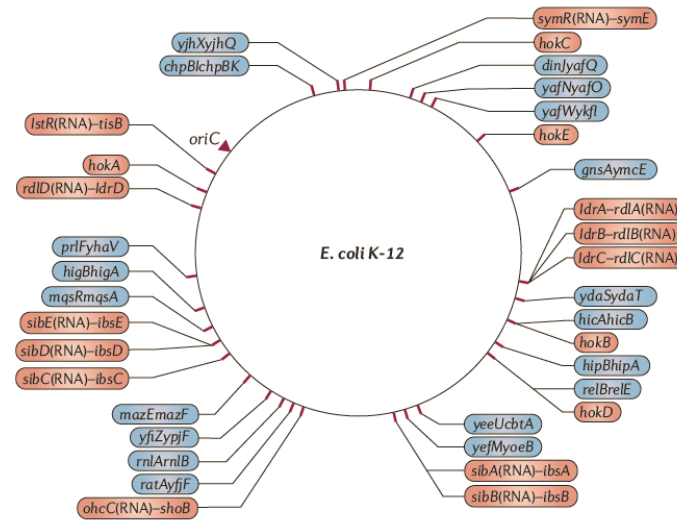
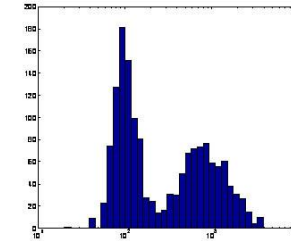
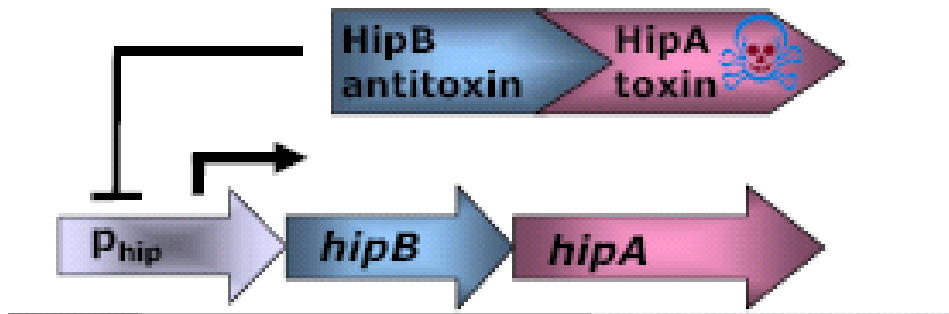


Figure 2 | Genomic map of the toxin-antitoxin modules in *Escherichia coli*.

Yoshihiro Yamaguchi and Masayori Inouye

Nature Reviews Microbiology, 2011

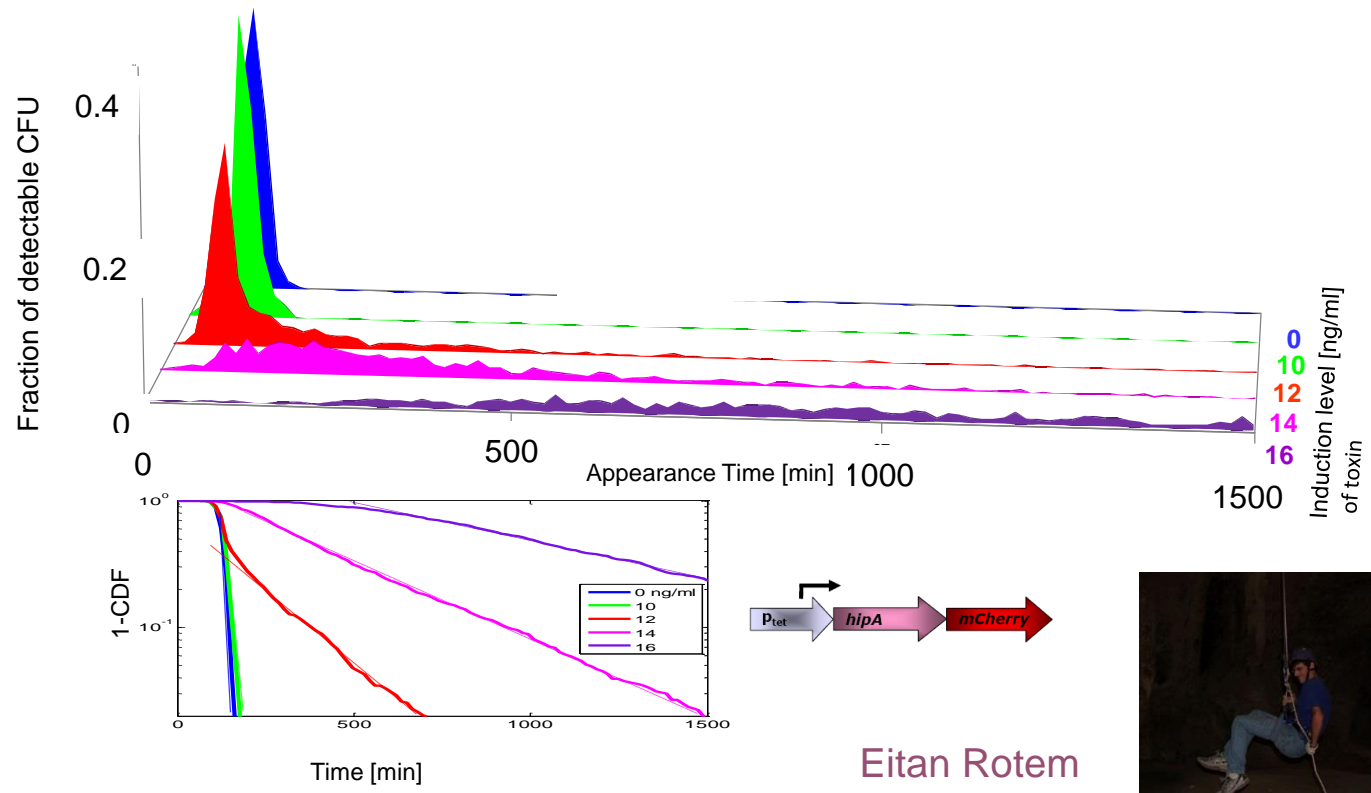
Toxin-antitoxin modules control of lag time duration



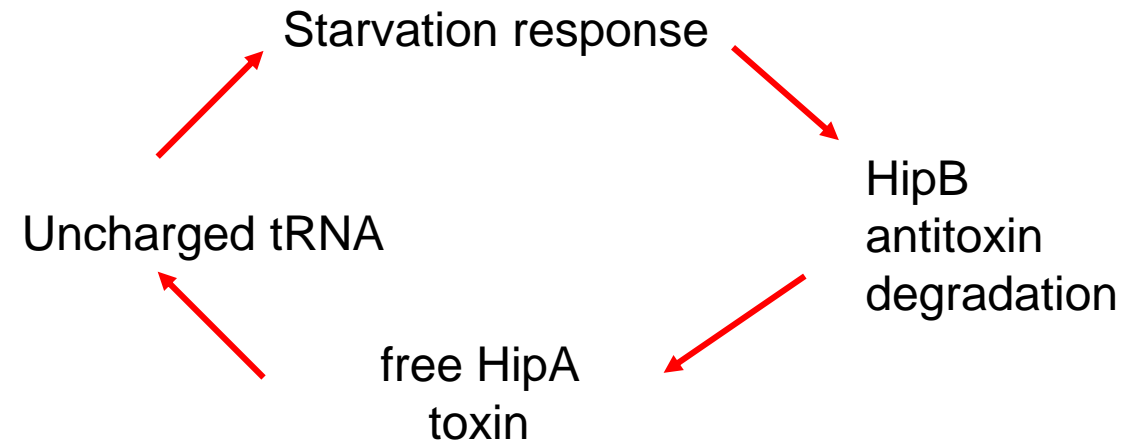
- Mutations in *hipA* lead to high persistence (Harris Moyed *J Bac*, 1983, 1988, 1991, 1994; Korch *et al Mol Mic* 2003)

The *hip* (*high persistence* toxin-antitoxin module)

The level of induction of the toxin determines the growth arrest duration



HipA toxin induces a long lag by phosphorylating GltX, the glutamyl tRNA synthetase

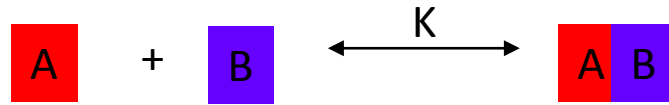


Kaspy I., *et al*, (*Nature Communication*, 2013)
Collaboration with Gad Glaser

The HipA toxin extends the lag by mimicking starvation



Threshold amplification of noise



A=toxin B=antitoxin

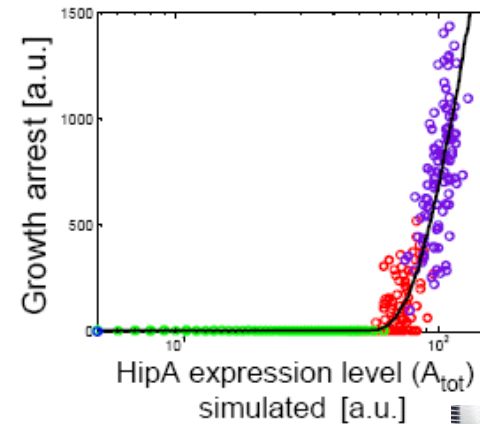
$$\frac{dA}{dt} = \alpha - k_+ A \cdot B + k_- AB - \mu A$$

$$\frac{dB}{dt} = \beta - k_+ A \cdot B + k_- AB - \mu^* B$$

$$\frac{dAB}{dt} = k_+ A \cdot B - k_- AB - \mu AB$$

(Lenz DL et al., 2004; Levine E. et al., 2007; Buchler NE, 2008)

Stochastic simulations of growth arrest

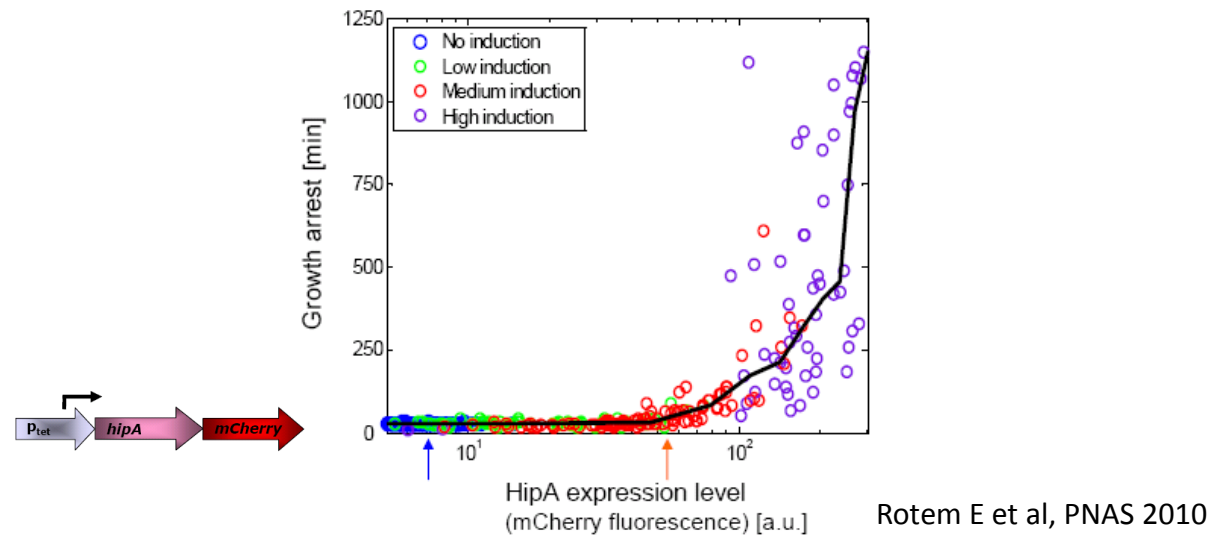
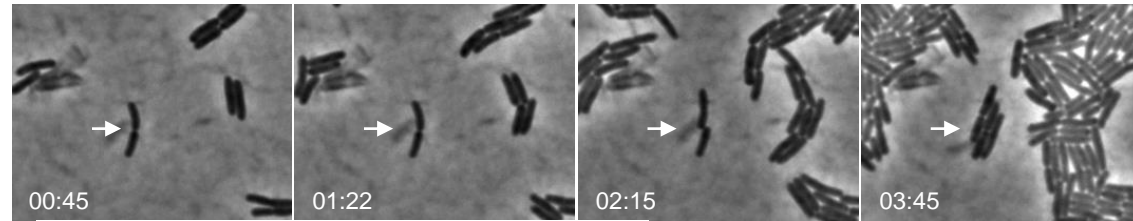


Adiel Loinger

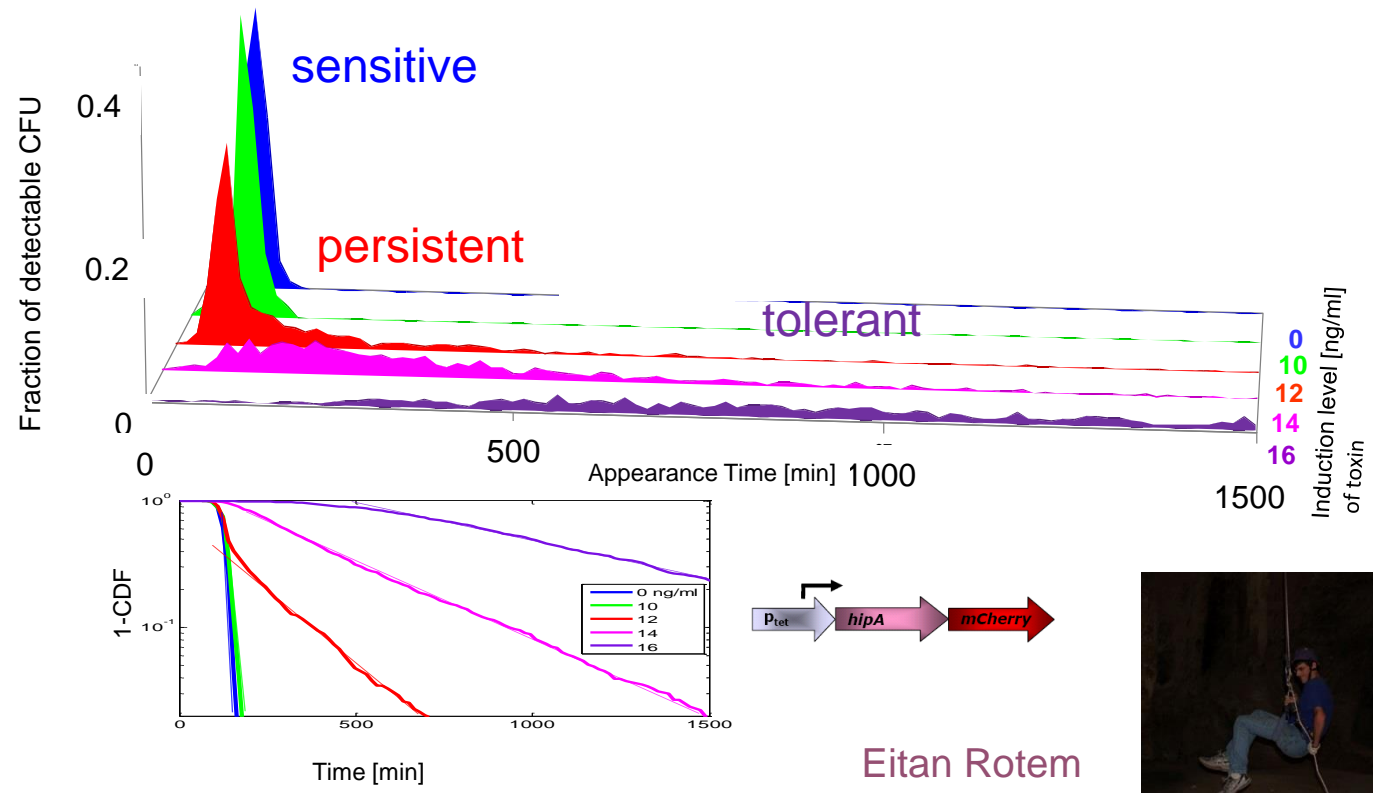
In collaboration with O. Biham



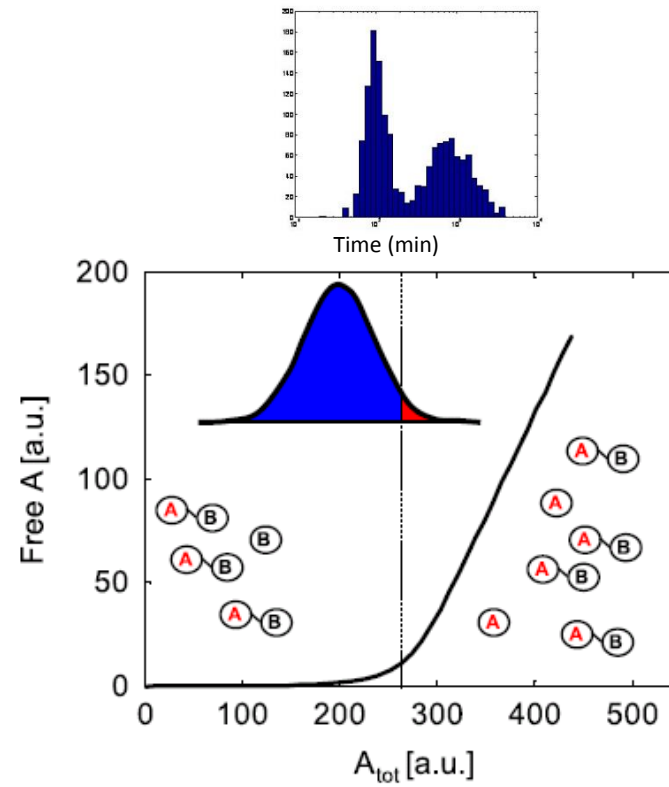
From molecular noise to phenotypic variability of growth at the single-cell level



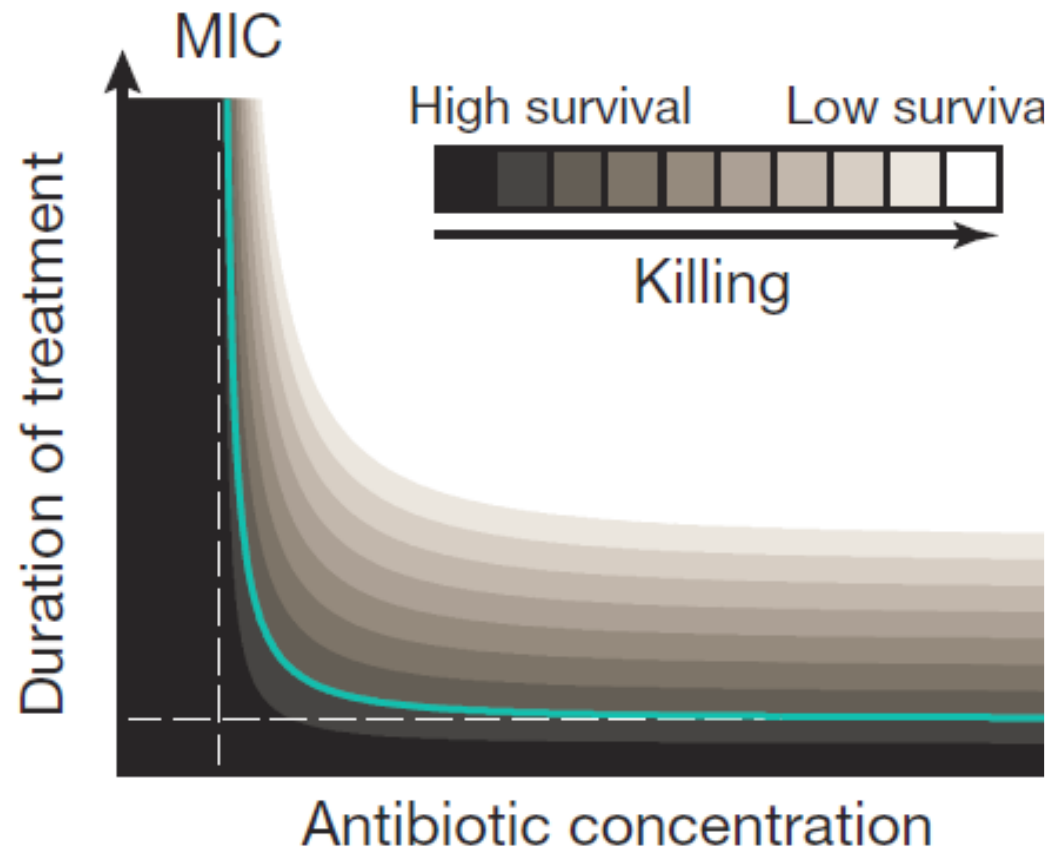
The level of induction of the toxin determines the growth arrest duration



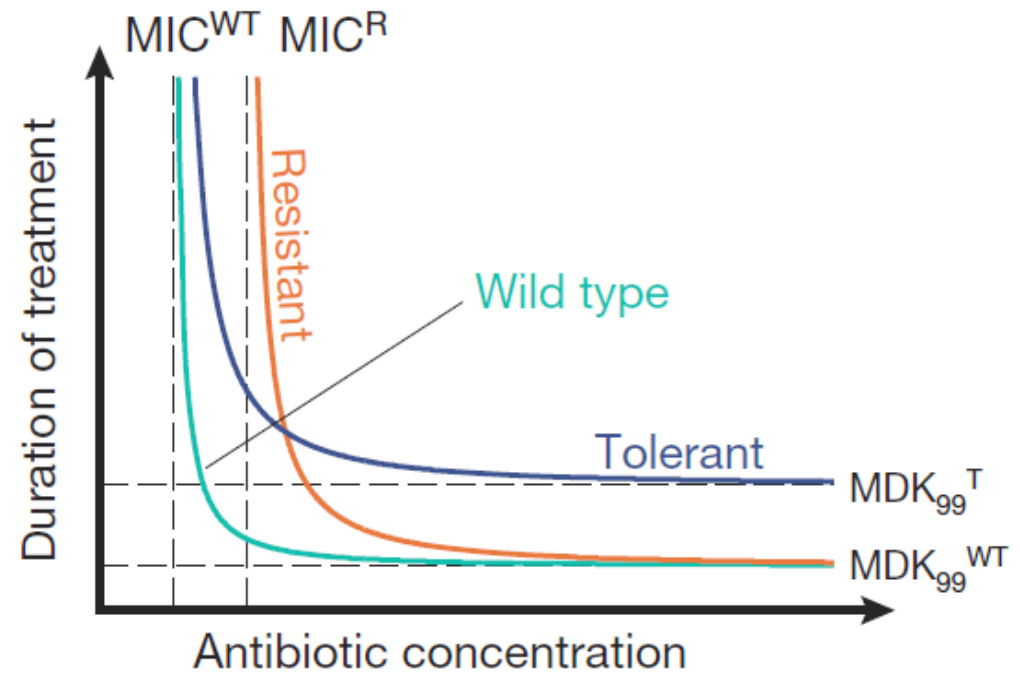
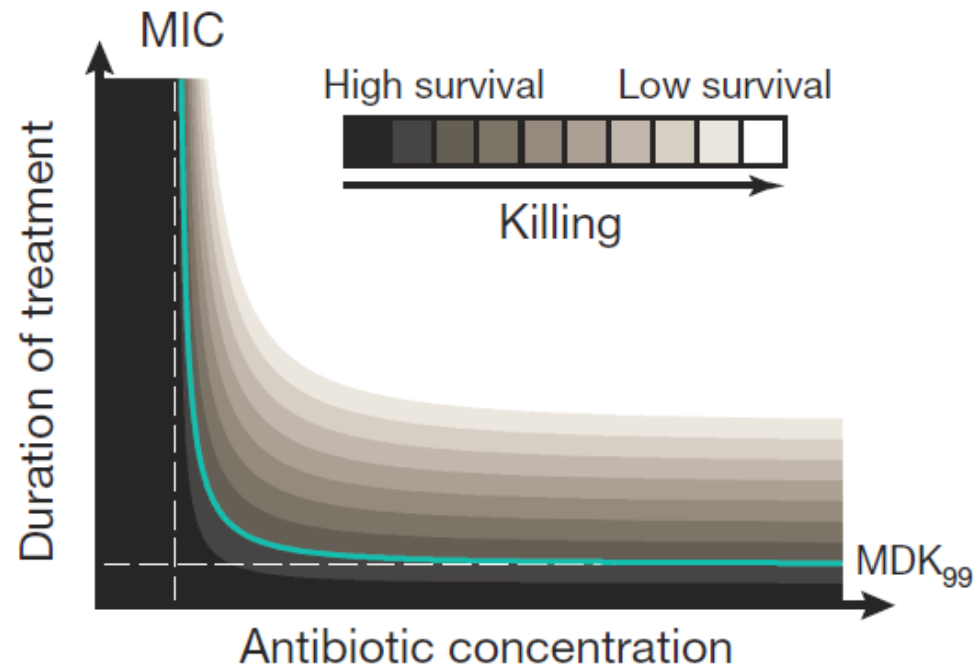
Difference between large numbers leads to amplification of noise



MIC and MDK: a global framework to define resistance, tolerance and persistence

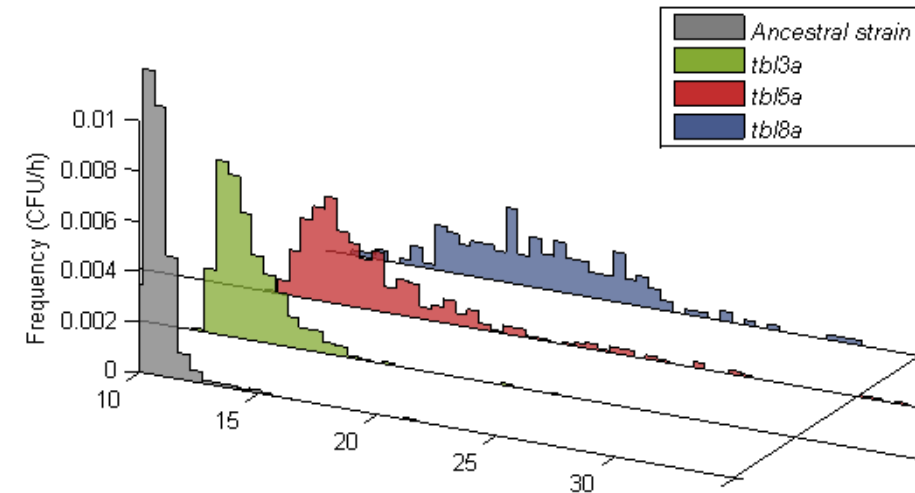
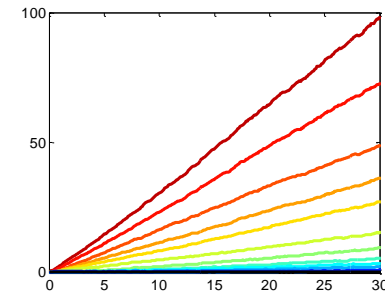


MIC and MDK



Summary

- **CASP: Constant Activity Stationary Phase** enables quantitative measurements on non growing cells
- Toxin-antitoxin modules are ideally suited to control tolerance and persistence by mimicking starvation
- Growth arrest can evolve to match the time scale of the stress
 - Understand *tbl* mutations
 - Search for similar effects in patients treated with cyclic exposure to antibiotics



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Broad Institute

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