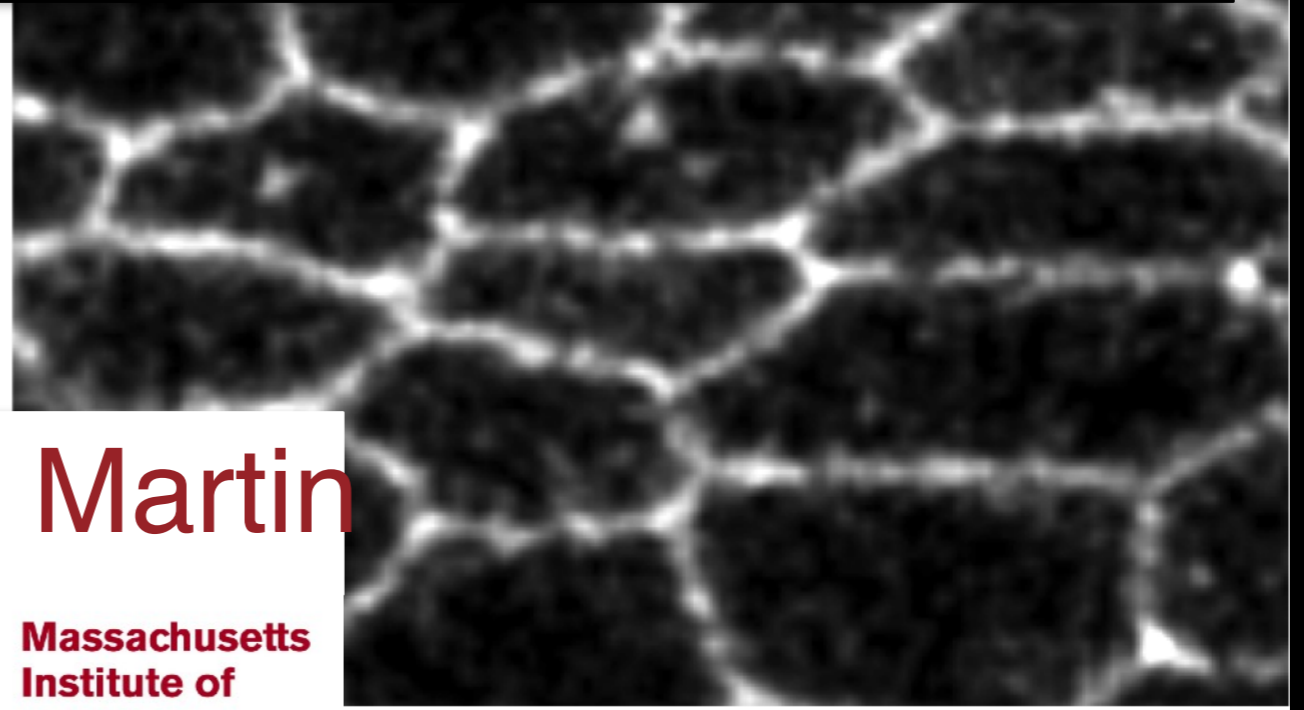
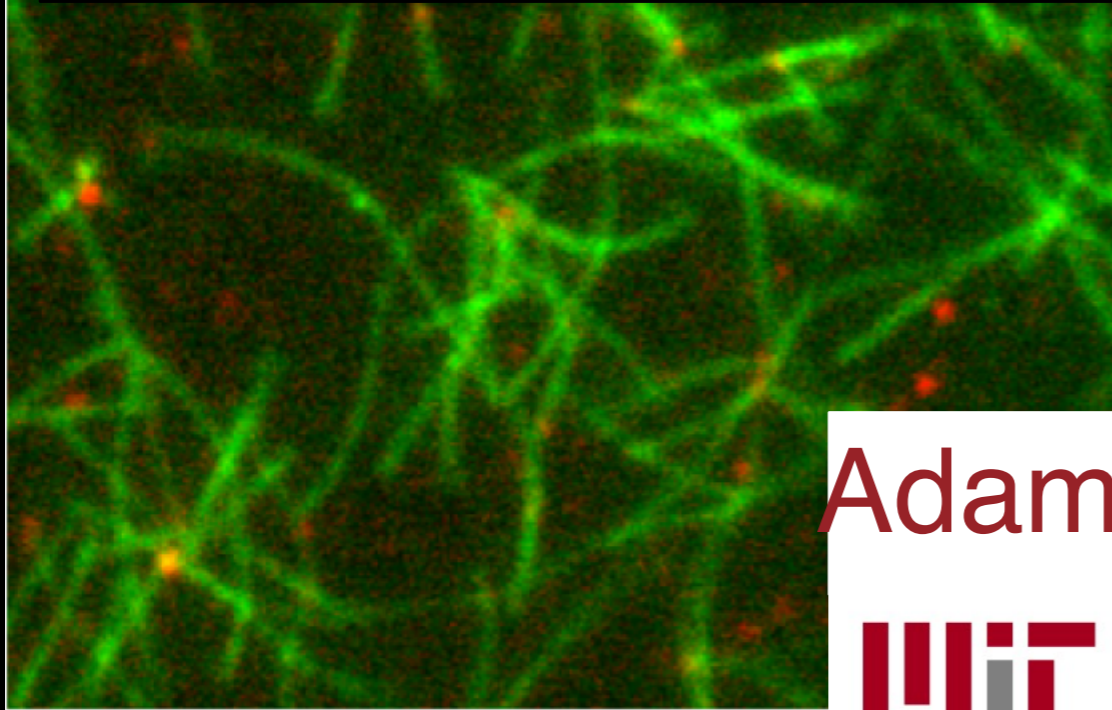


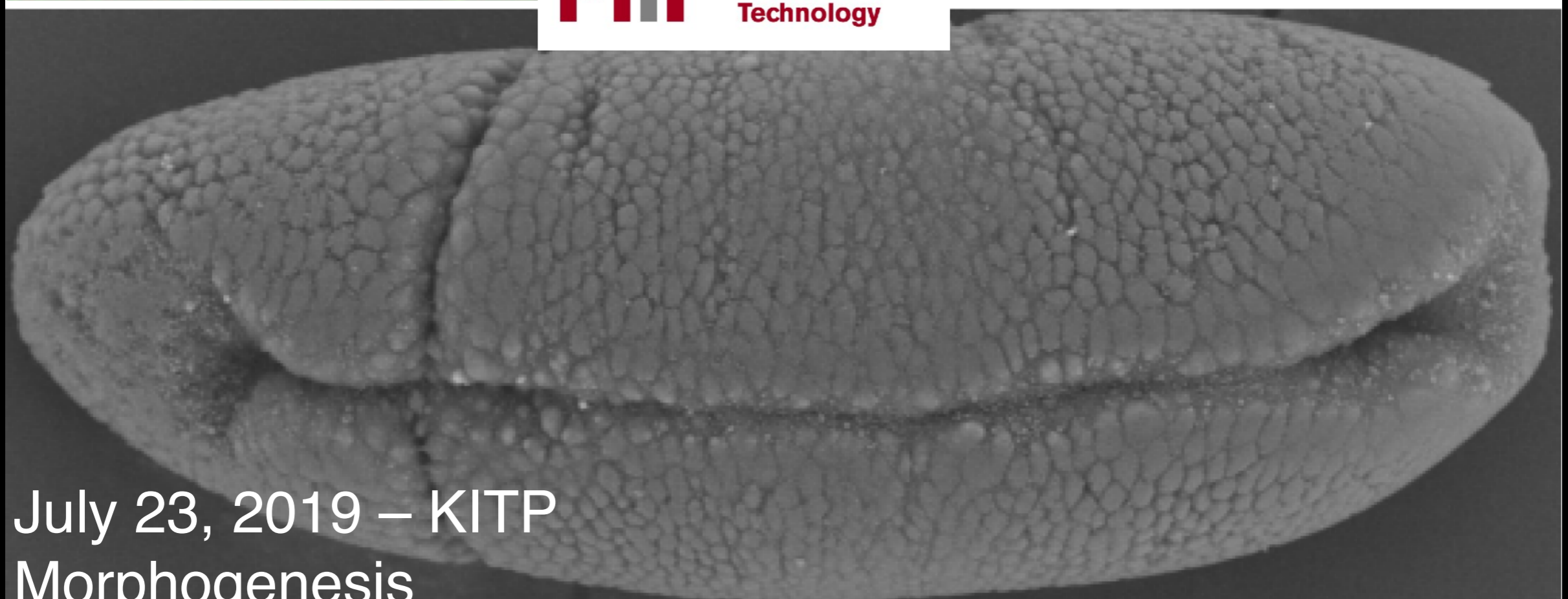
The early *Drosophila* embryo: a little something for everyone



Adam Martin

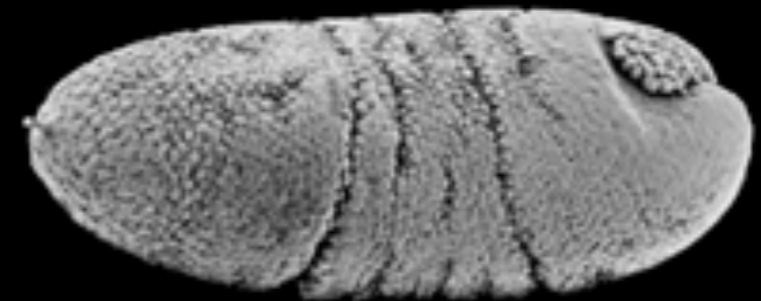
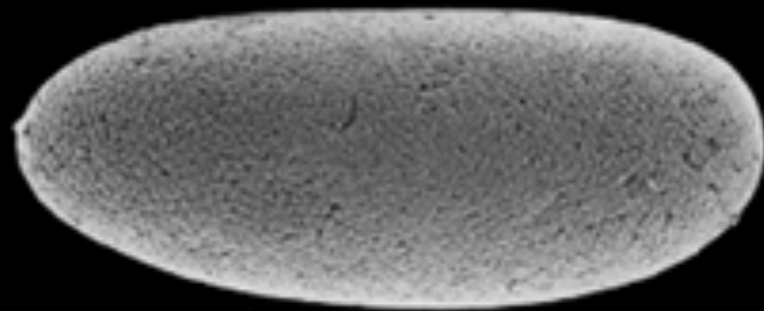


Massachusetts
Institute of
Technology



July 23, 2019 – KITP
Morphogenesis

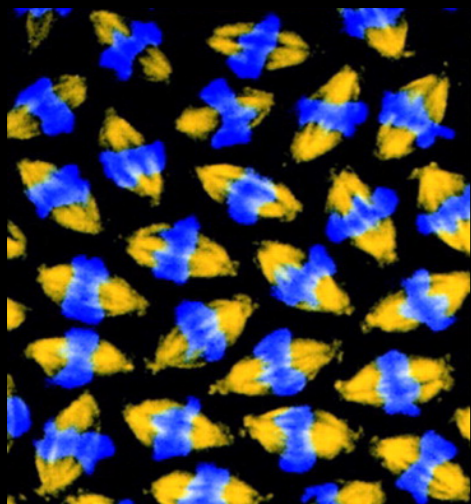
Drosophila embryo: Everything a Biophysicist could want (in a couple hrs)



Time (4 hours)



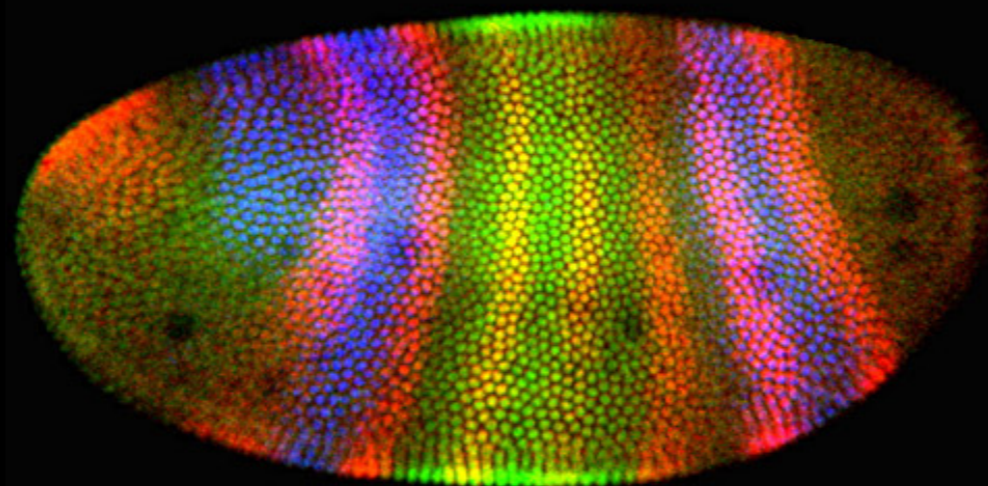
1) Nuclear division



Microtubules

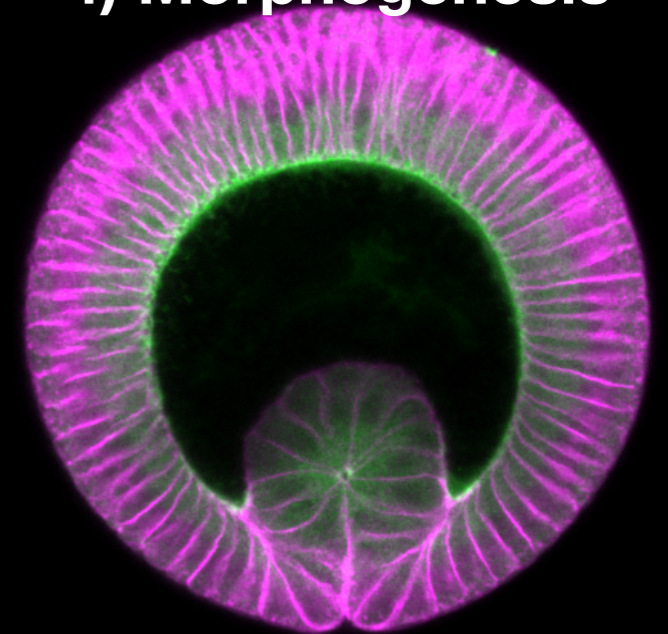
DNA

2) Patterned gene expression
3) Cellularization



Different transcription factors

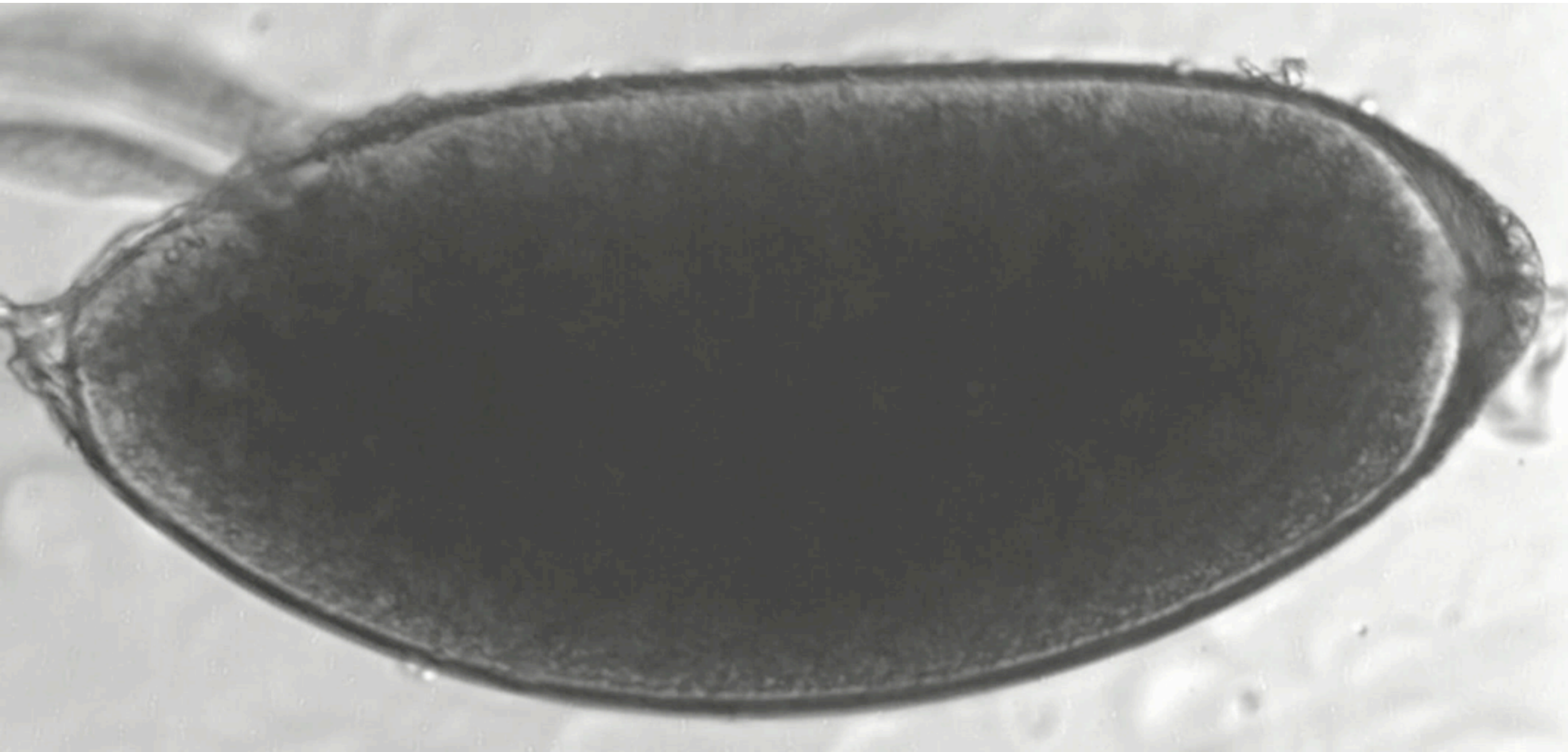
4) Morphogenesis



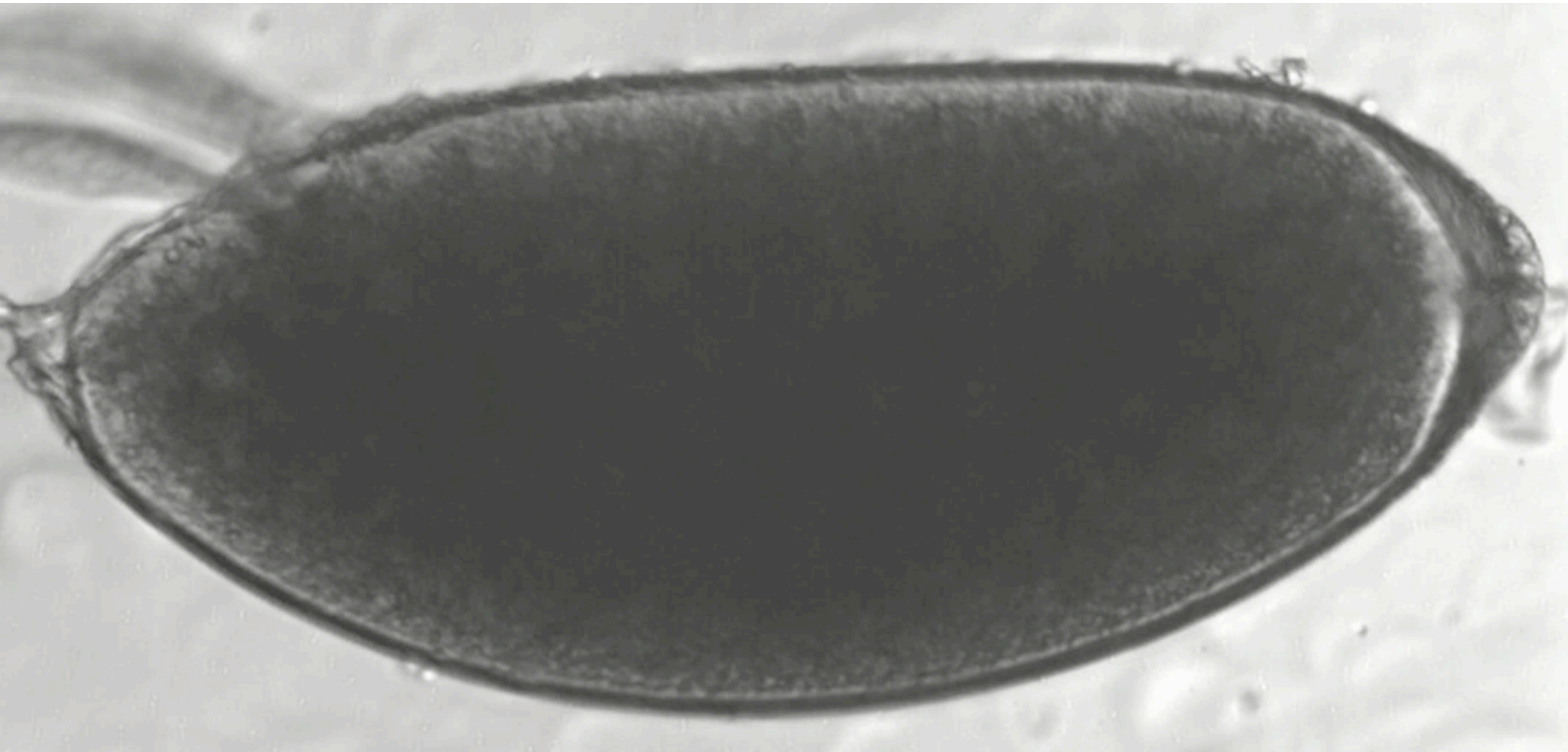
Plasma Membrane

Myosin motor

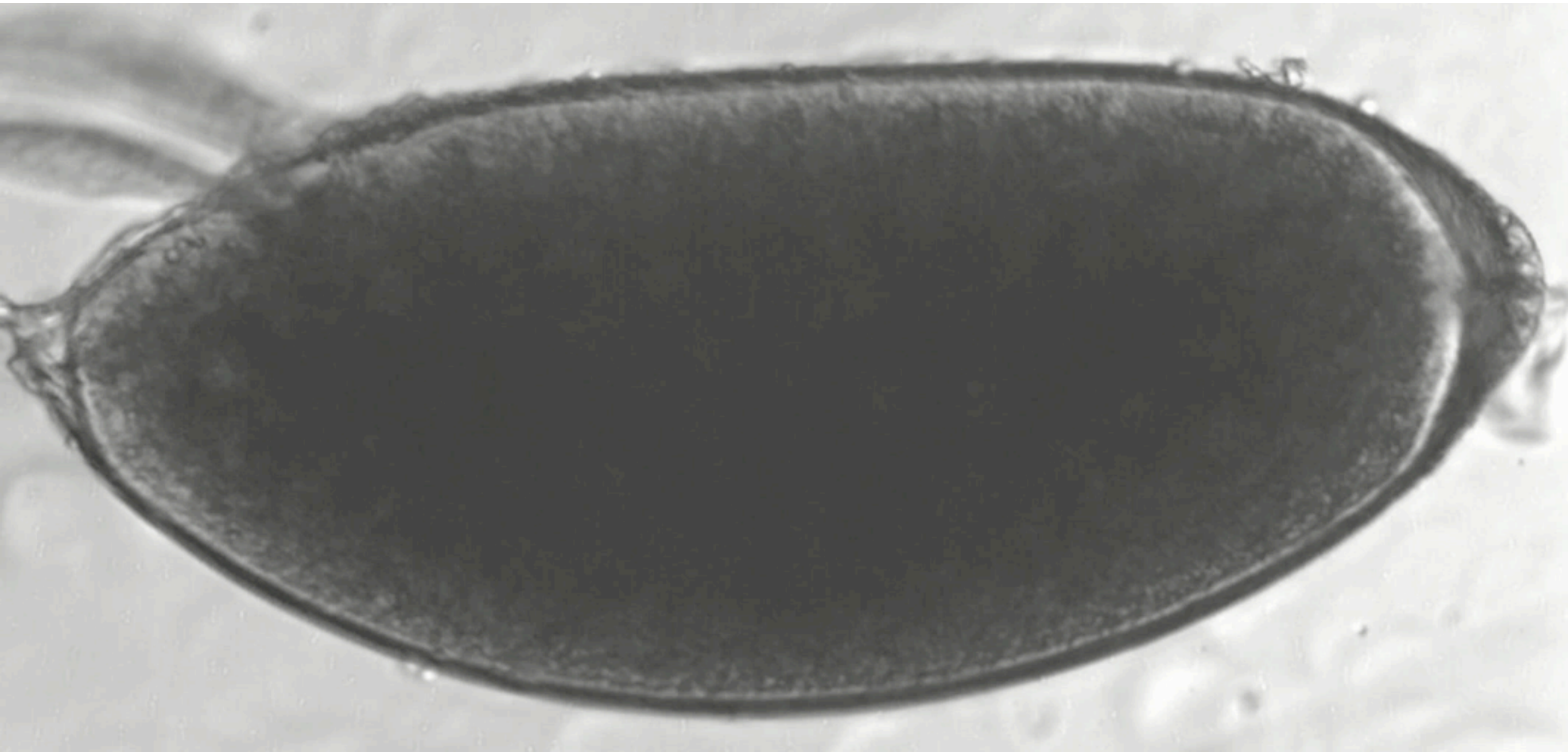
Development involves global tissue movements



Development involves global tissue movements



Development involves global tissue movements



Cell cycle transitions during early development

nuclear replication
(cell cycle ~nine minutes)

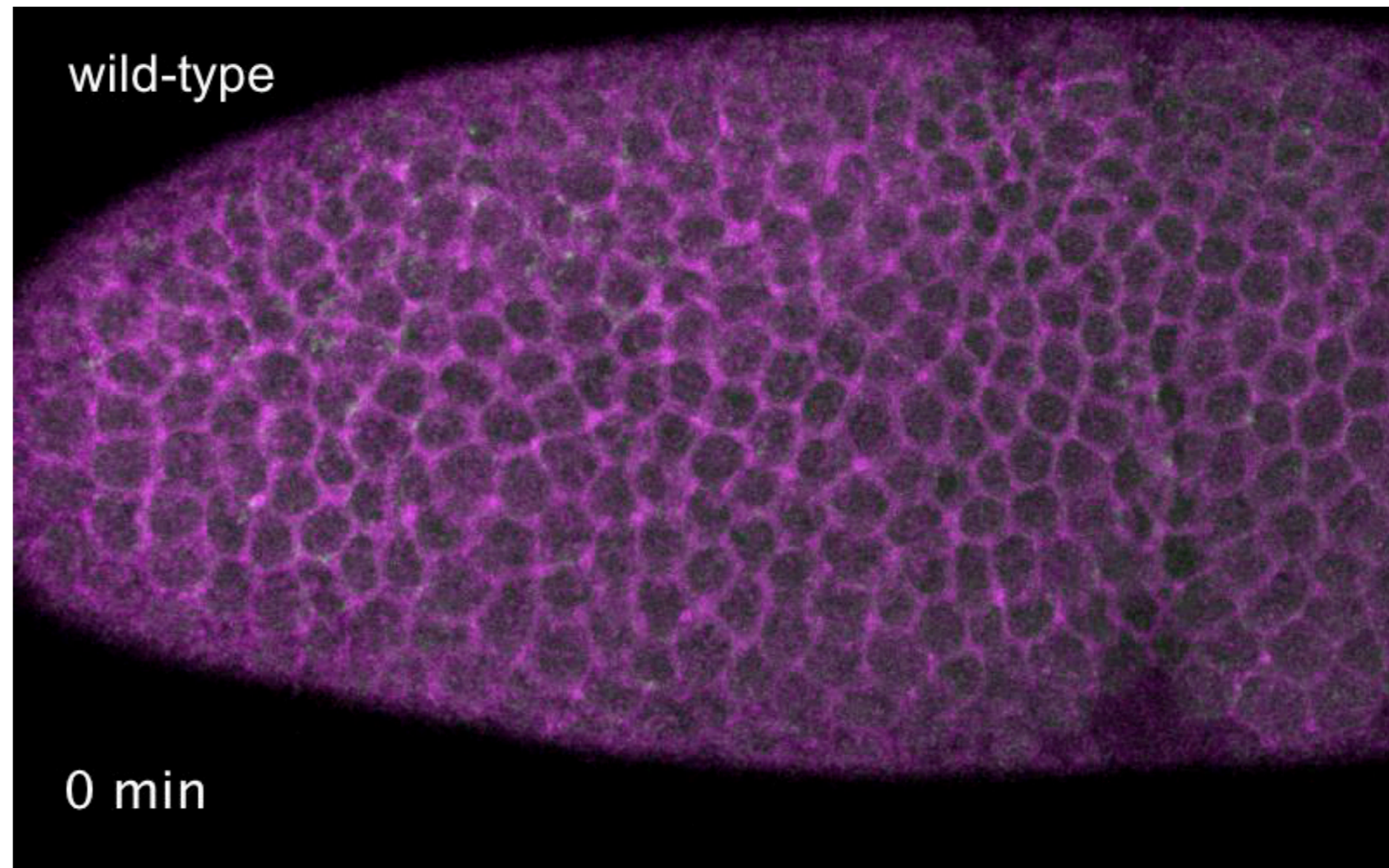
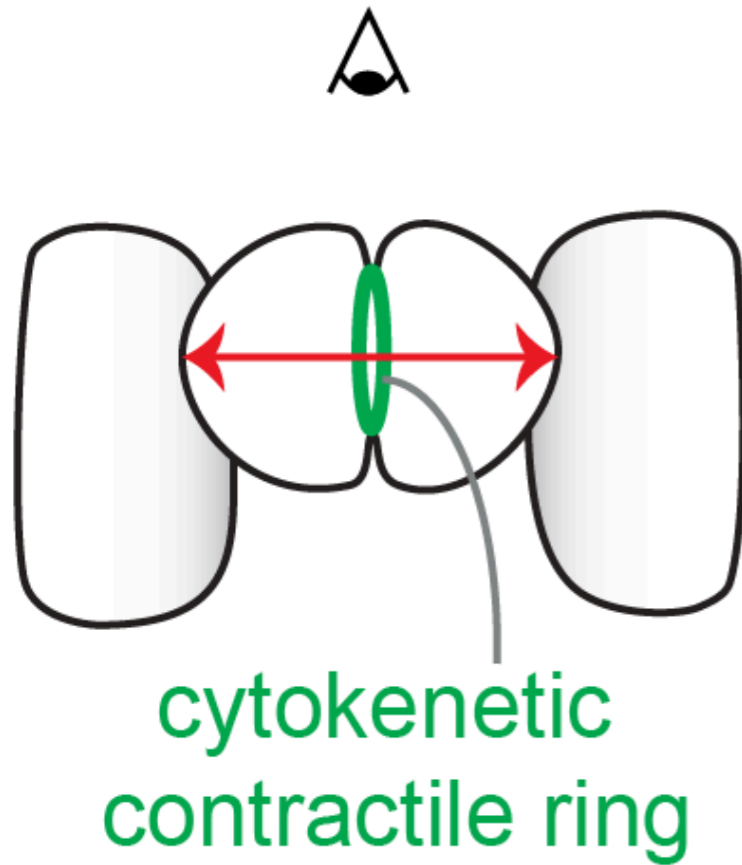
Cell cycle transitions during early development

nuclear replication
(cell cycle ~nine minutes)

Cell cycle transitions during early development

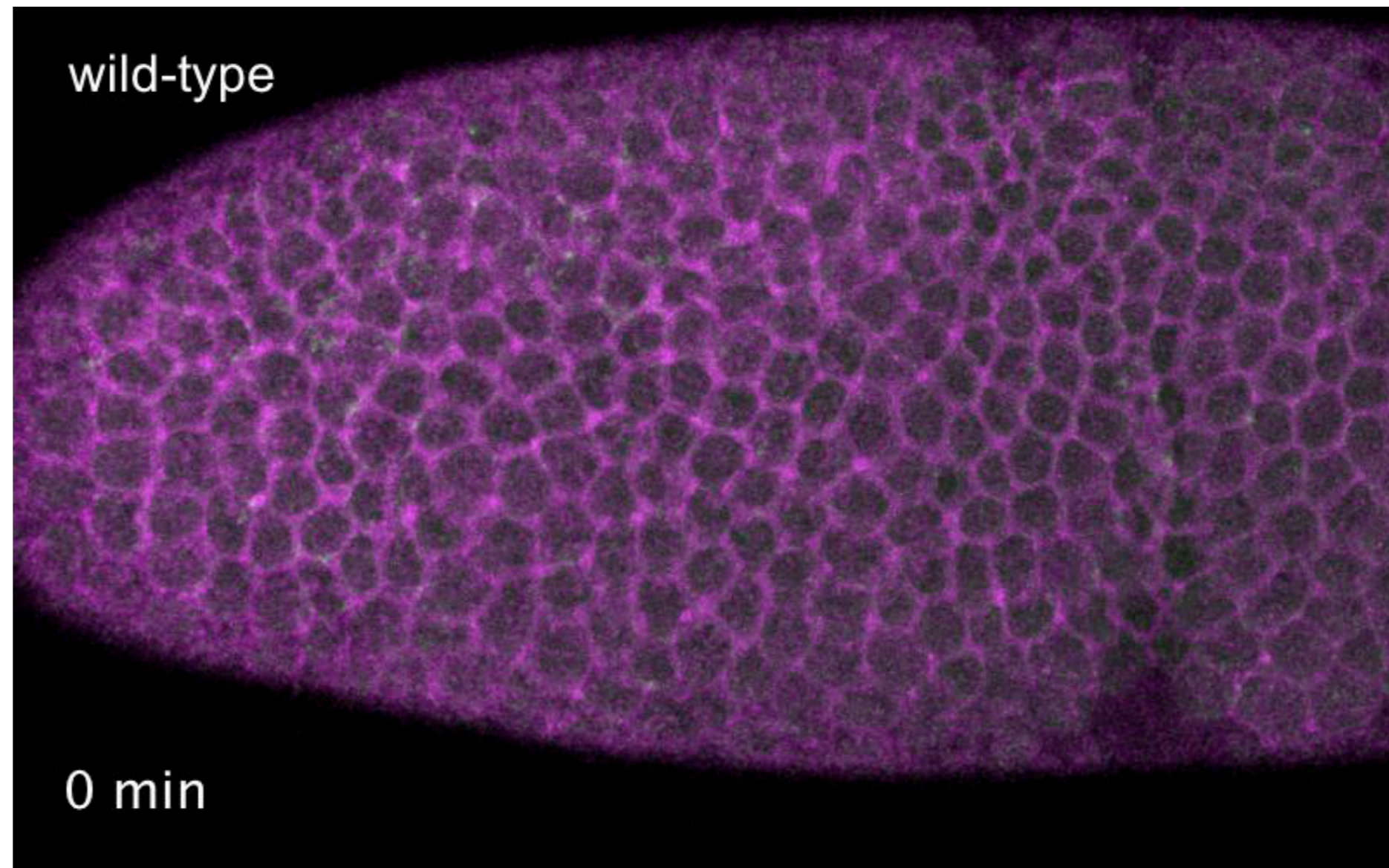
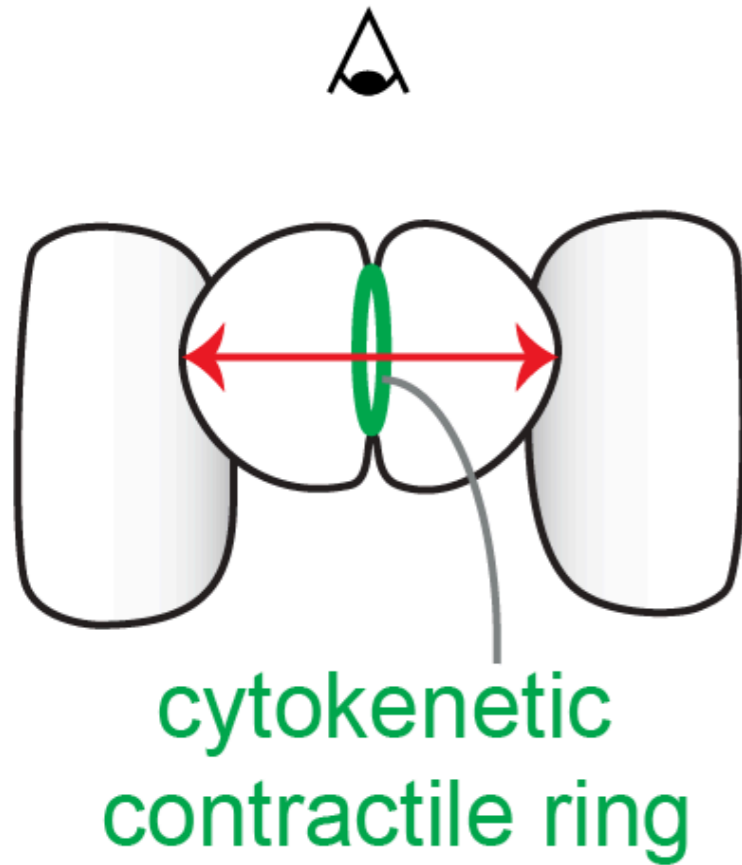
nuclear replication
(cell cycle ~nine minutes)

After cells form, cell division occurs in mitotic domains



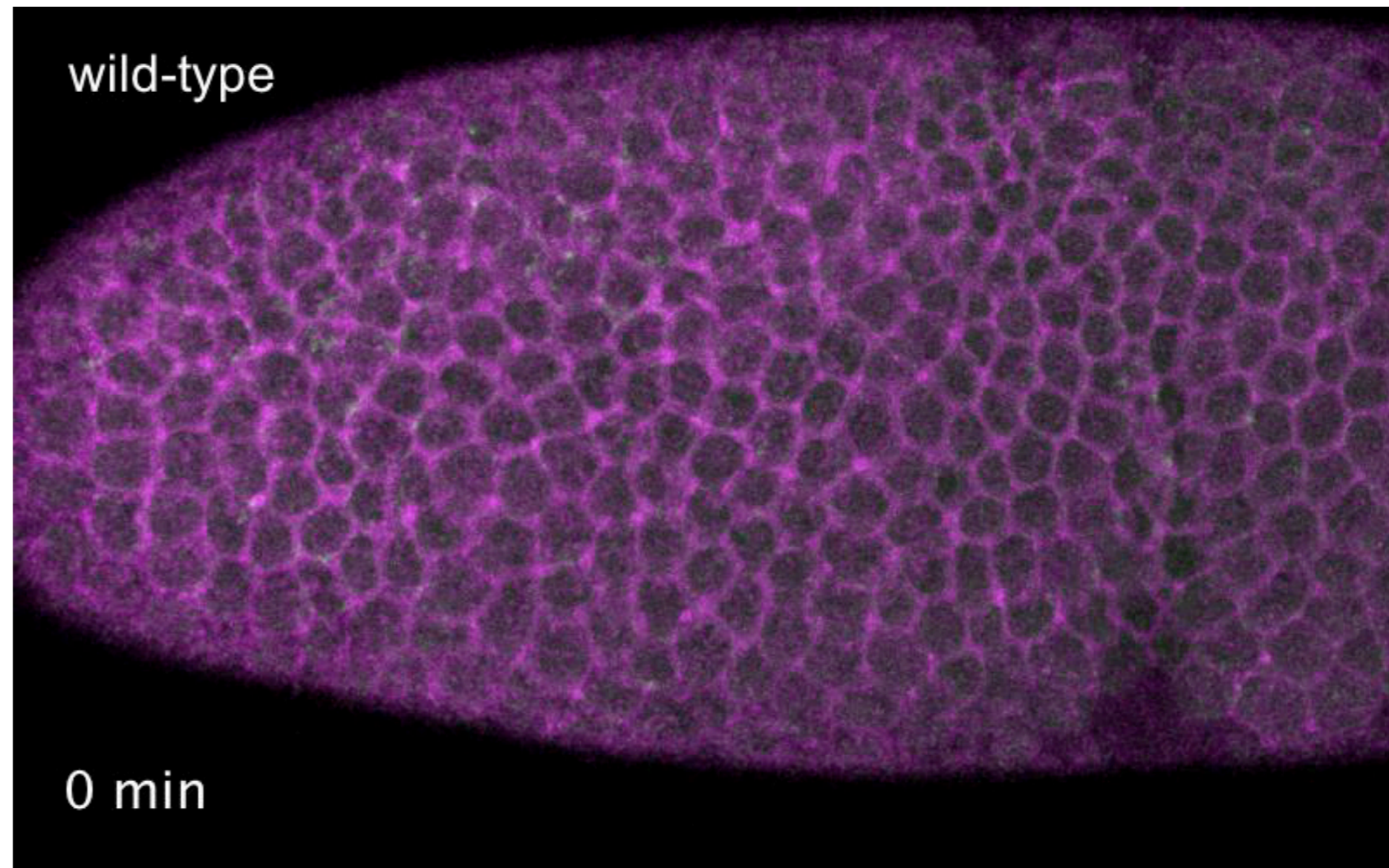
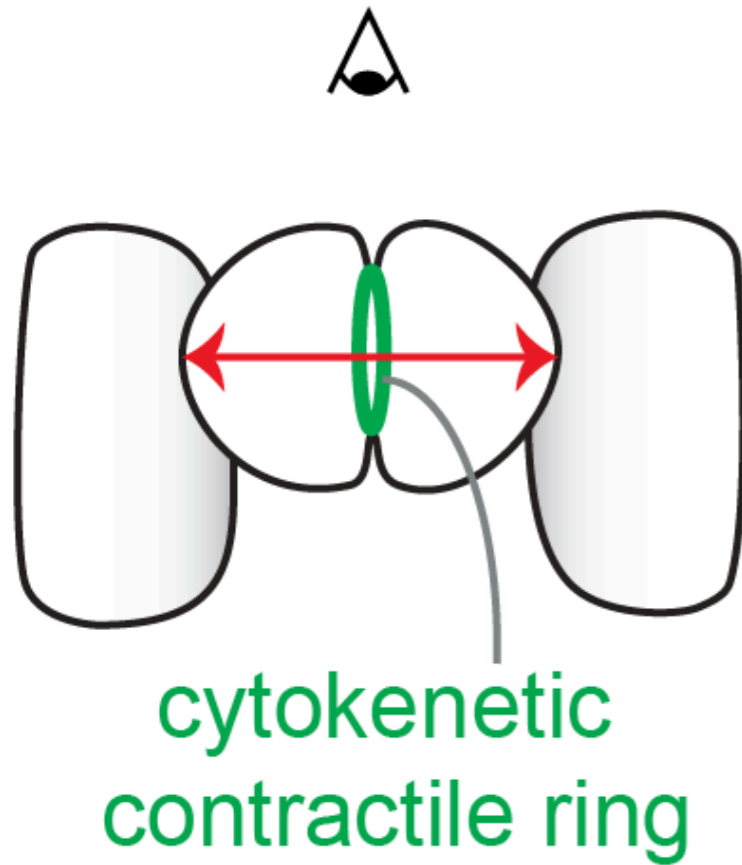
Chanet et al., 2017

After cells form, cell division occurs in mitotic domains



Chanet et al., 2017

After cells form, cell division occurs in mitotic domains



Chanet et al., 2017

Making heads or tails of development

Embryo



Making heads or tails of development

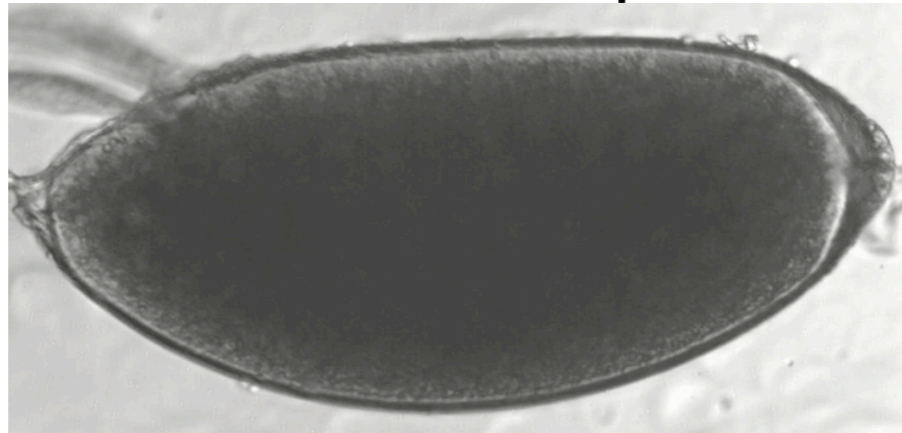
Embryo



Making heads or tails of development

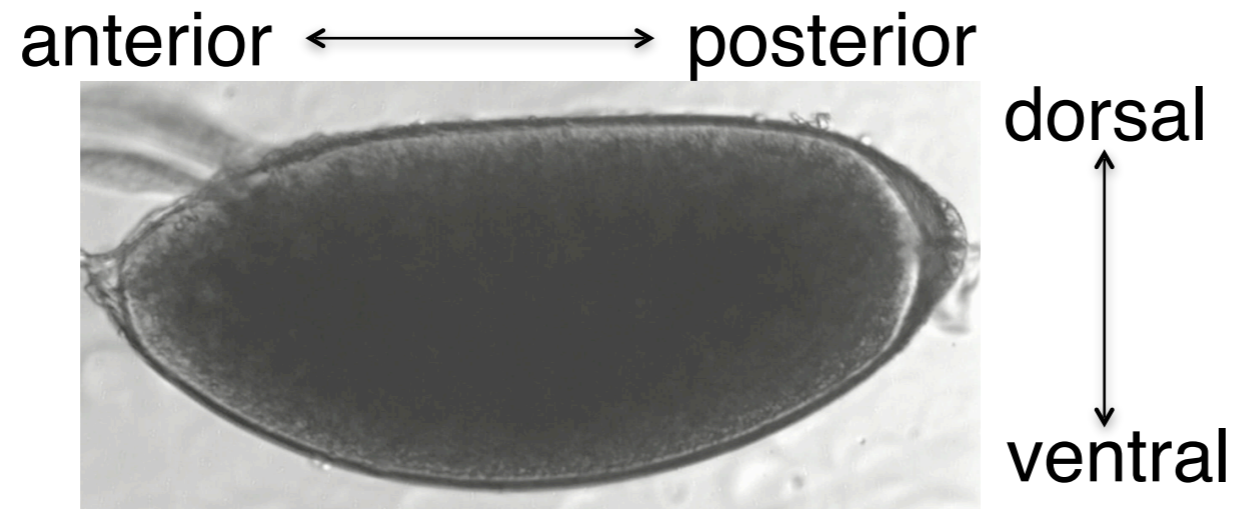
anterior ← → posterior

Embryo



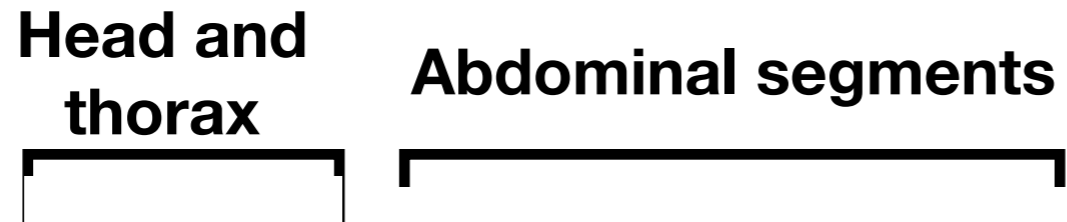
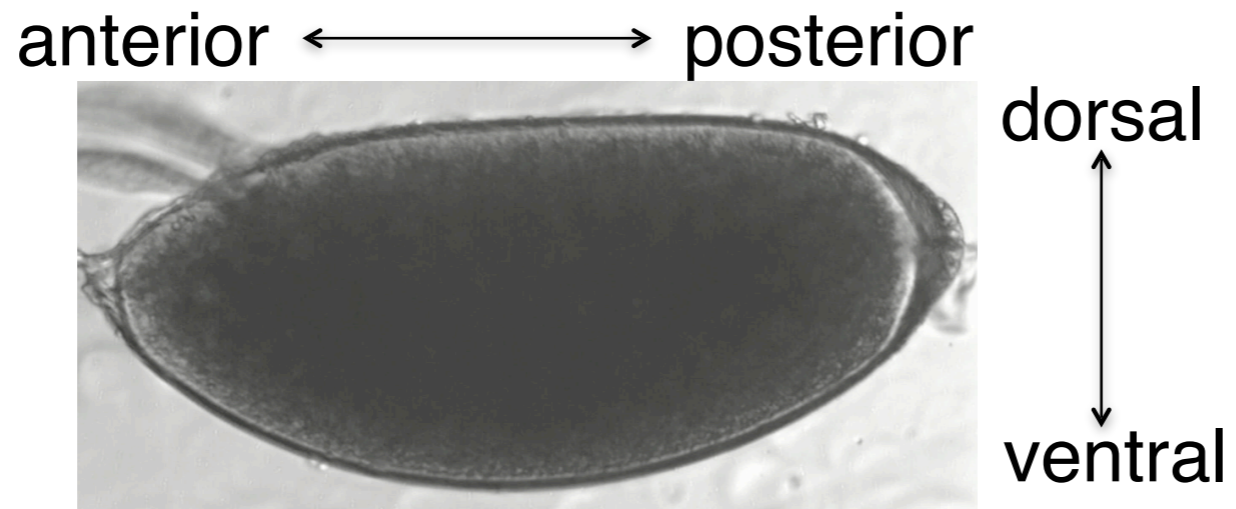
Making heads or tails of development

Embryo



Making heads or tails of development

Embryo

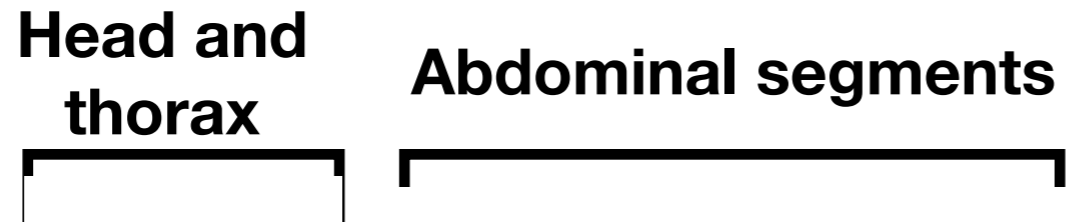
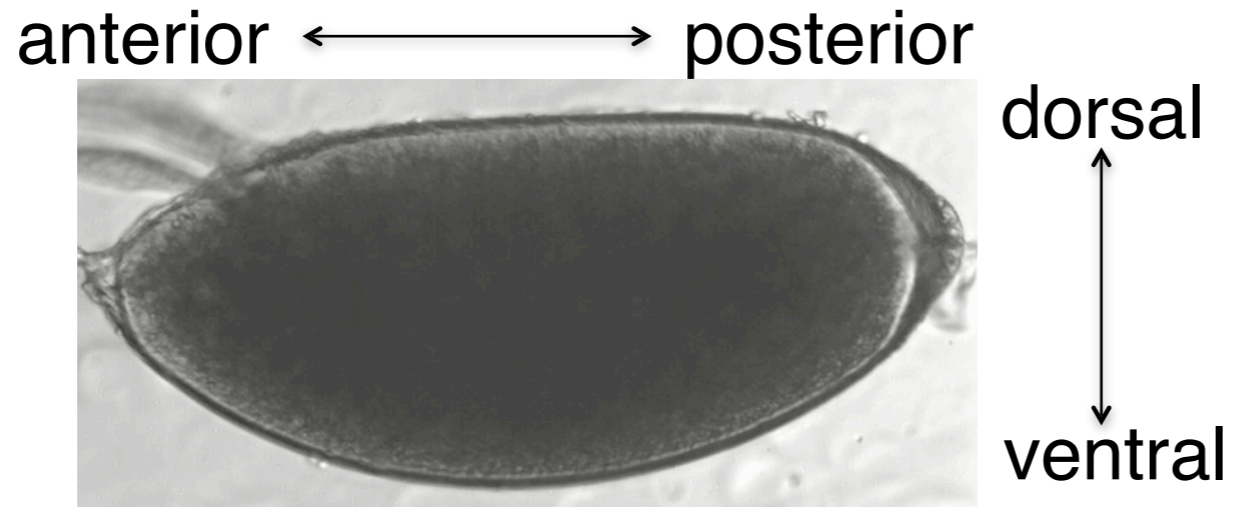


Larva



Making heads or tails of development

Embryo



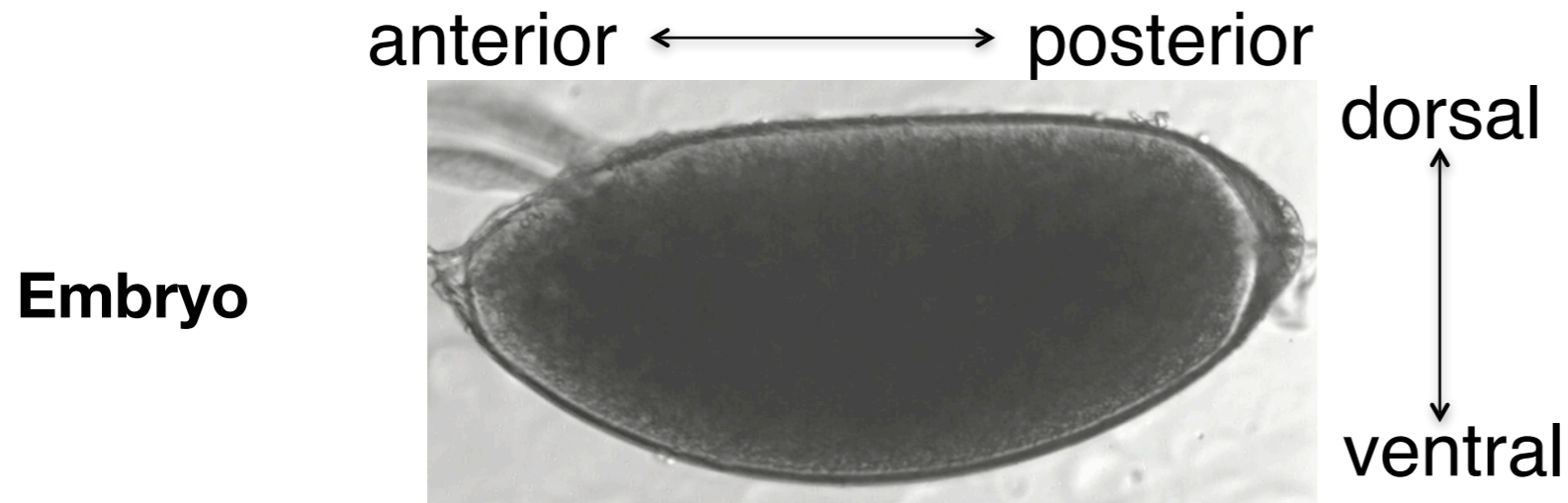
Larva



Adult



Making heads or tails of development

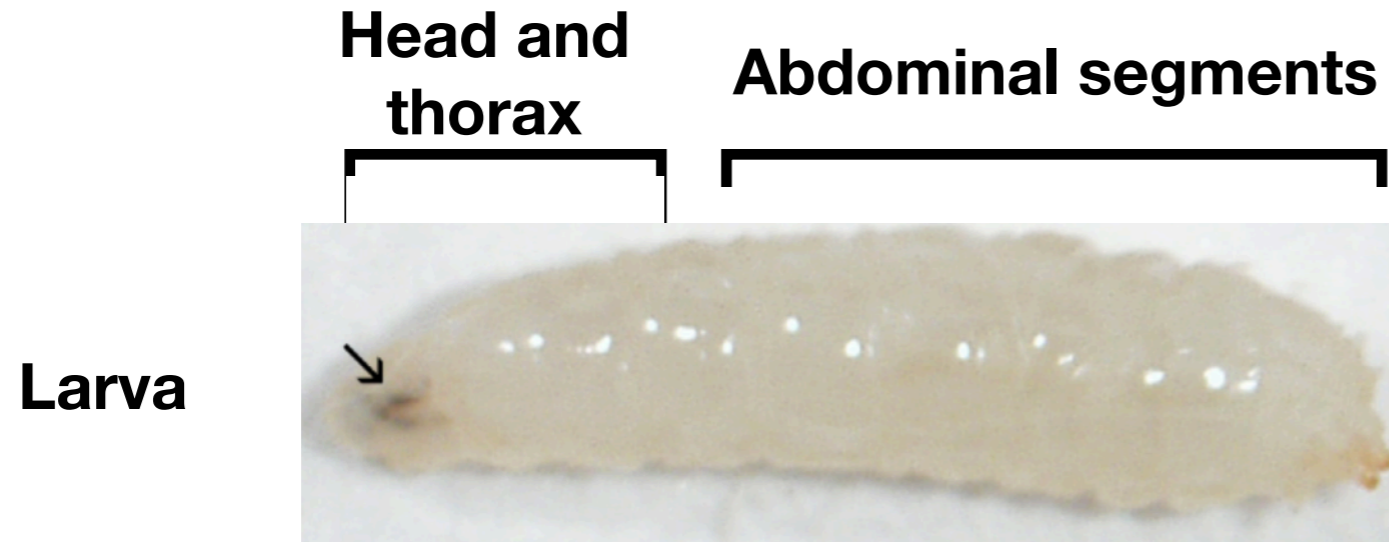


Maternal effect gene

Zygotic genes

Cell fate

Forces/Form

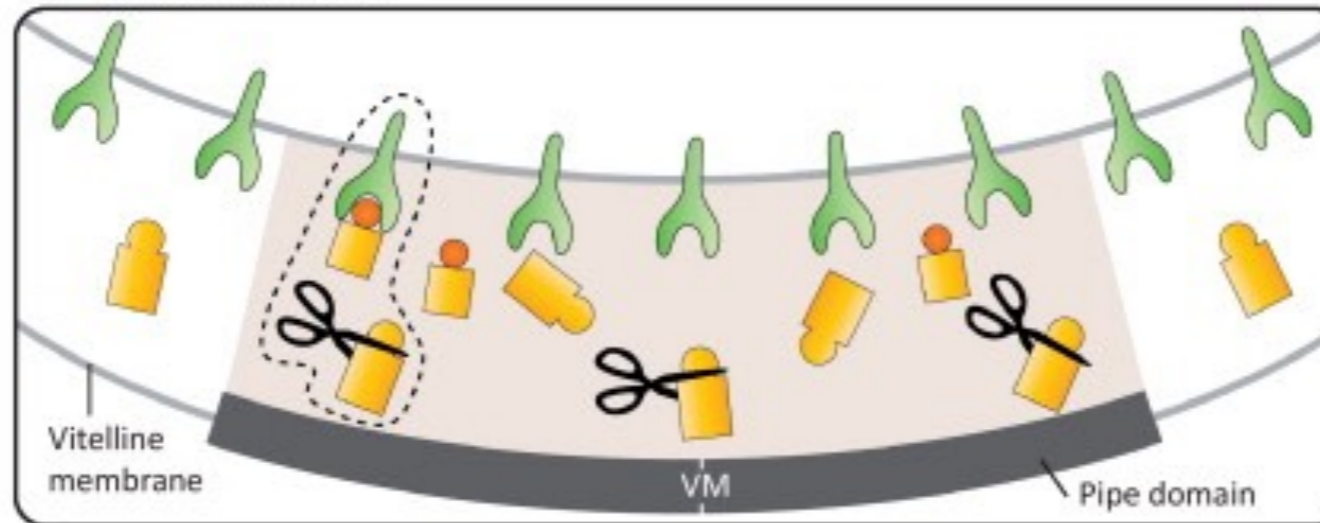


Adult

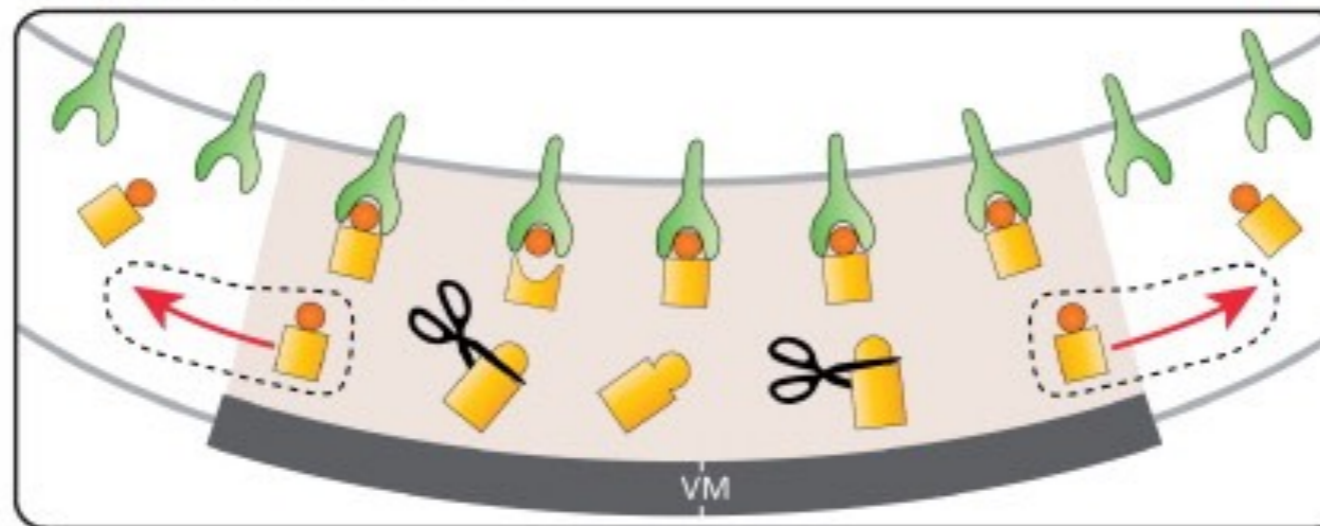


Morphogen signaling leads to gradient in nuclear Dorsal

(a) Spz cleavage by Ea



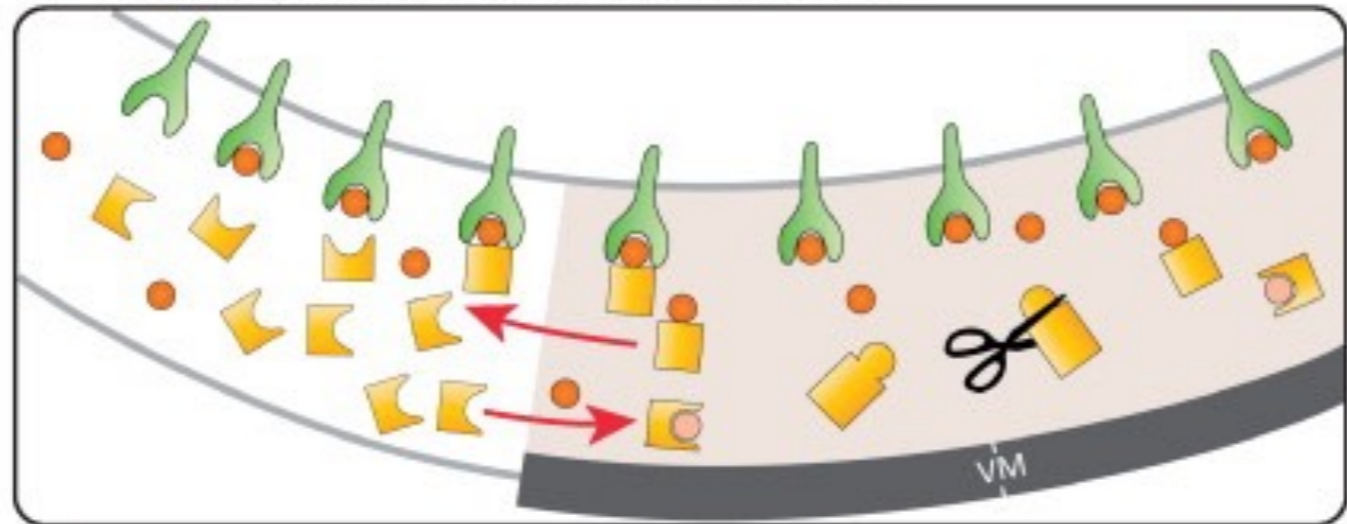
(b) N-C Spz diffuses laterally



Shilo et al., 2013

Morphogen signaling leads to gradient in nuclear Dorsal

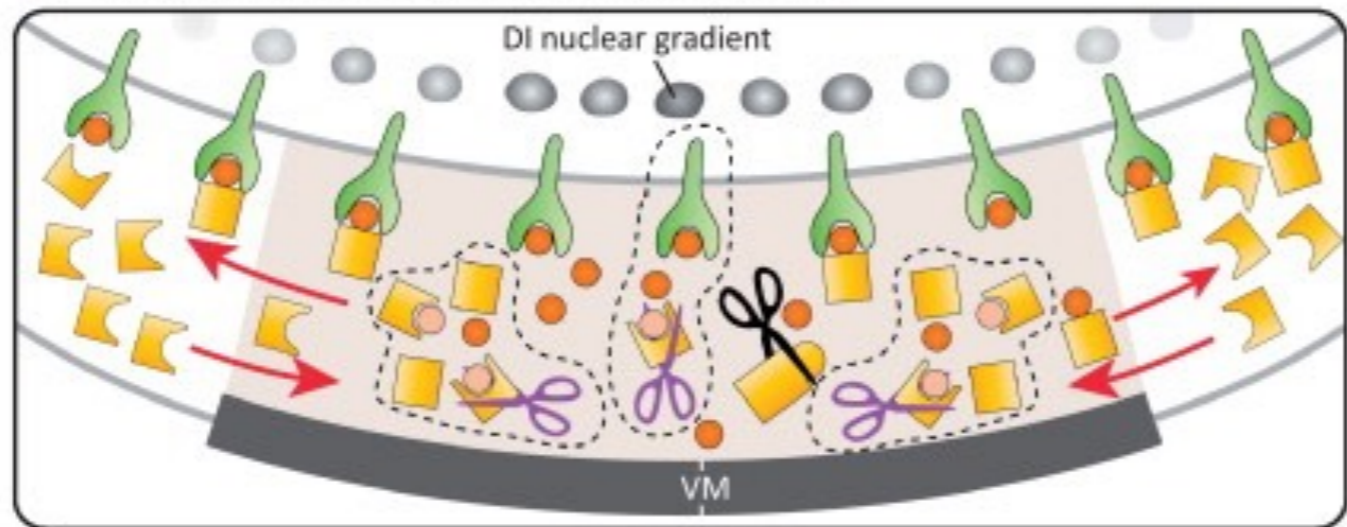
(c) N Spz peaks at edges: ventral flux of C Spz+N Spz



●
■
C Spz (active) and
N Spz (free)

■
○
Reassociation
C Spz+N Spz
(inactive)

(d) Preferential ventral release of C Spz: Shuttling



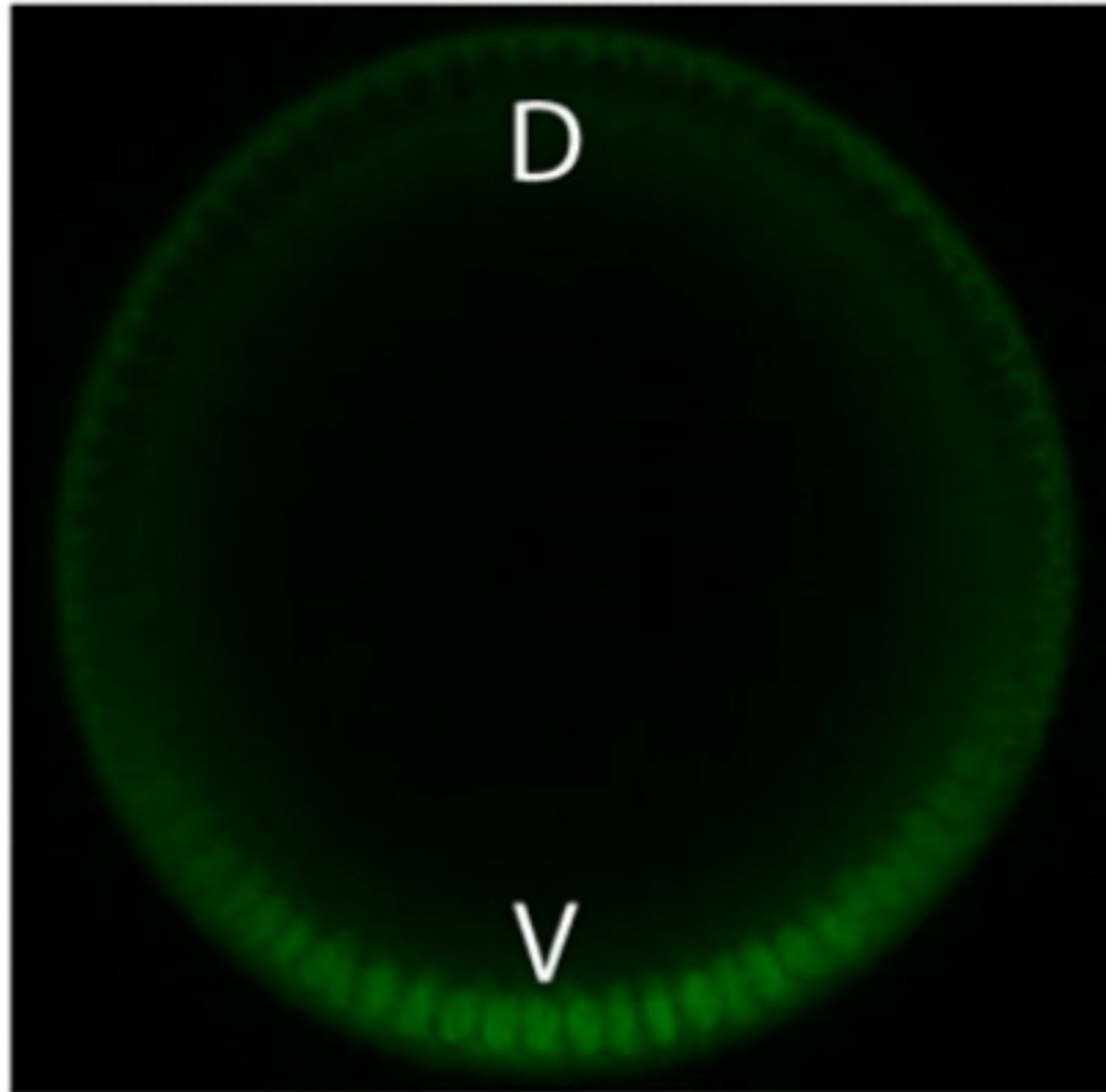
■
○
Cleavage of N Spz
C Spz (active)

TRENDS in Genetics

Shilo et al., 2013

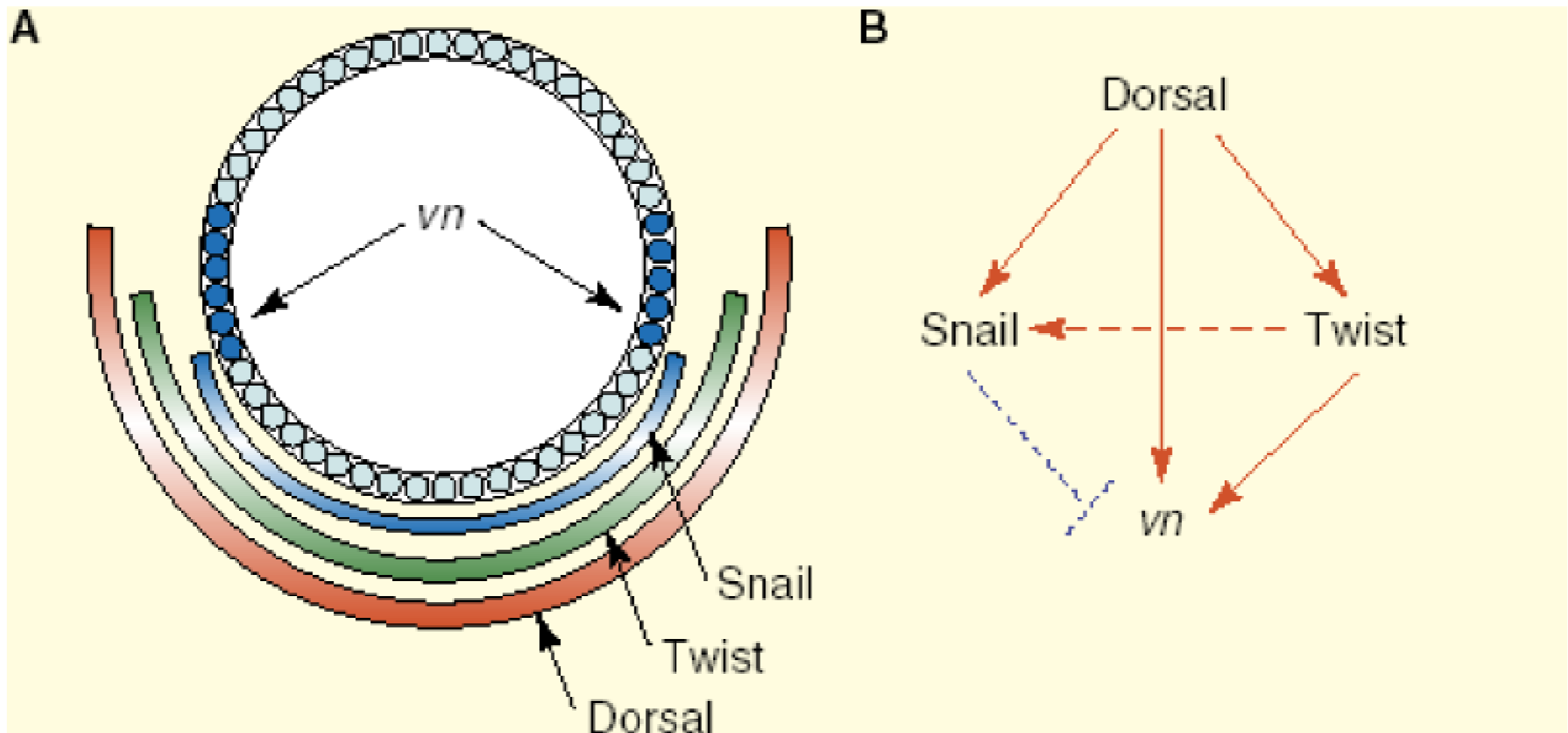
The Dorsal protein defines the ventral side of the embryo

**Dorsal protein
(NF- κ B
transcription
factor)**



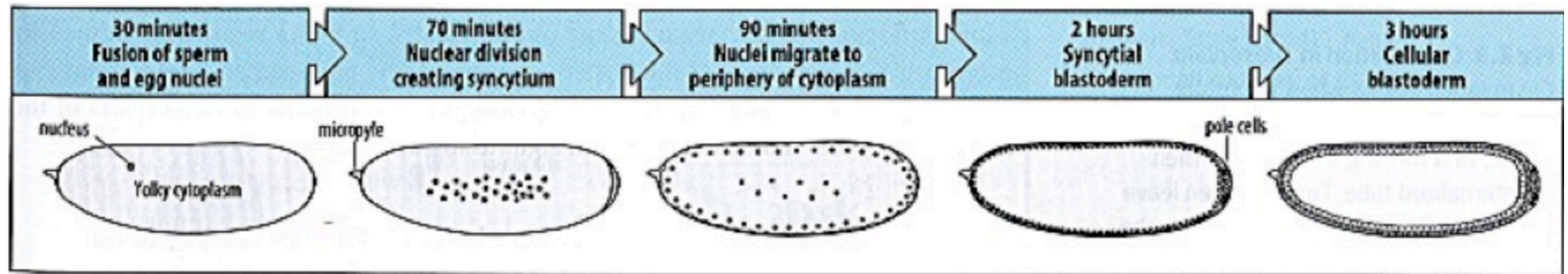
Kanodia et al., 2009

Dorsal concentration and cross-regulation establish domains of gene expression

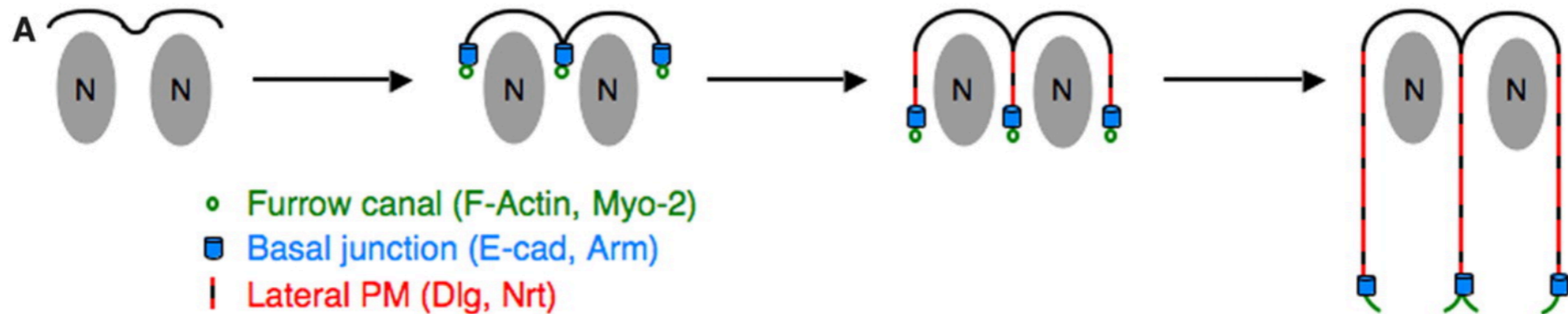
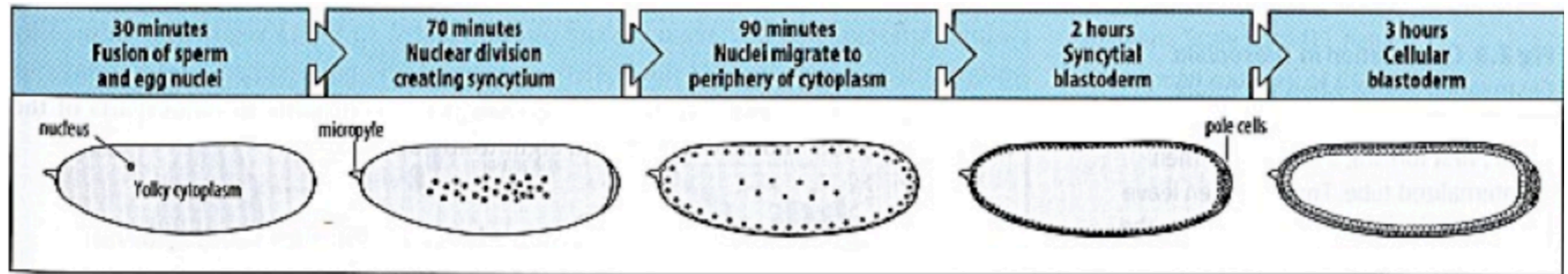


M. Levine's lab

Cellularization: 1 to 6,000 cells in 1 hour

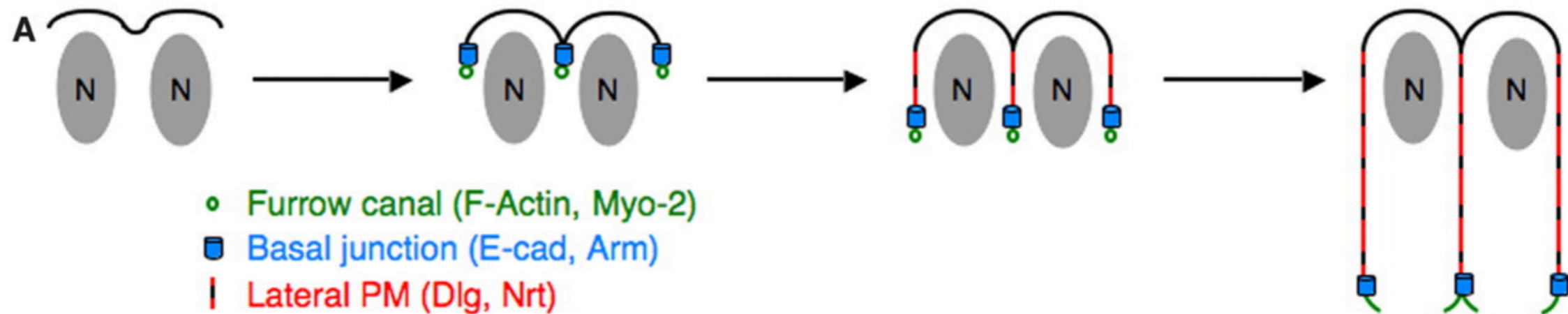
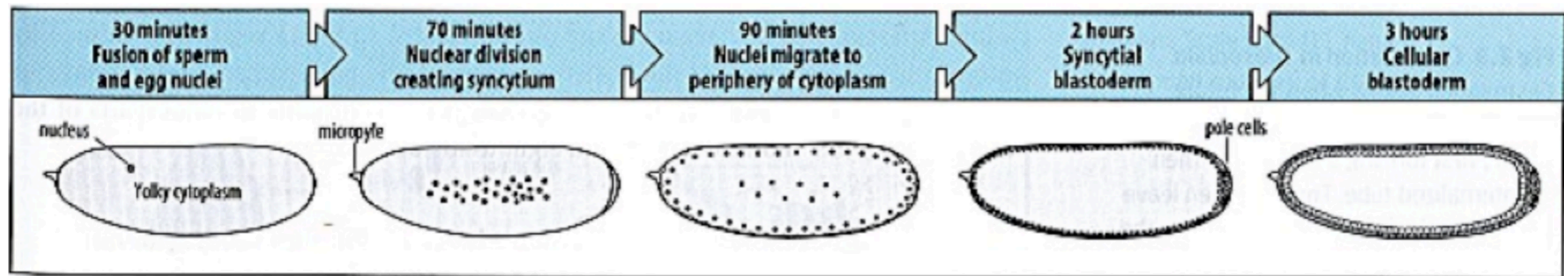


Cellularization: 1 to 6,000 cells in 1 hour

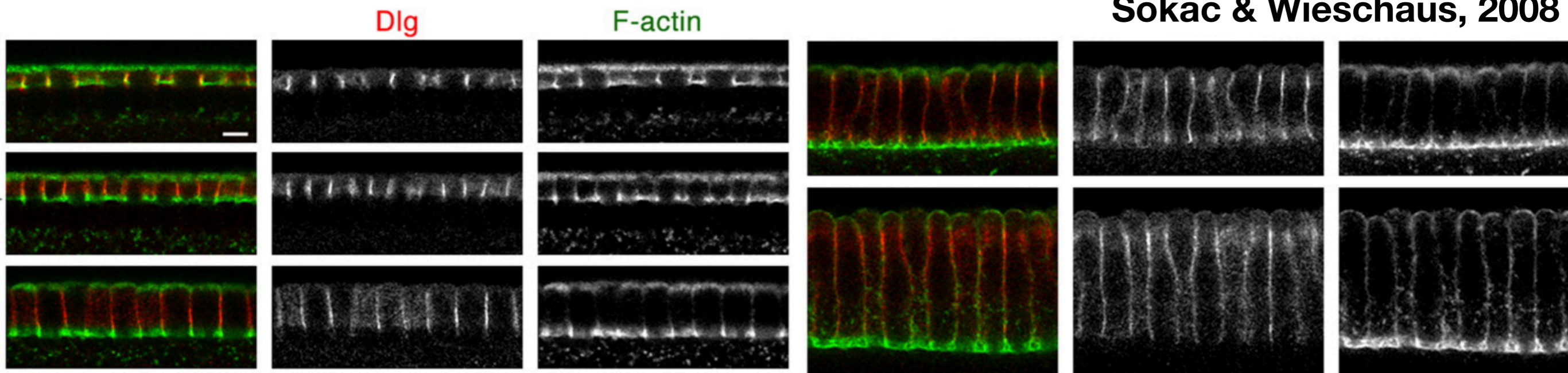


Sokac & Wieschaus, 2008

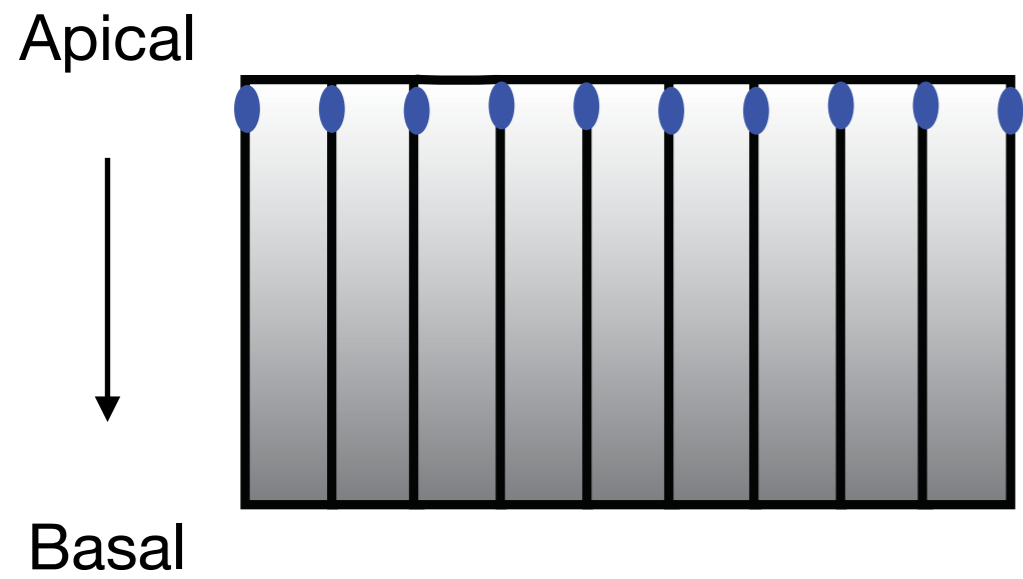
Cellularization: 1 to 6,000 cells in 1 hour



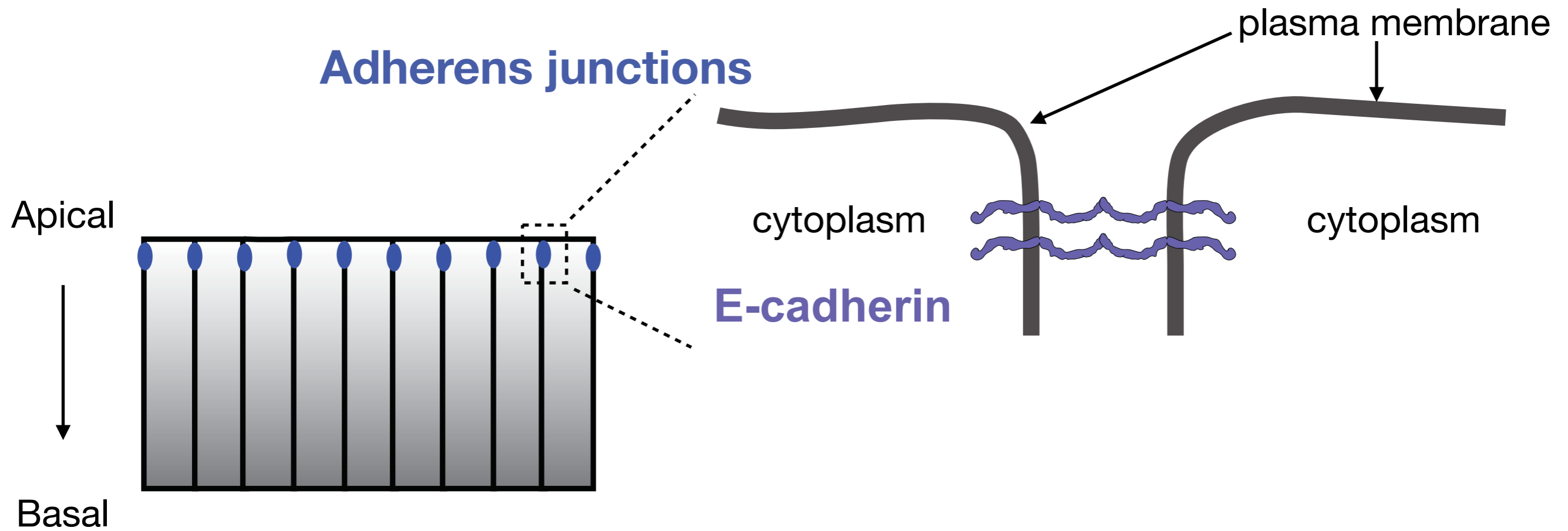
Sokac & Wieschaus, 2008



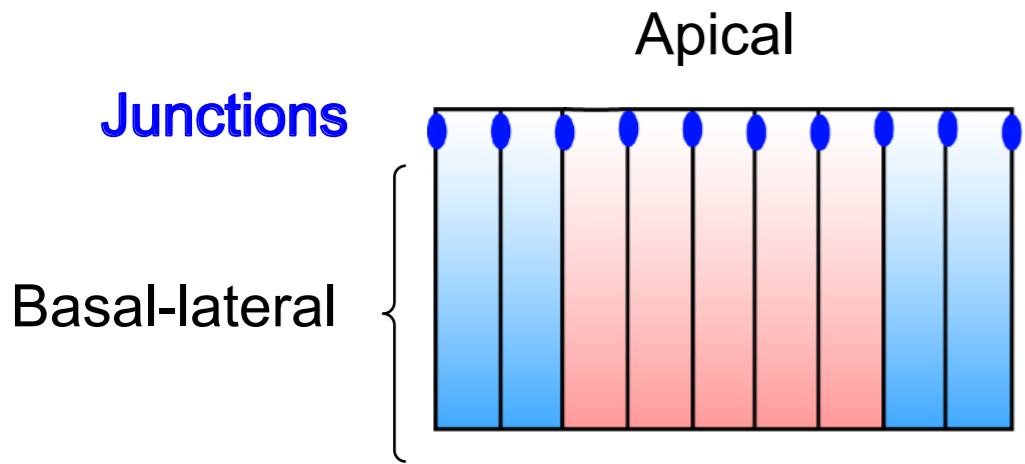
Cells that form are epithelial in nature



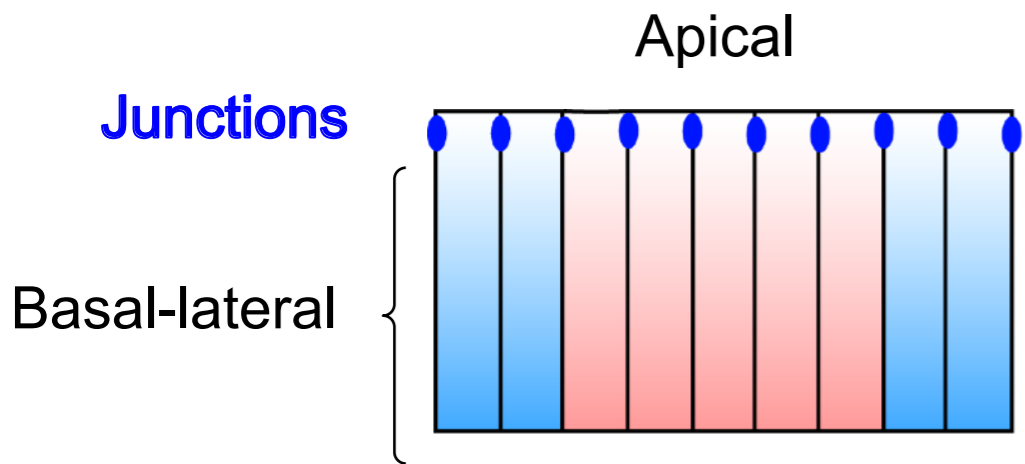
Cells that form are epithelial in nature



Epithelial cells undergo various shape changes and rearrangements to sculpt tissues



Epithelial cells undergo various shape changes and rearrangements to sculpt tissues



(a) Cell intercalation



(b) Rotation



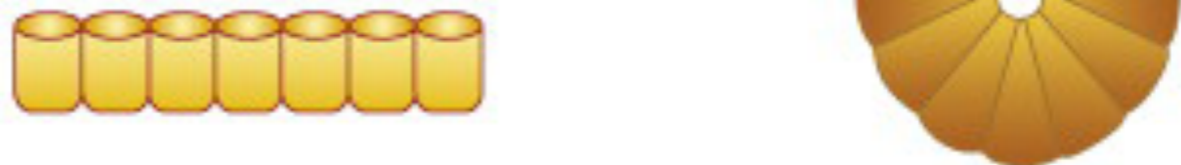
(c) Invagination



(d) Sheet extension

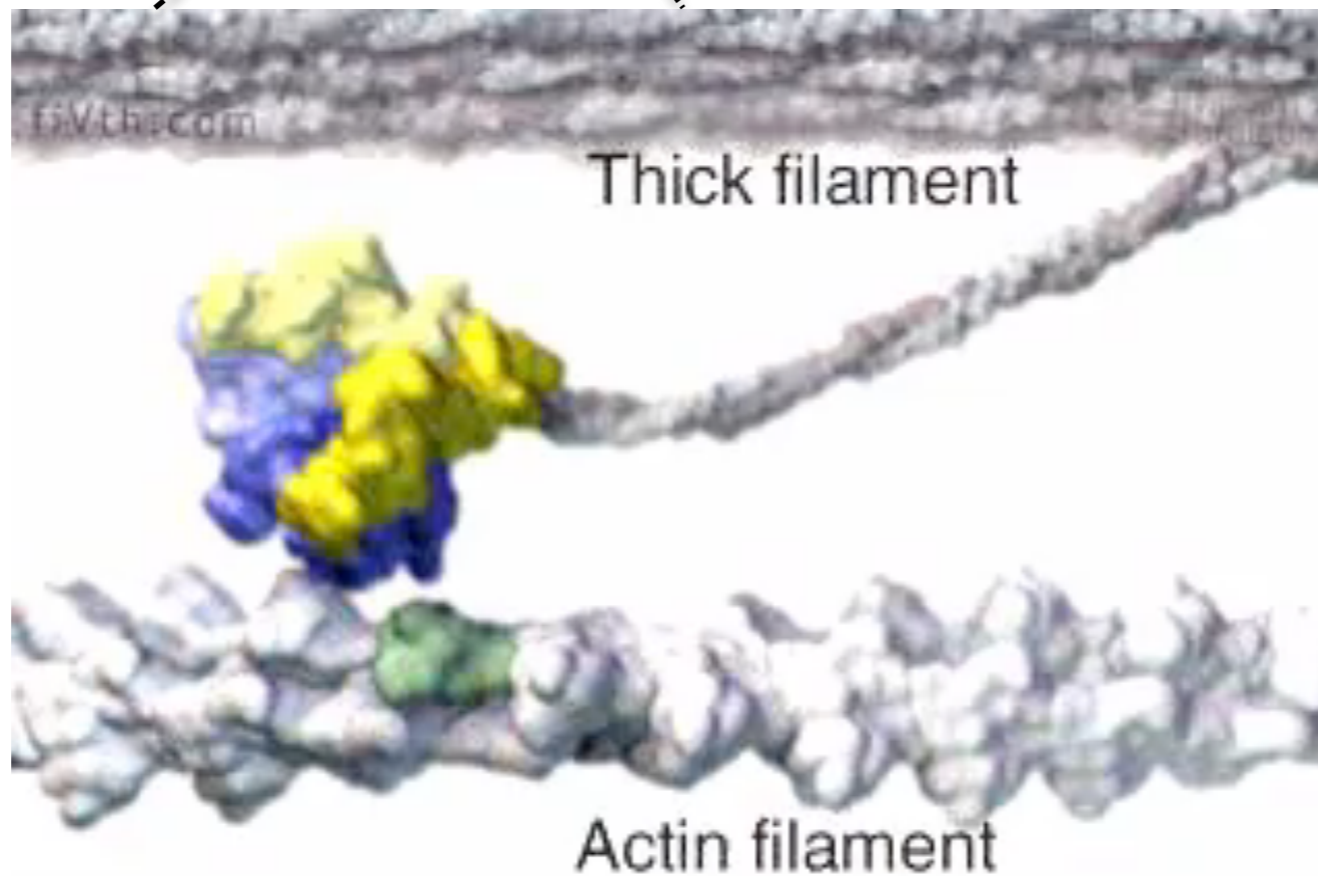
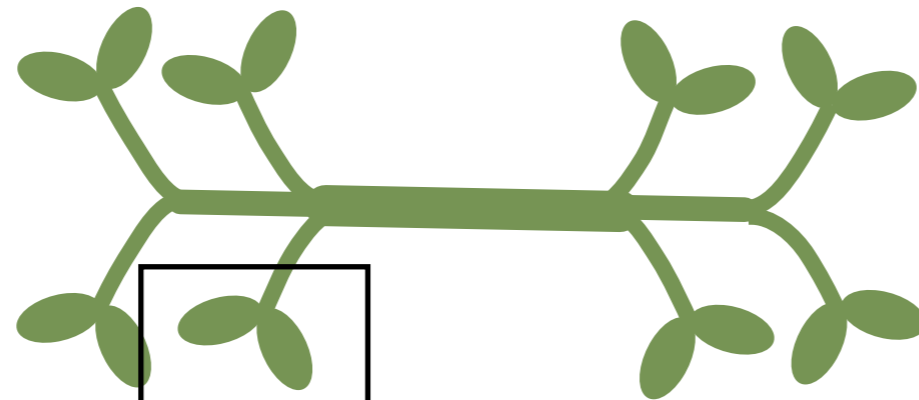


(e) Tube formation



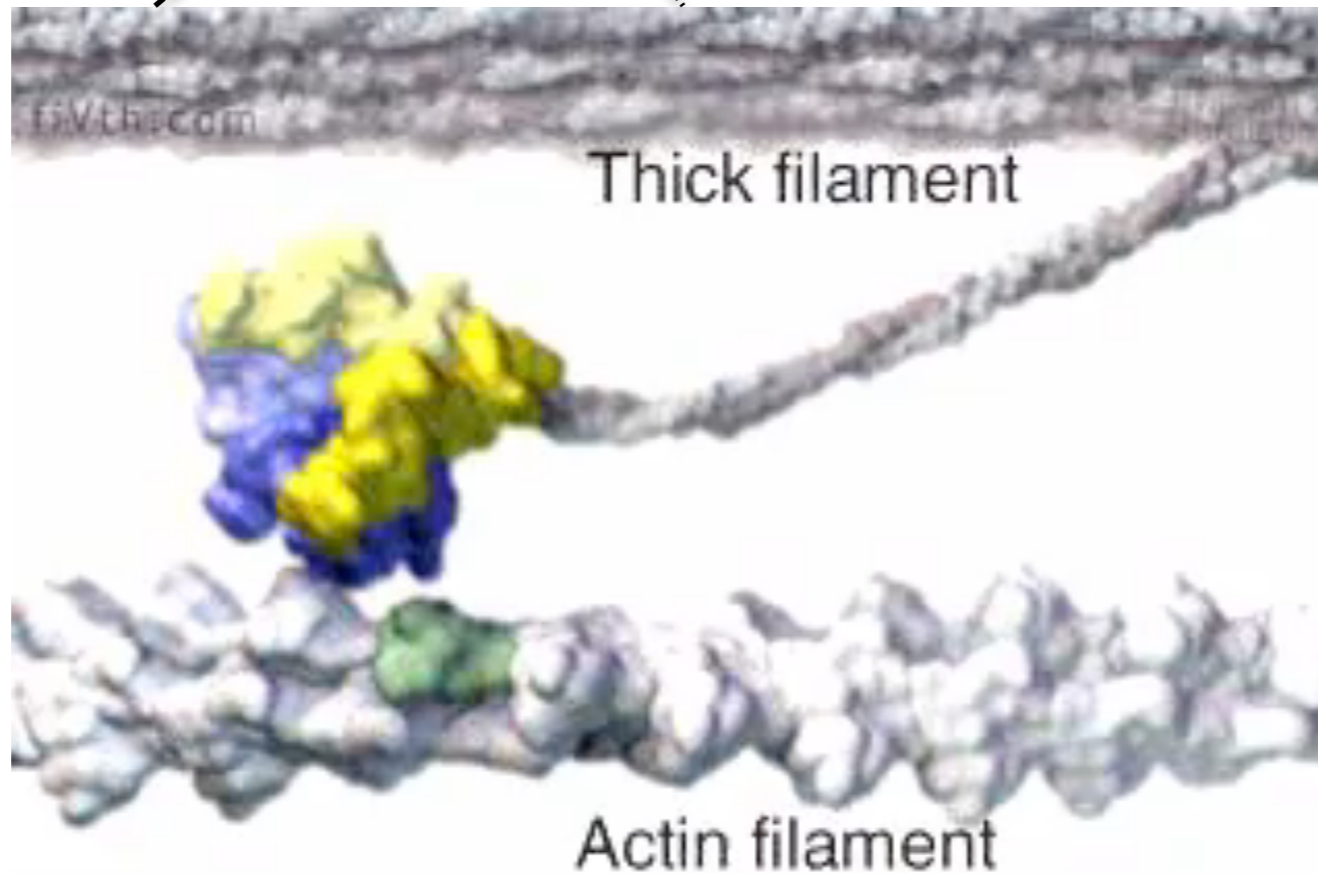
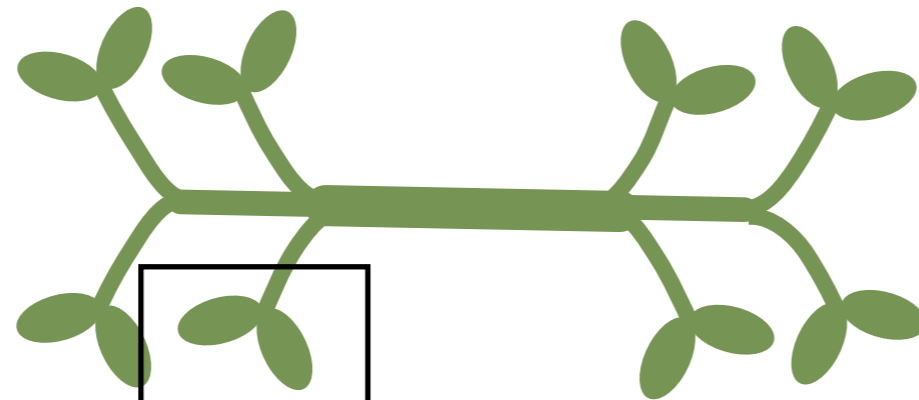
Quintin . . . Labouesse.
Trends Genet. 2008.

Myosin 2 (myosin) is thought to generate force to drive morphogenesis



Vale & Milligan, *Science* (2000).

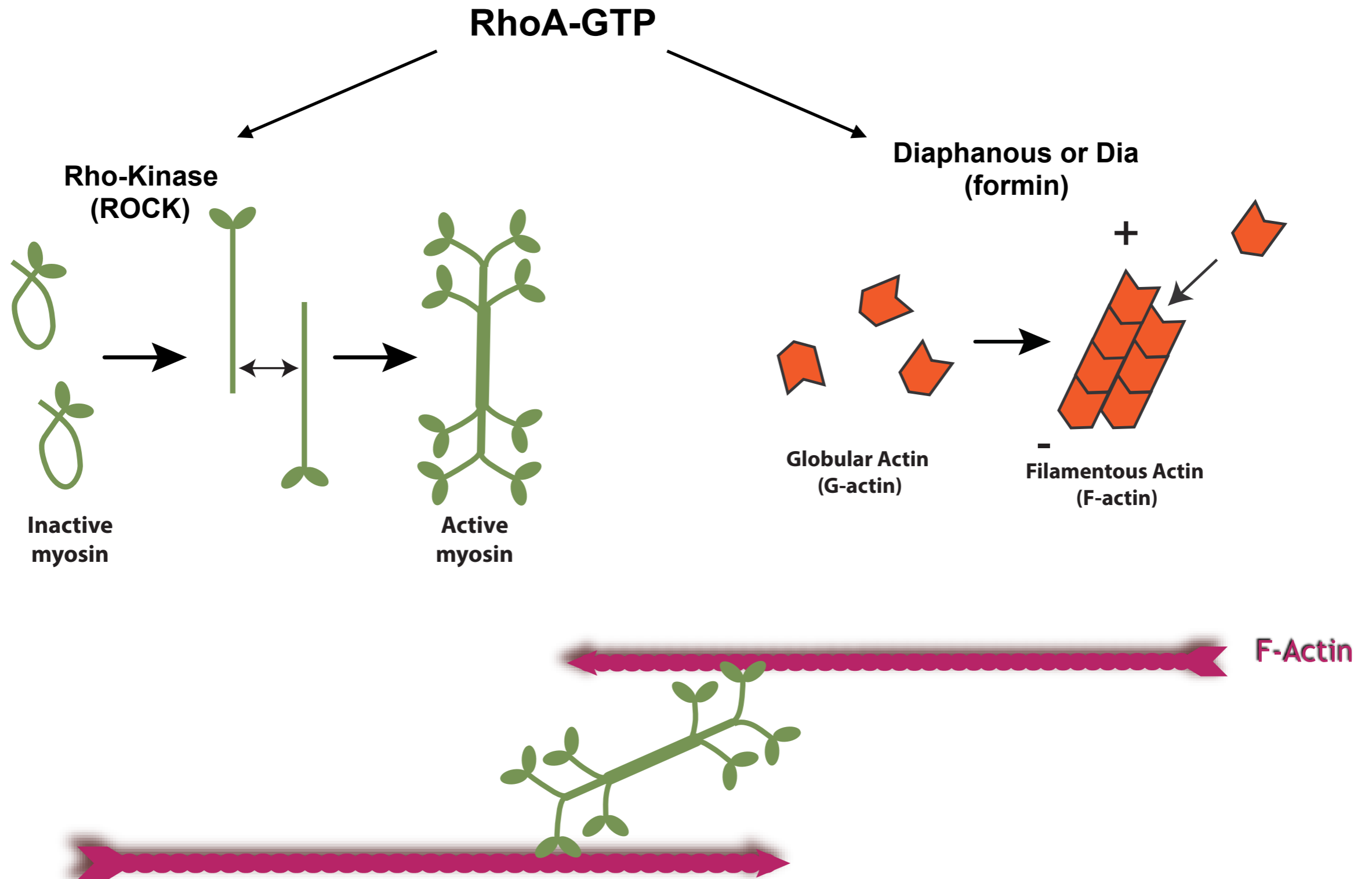
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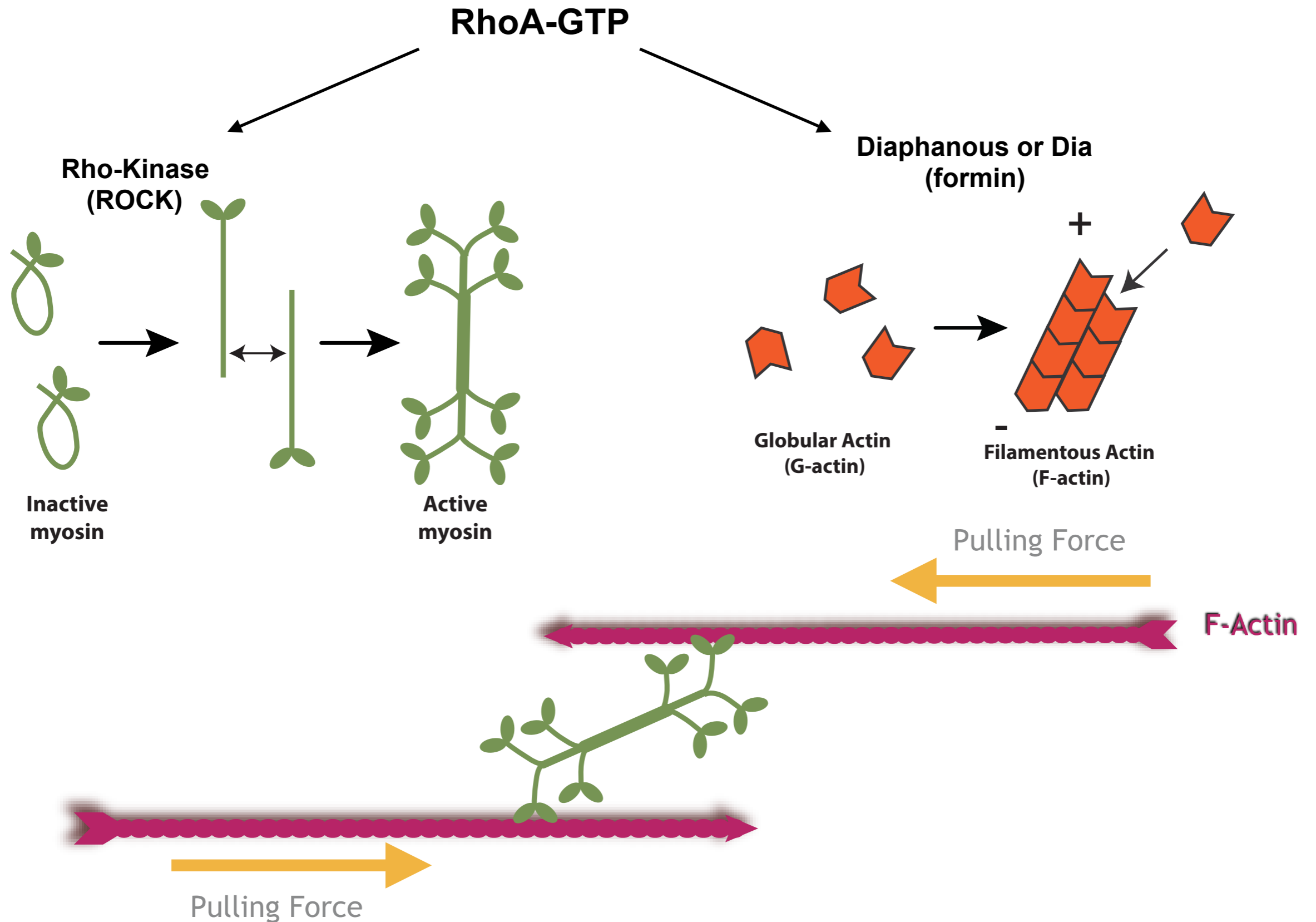
Vale & Milligan, *Science* (2000).

tiVth.com

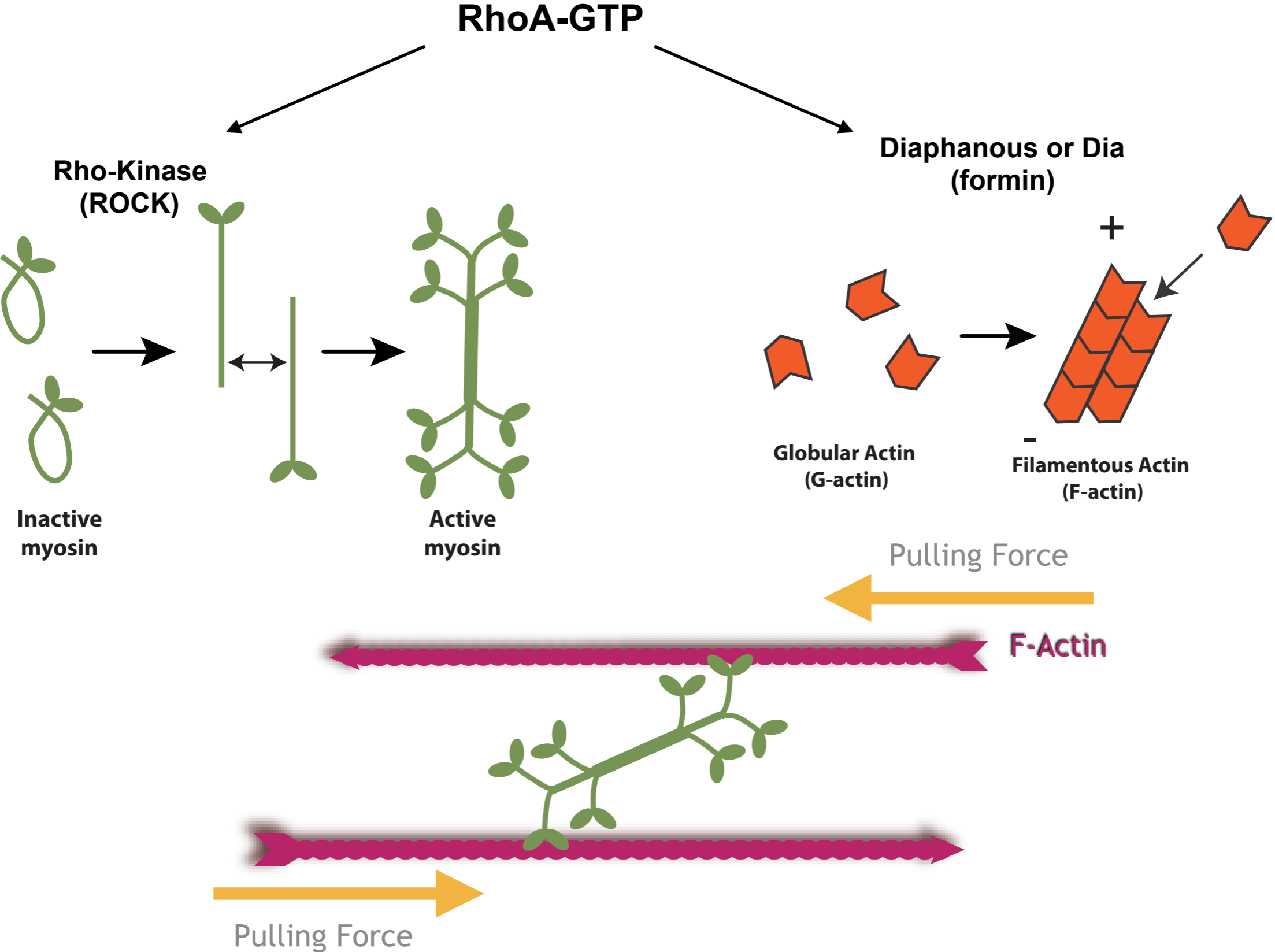
RhoA promotes actomyosin contraction



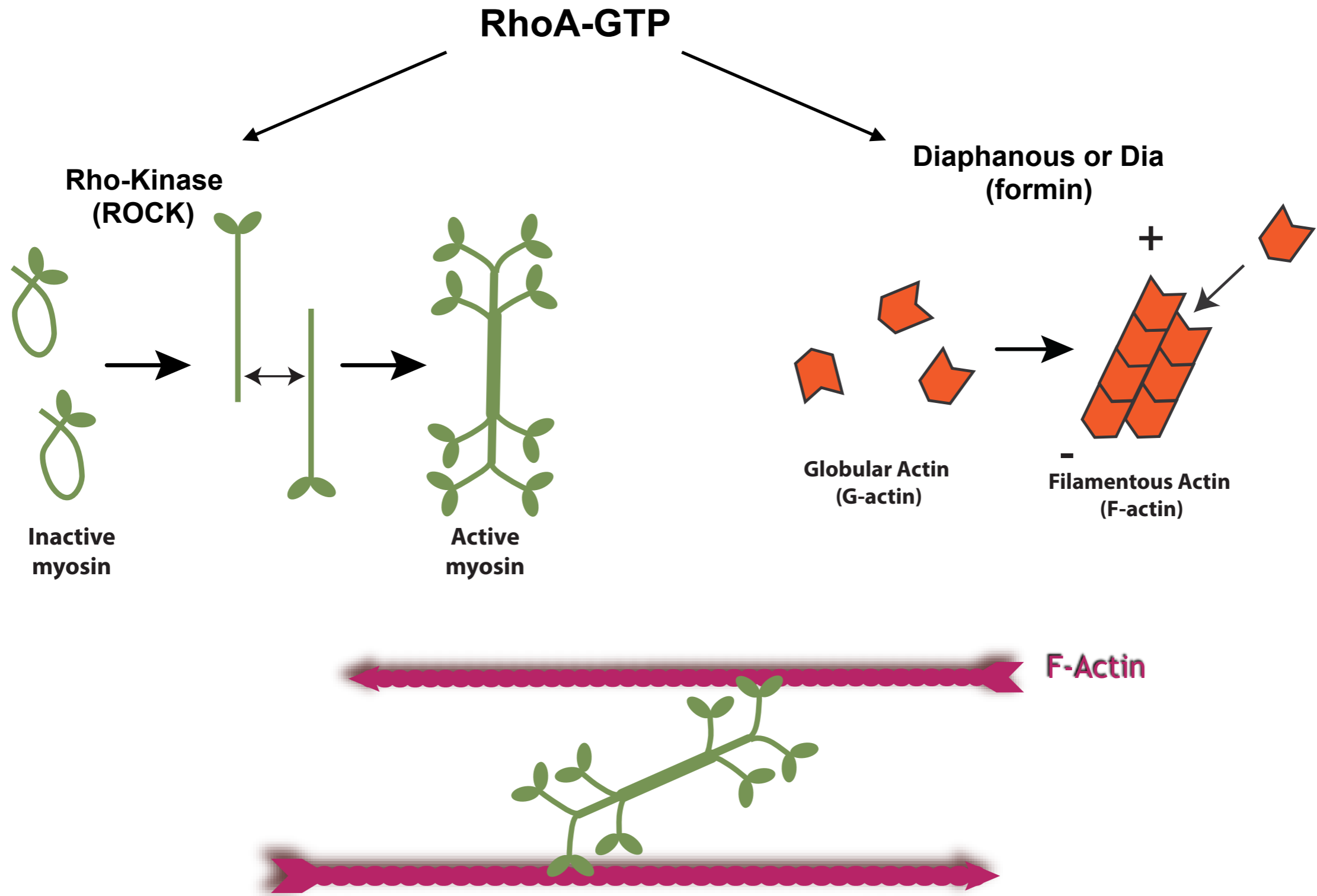
RhoA promotes actomyosin contraction



RhoA promotes actomyosin contraction

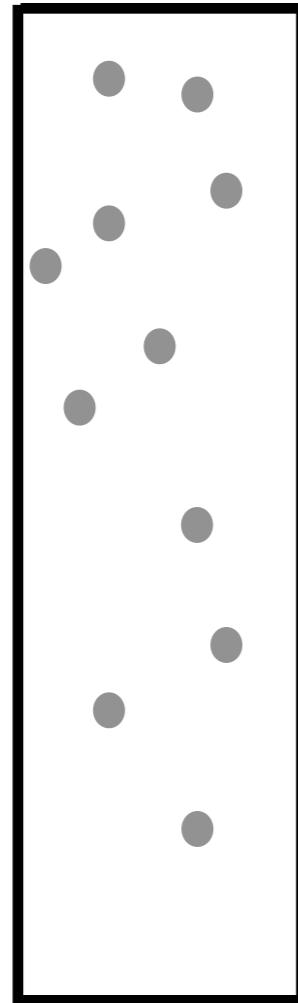


RhoA promotes actomyosin contraction



Simple model for constriction

Apical

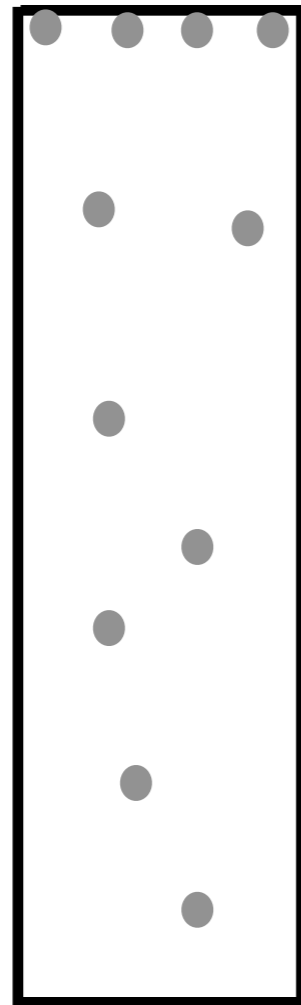


● **Inactive RhoA-GDP**

Basal

Simple model for constriction

Apical

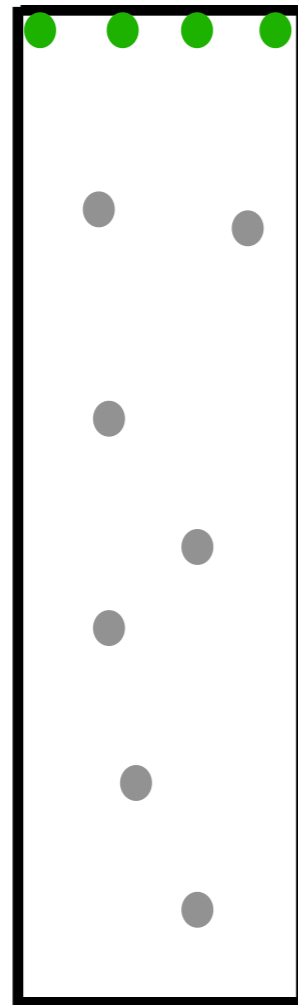


● **Inactive RhoA-GDP**

Basal

Simple model for constriction

Apical



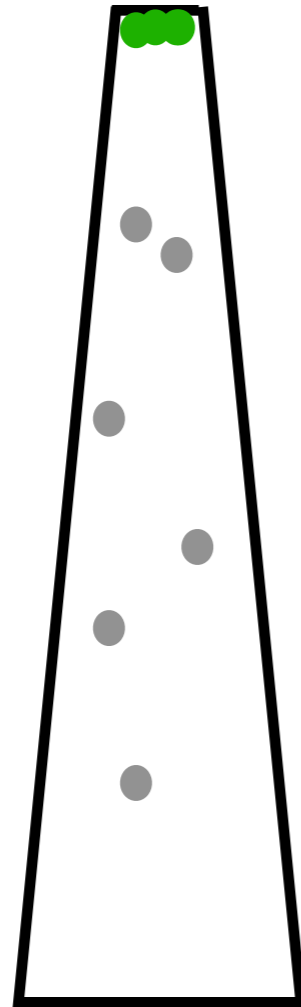
● Inactive RhoA-GDP

● Active RhoA-GTP

Basal

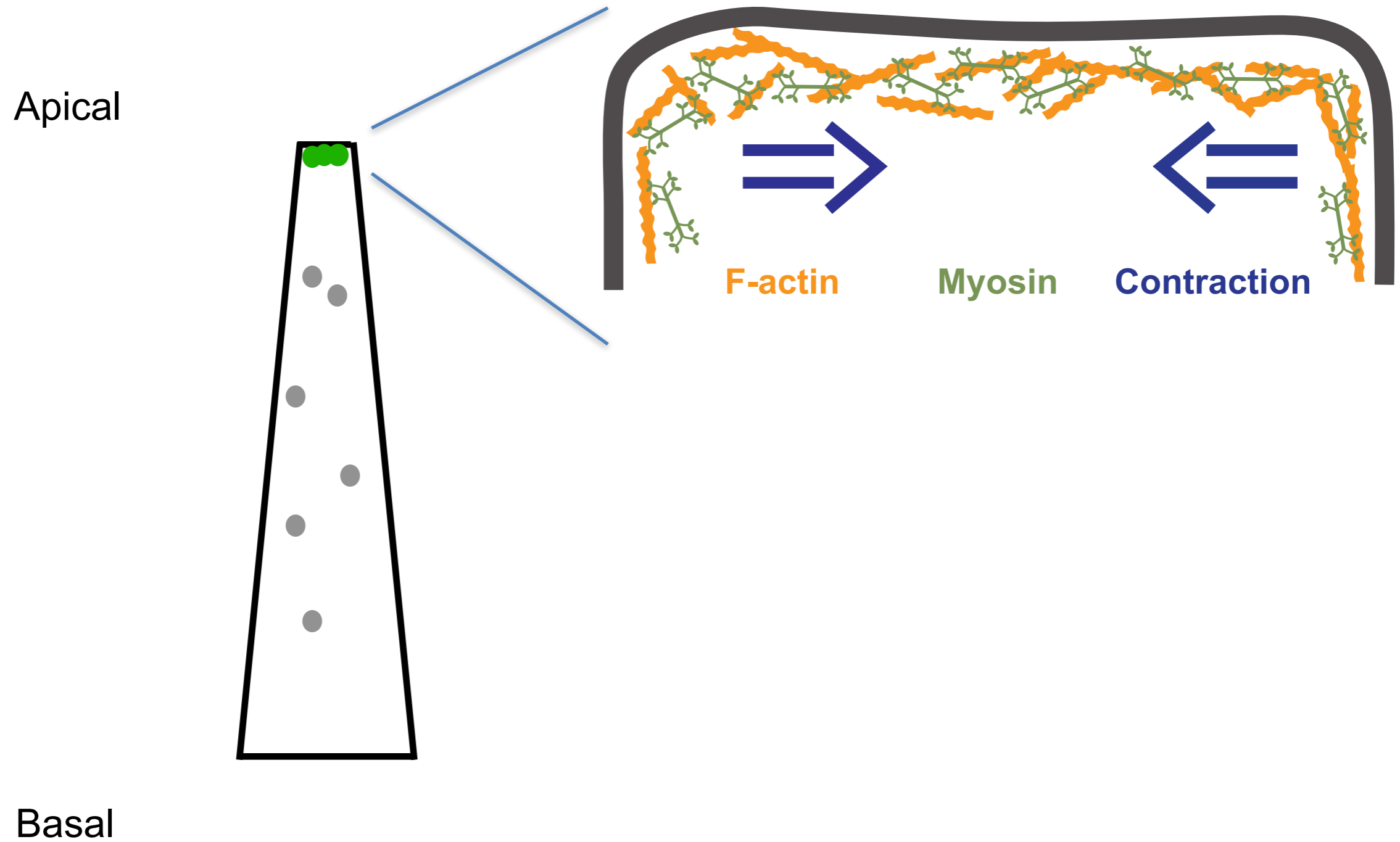
Simple model for constriction

Apical

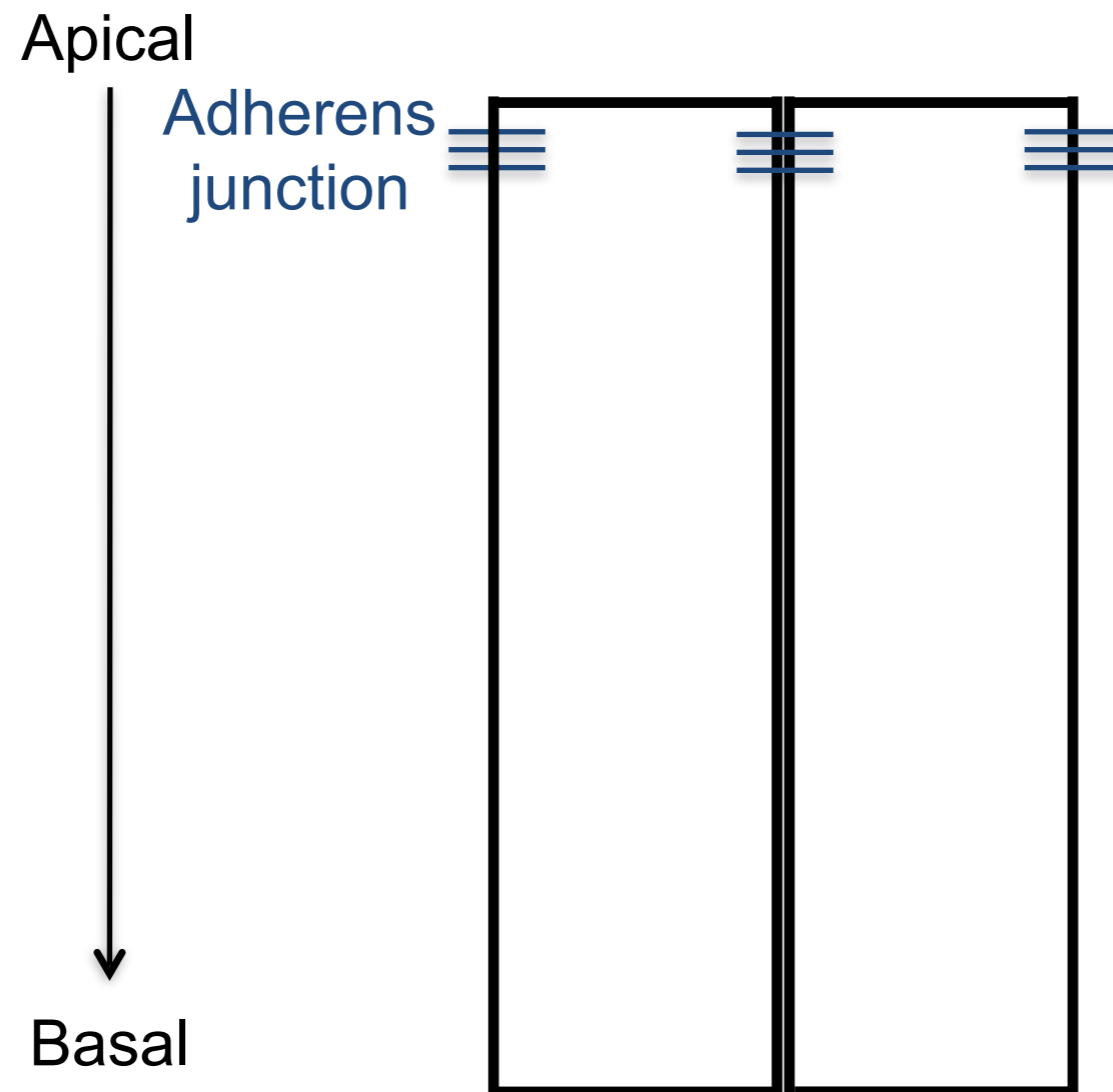


Basal

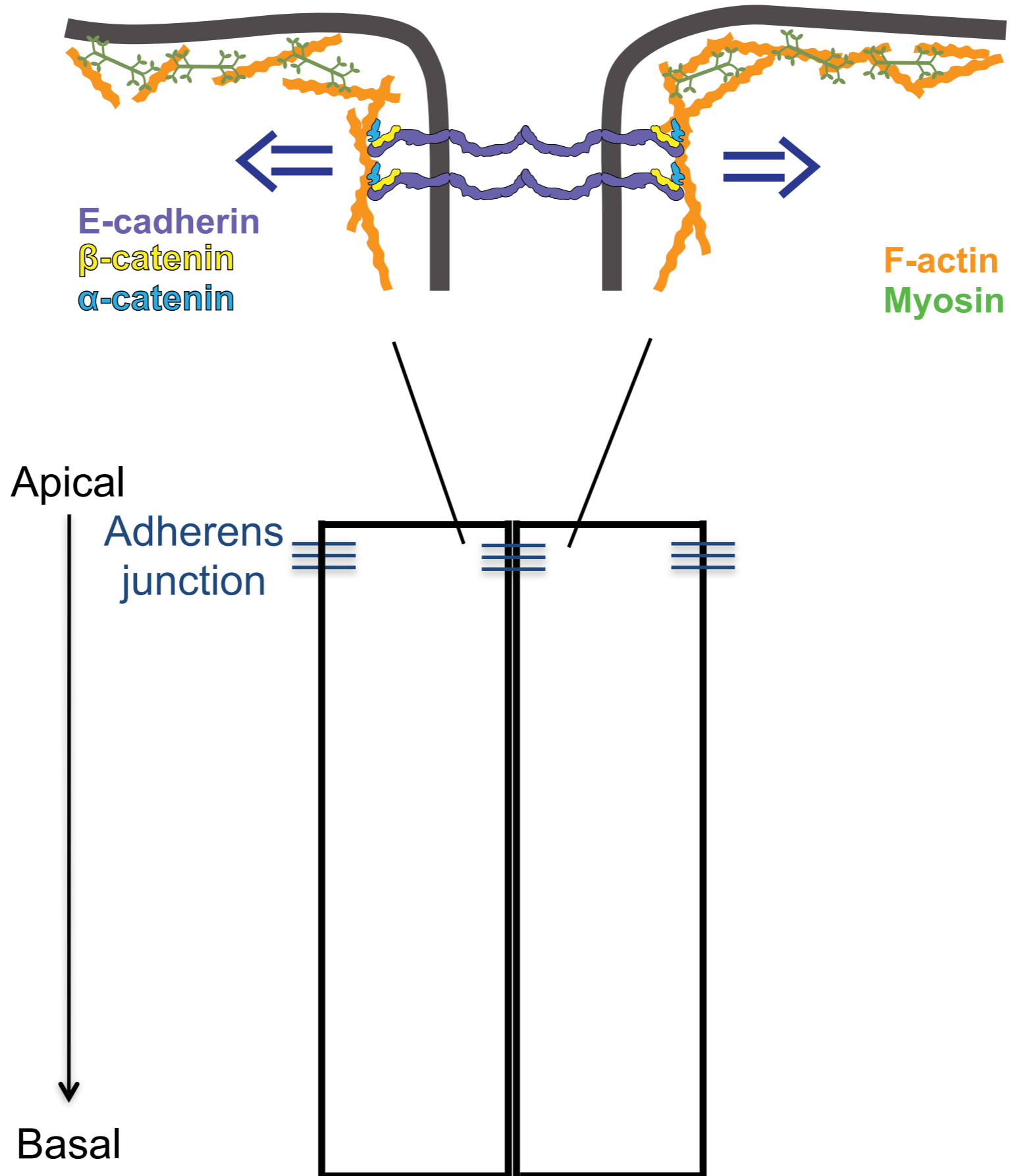
Simple model for constriction



Adherens junctions serve as anchor points for contraction to pull

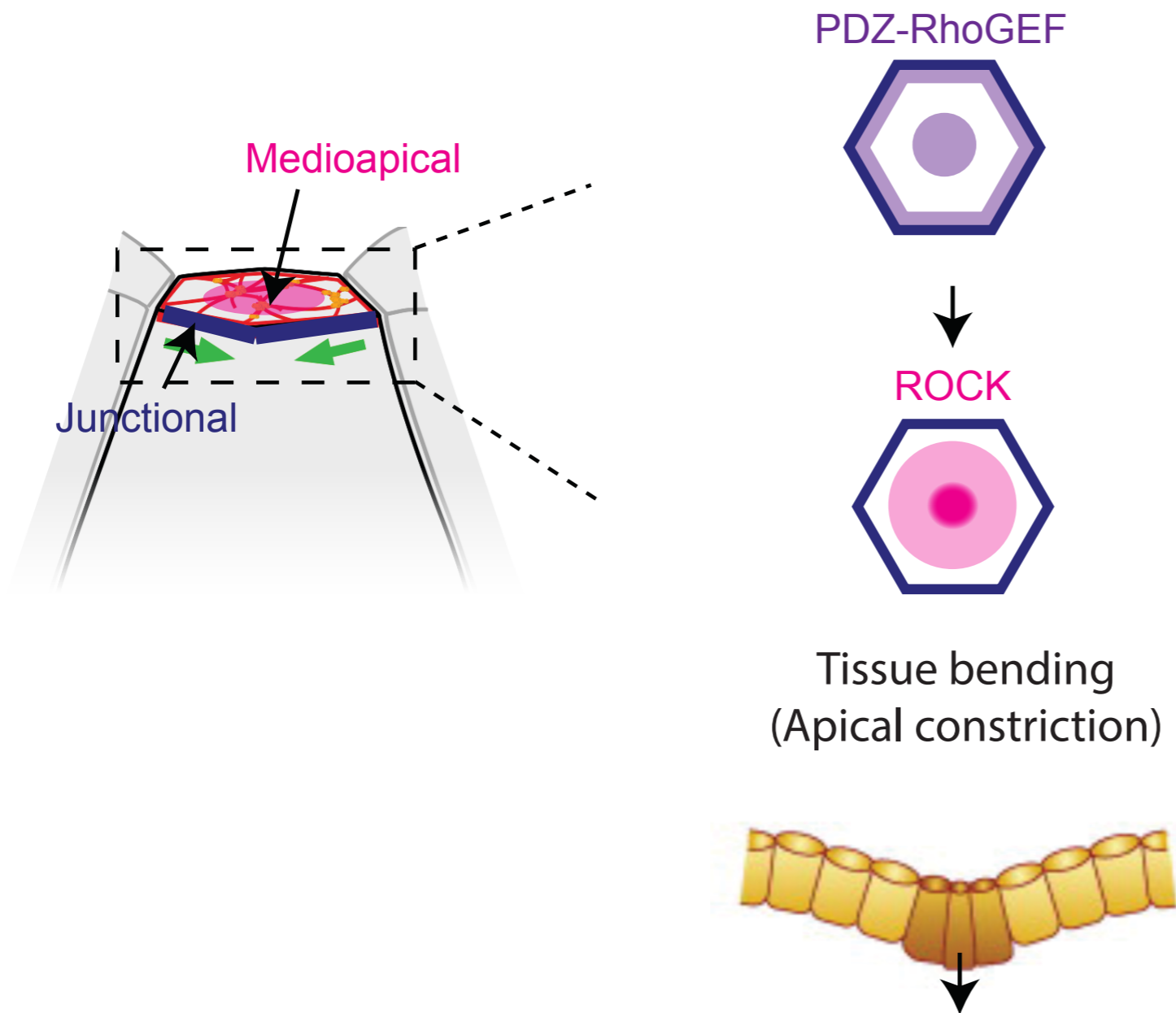


Adherens junctions serve as anchor points for contraction to pull



Distinct morphogenetic programs arise from different patterns of cell contractility

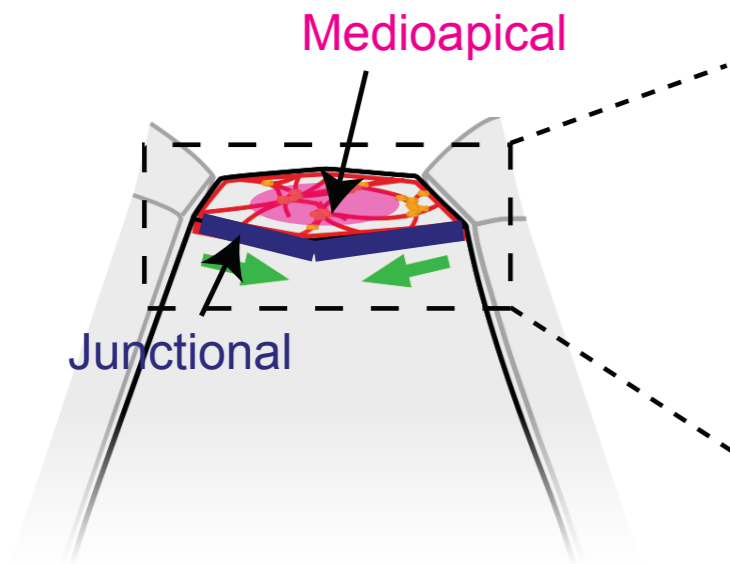
Radial Cell Polarity (RCP)



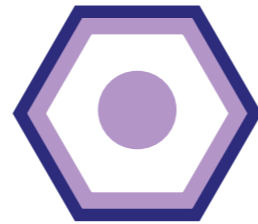
Distinct morphogenetic programs arise from different patterns of cell contractility

Radial Cell Polarity (RCP)

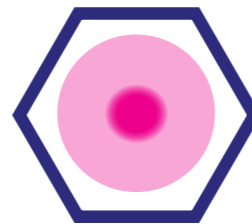
Planar Cell Polarity (PCP)



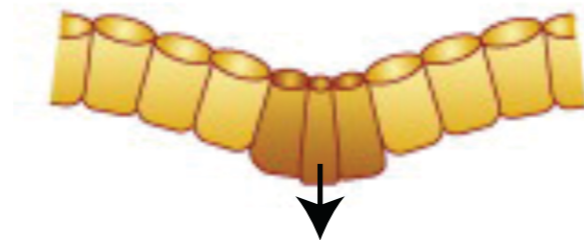
PDZ-RhoGEF



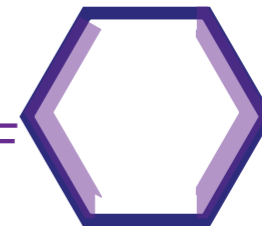
ROCK



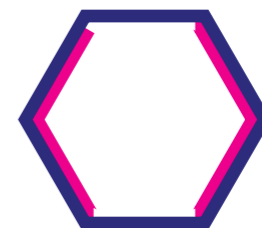
Tissue bending
(Apical constriction)



PDZ-RhoGEF



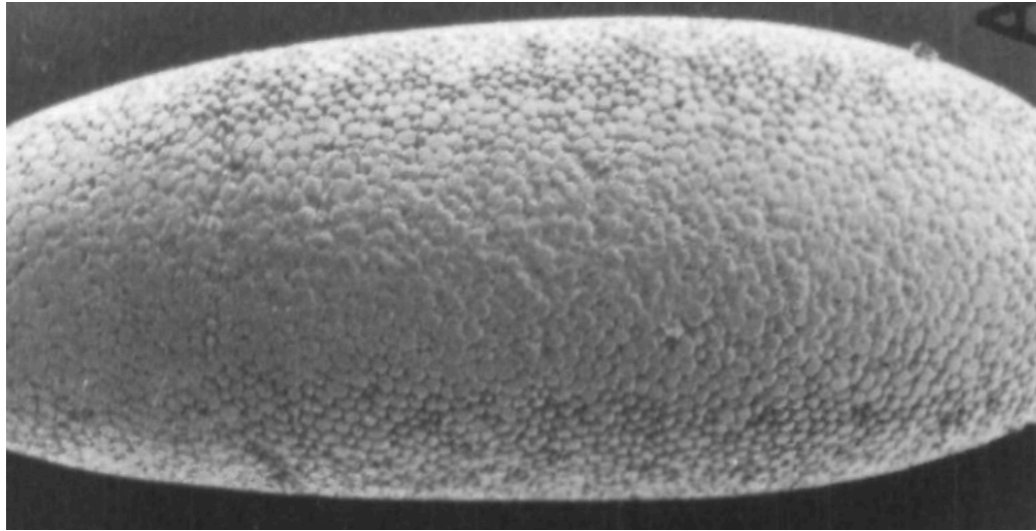
ROCK



Tissue elongation
(Cell intercalation)

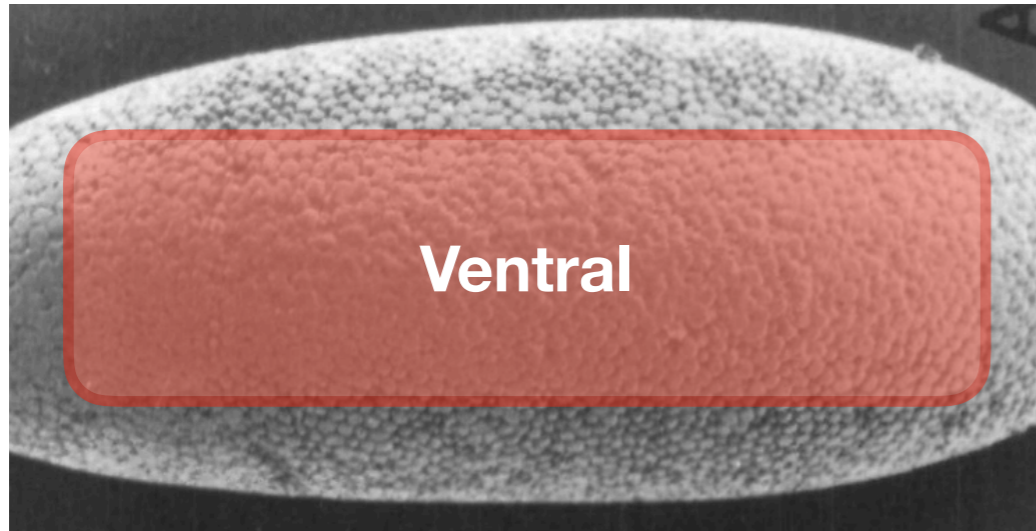


In Drosophila, ventral cells fold inward / invaginate



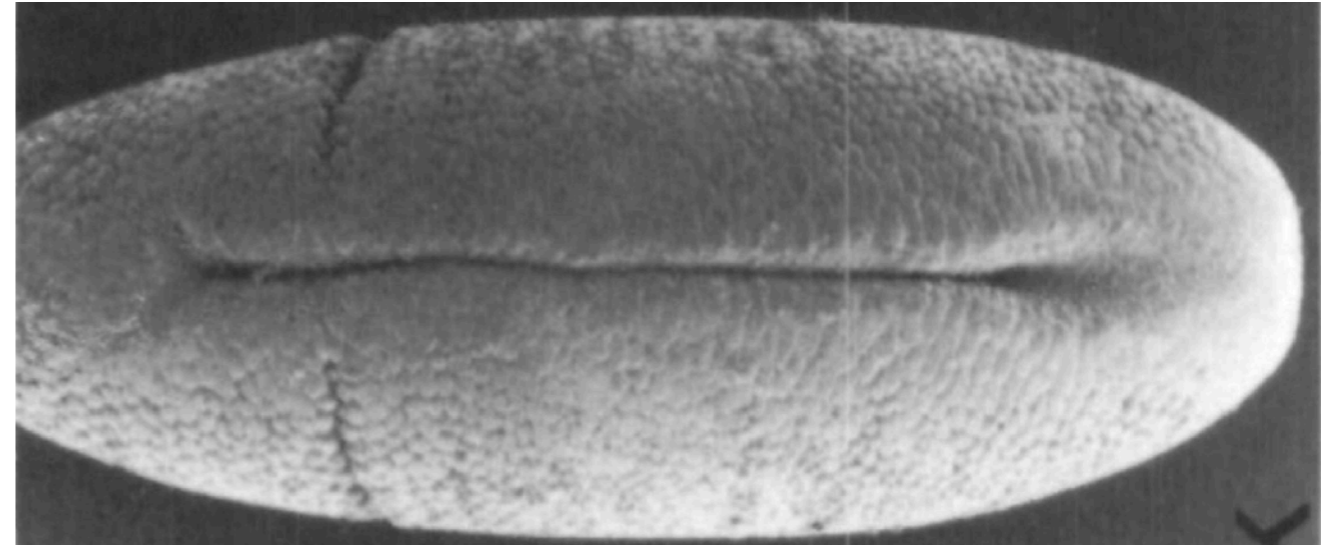
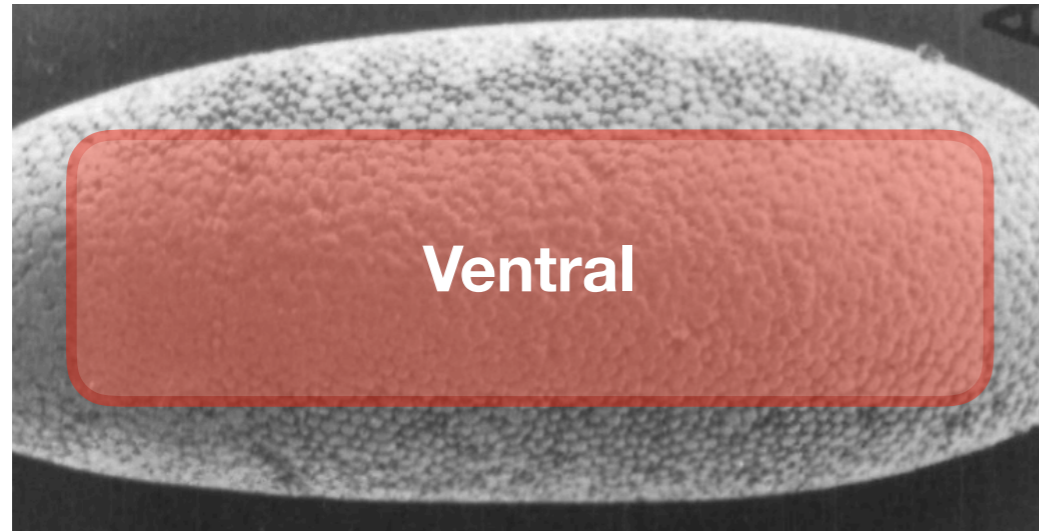
Sweeton, D., et al. *Development* (1991)

In Drosophila, ventral cells fold inward / invaginate



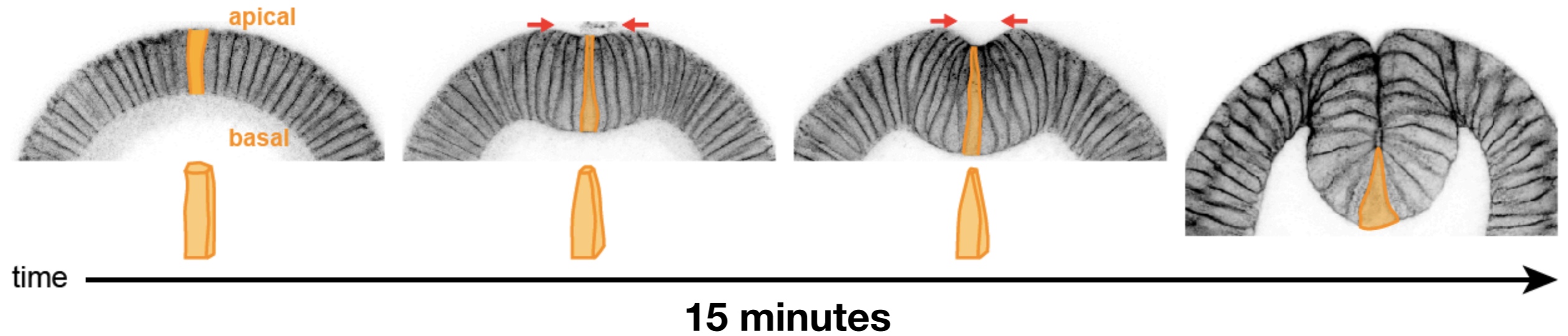
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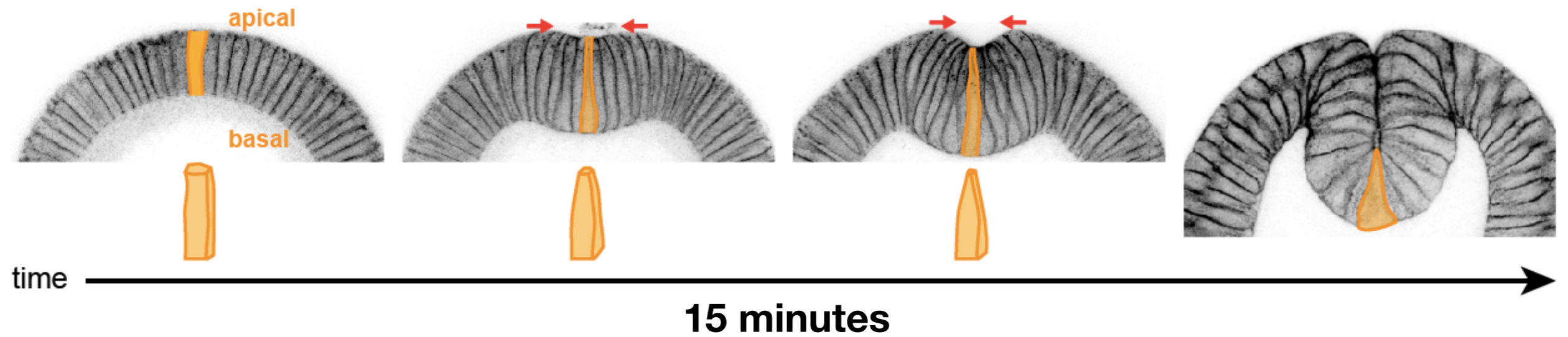


Sweeton, D., et al. *Development* (1991)

Apical constriction is associated with tissue folding



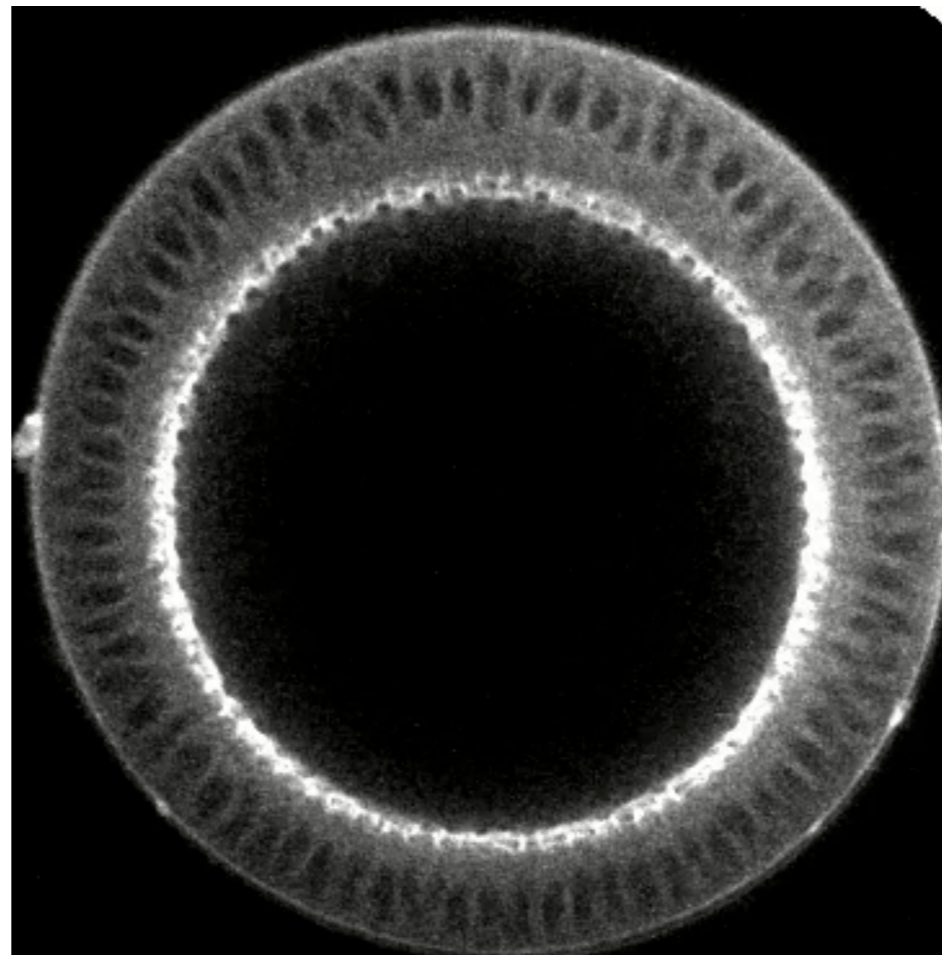
Apical constriction is associated with tissue folding



Ventral

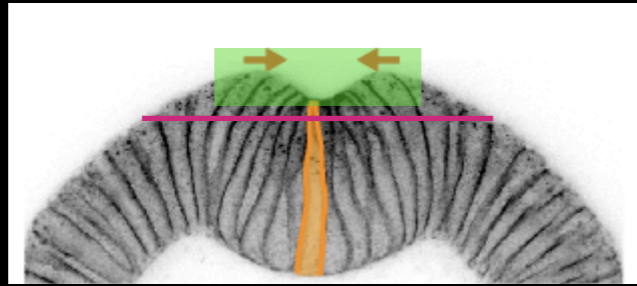


Dorsal



Brodland et al., *PNAS* (2000).

Myosin 2 (myosin) accumulates apically during folding

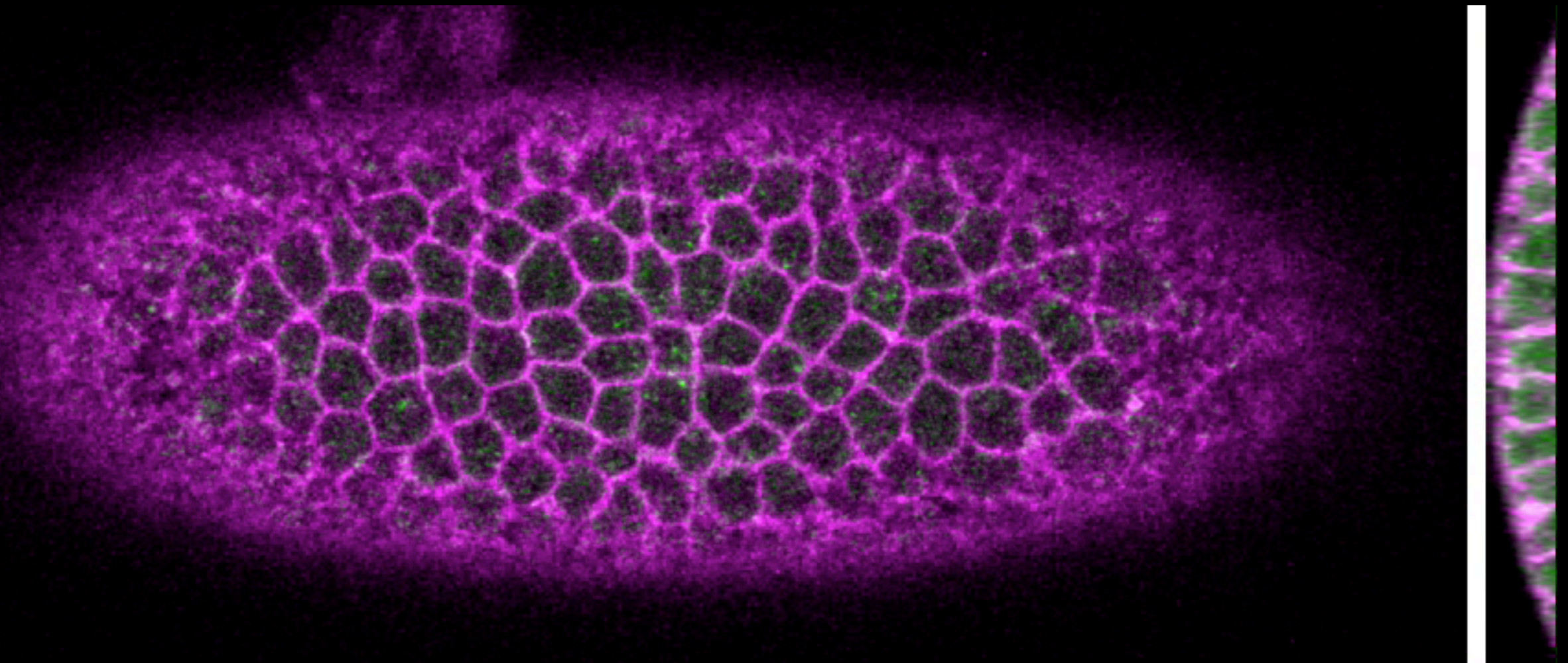


Myosin

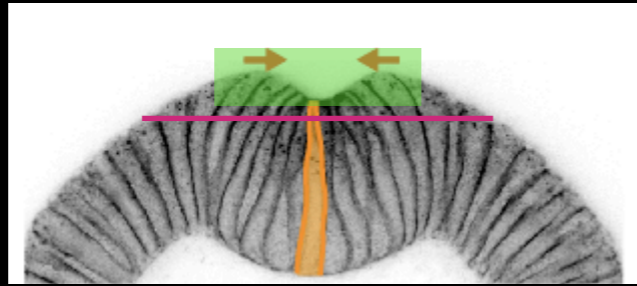
Membrane

Claudia Vasquez

Cross-
Section
Apical → Basal



Myosin 2 (myosin) accumulates apically during folding

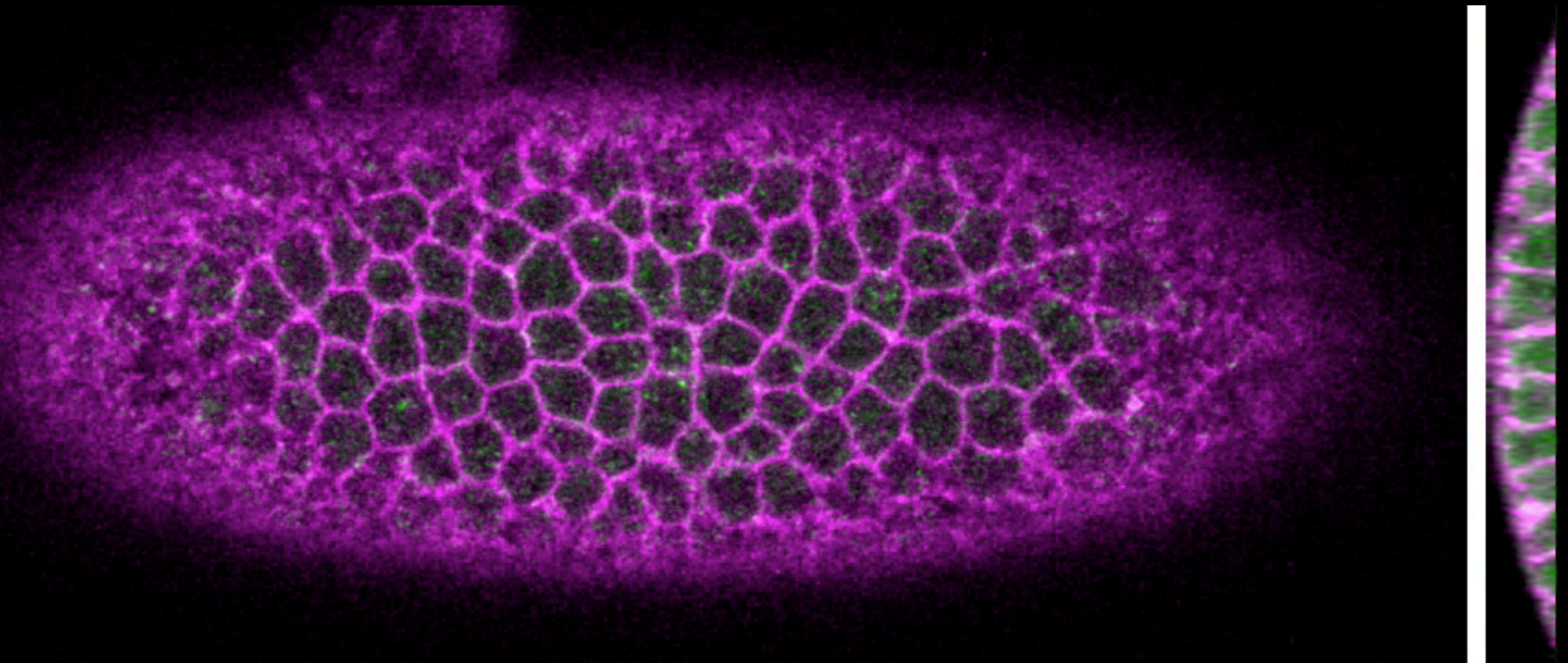


Myosin

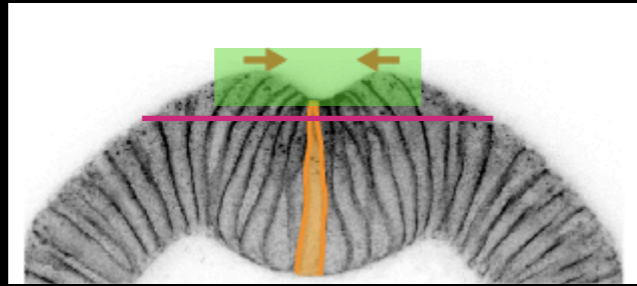
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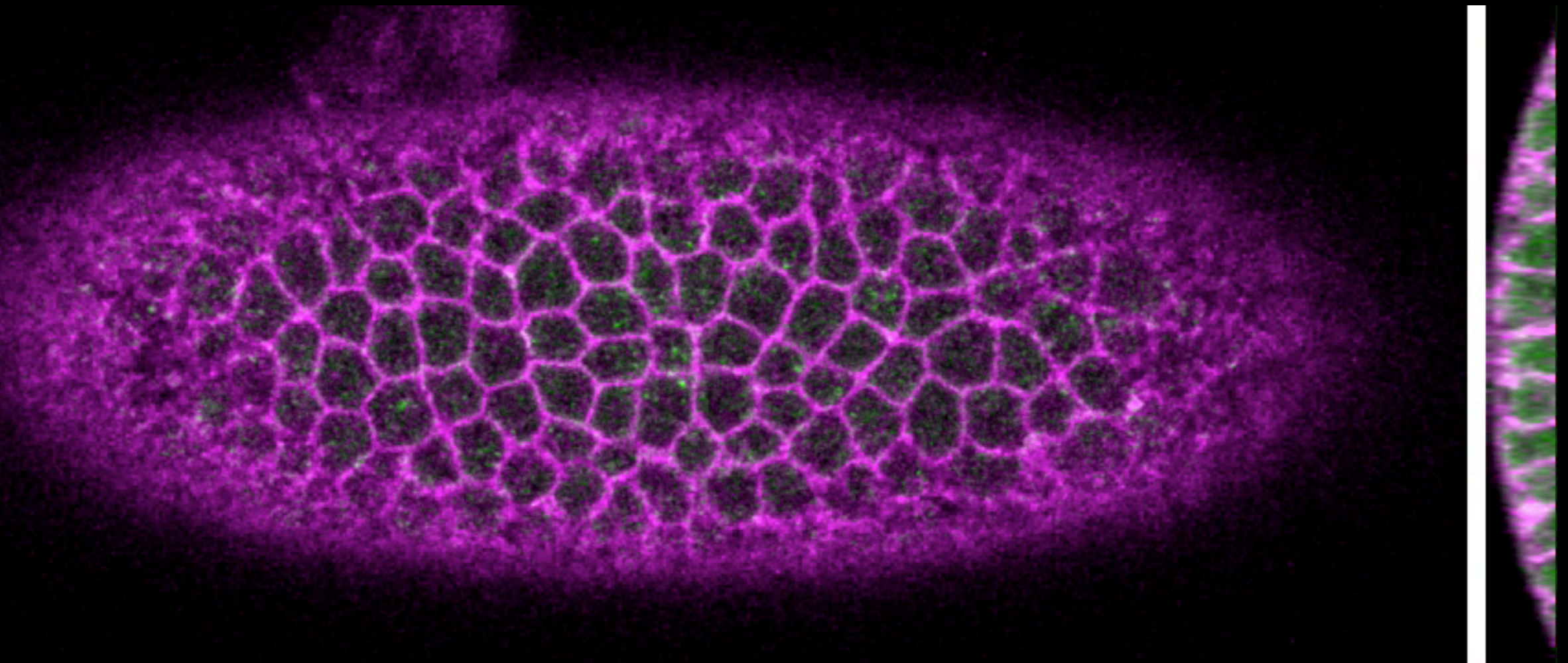


Myosin

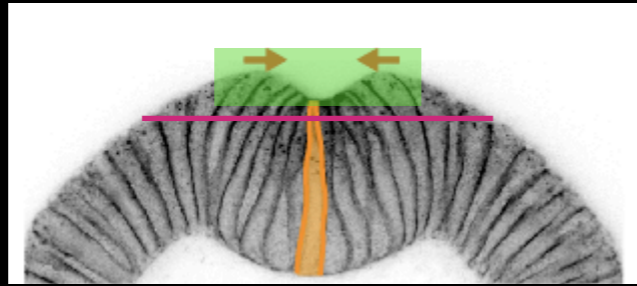
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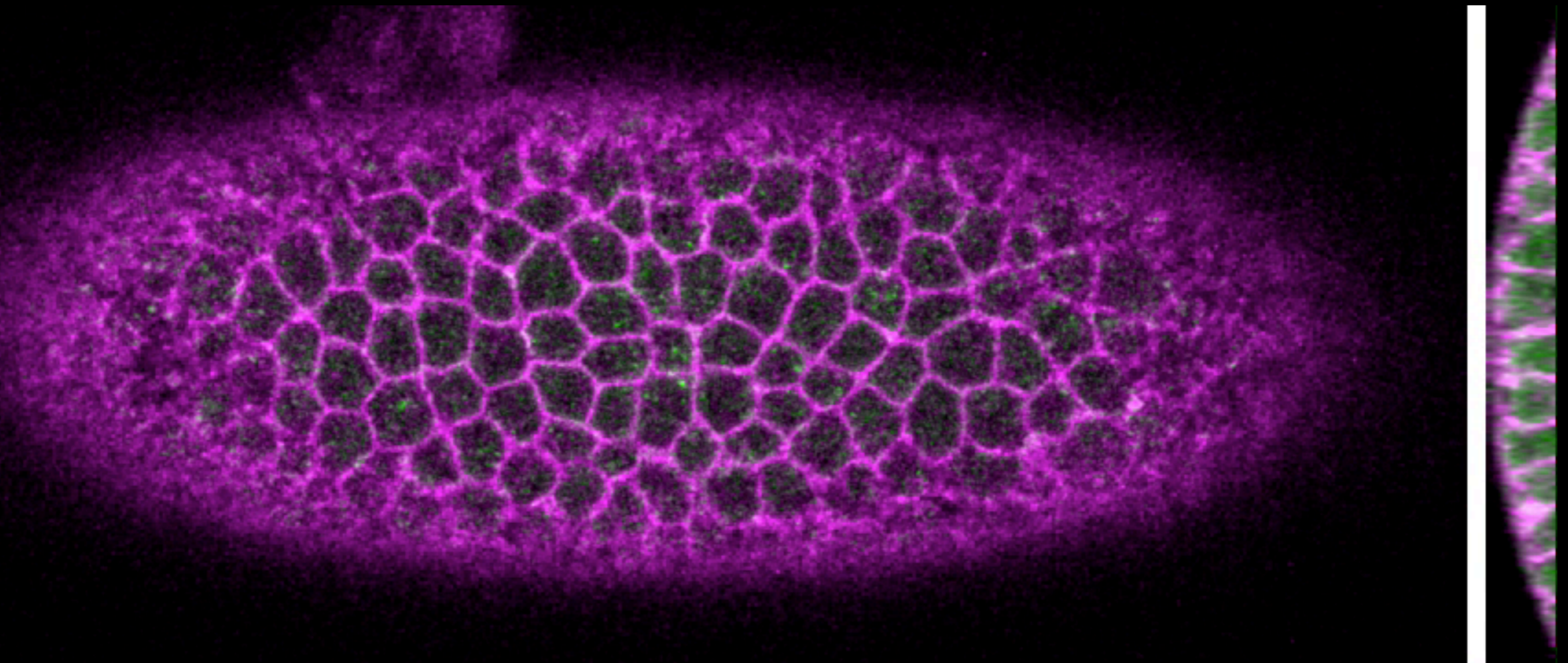


Myosin → Cytoskeleton /Regulator

Membrane → Cell outline

Claudia Vasquez

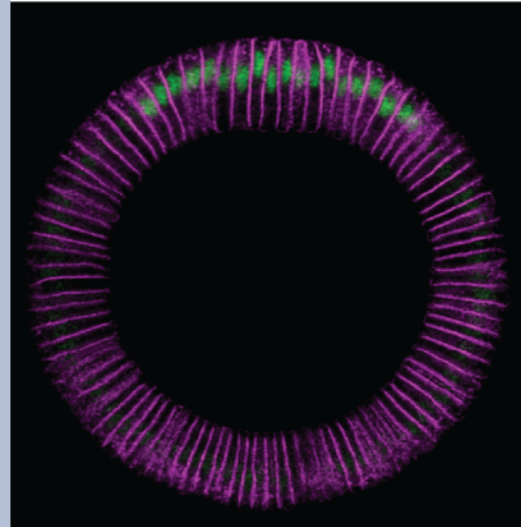
Cross-
Section
Apical → Basal



Ventral transcription factor promotes contractility

Regulation of
gene expression

**Snail and Twist
Transcription
factors**



Ventral

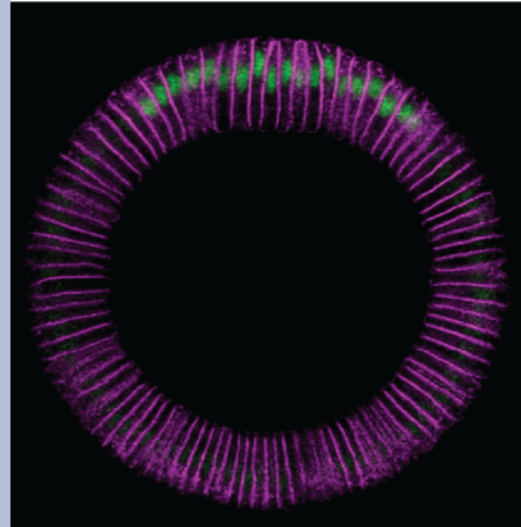


Dorsal

Ventral transcription factor promotes contractility

Regulation of
gene expression

**Snail and Twist
Transcription
factors**



Ventral



Dorsal

Leptin and Grunewald, 1990; Leptin, 1991, Costa et al., 1994.
Barrett et al., 1997., Hacker and Perrimon, 1998.,
Dawes-Hoang et al., 2005., Kolsch et al., 2007., Fox and Peifer, 2007, Homem and Peifer, 2008.

Ventral transcription factor promotes contractility

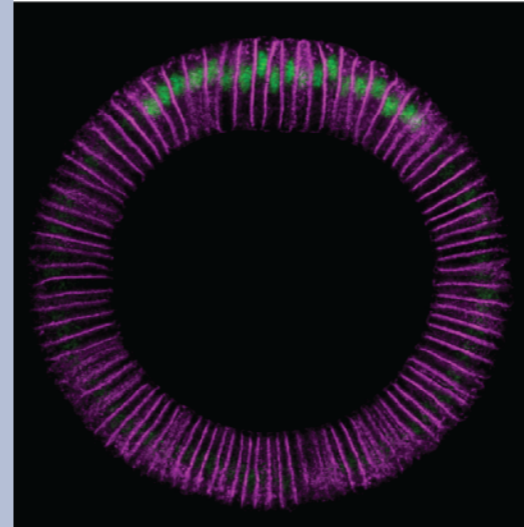
Regulation of
gene expression

**Snail and Twist
Transcription
factors**

Ligand **T48**

GPCR

RhoGEF2



Ventral

Dorsal

Leptin and Grunewald, 1990; Leptin, 1991, Costa et al., 1994.
Barrett et al., 1997., Hacker and Perrimon, 1998.,
Dawes-Hoang et al., 2005., Kolsch et al., 2007., Fox and Peifer, 2007, Homem and Peifer, 2008.

Ventral transcription factor promotes contractility

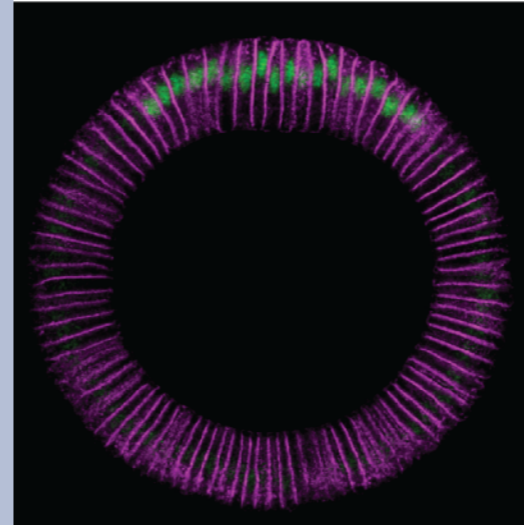
Regulation of
gene expression

**Snail and Twist
Transcription
factors**

Ligand **T48**

GPCR

RhoGEF2



Ventral

Dorsal

Signal transduction
(post-transcriptional)

Ventral transcription factor promotes contractility

Regulation of
gene expression

**Snail and Twist
Transcription
factors**

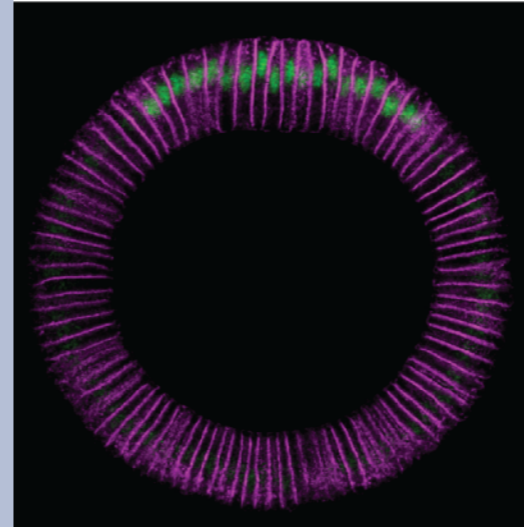
Ligand **T48**

GPCR

RhoGEF2

RhoA
GDP

**RhoA
GTP**



Ventral

Dorsal

Signal transduction
(post-transcriptional)

Ventral transcription factor promotes contractility

Regulation of gene expression

**Snail and Twist
Transcription
factors**

Ligand **T48**

GPCR

RhoGEF2

RhoA
GDP

**RhoA
GTP**

ROCK

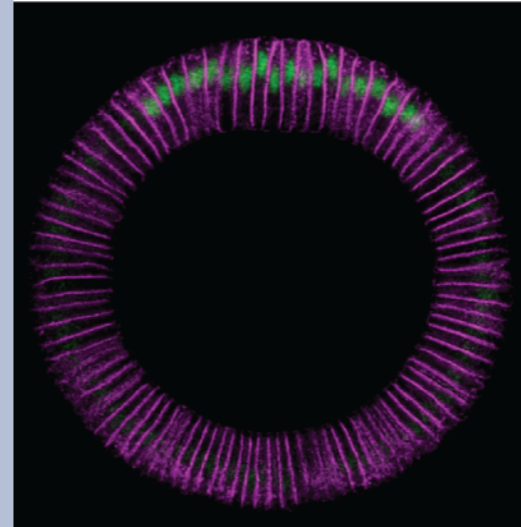
Dia

Myosin
Inactive

**Myosin
Active**

G-actin

F-actin



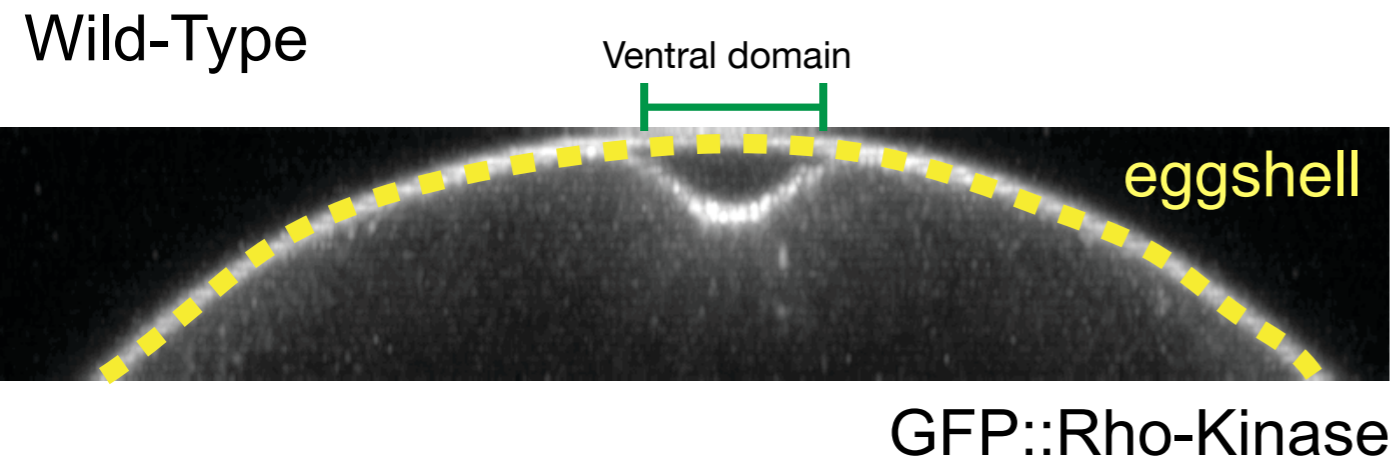
Ventral

Dorsal

Signal transduction
(post-transcriptional)

Leptin and Grunewald, 1990; Leptin, 1991, Costa et al., 1994.
Barrett et al., 1997., Hacker and Perrimon, 1998.,
Dawes-Hoang et al., 2005., Kolsch et al., 2007., Fox and Peifer, 2007, Homem and Peifer, 2008.

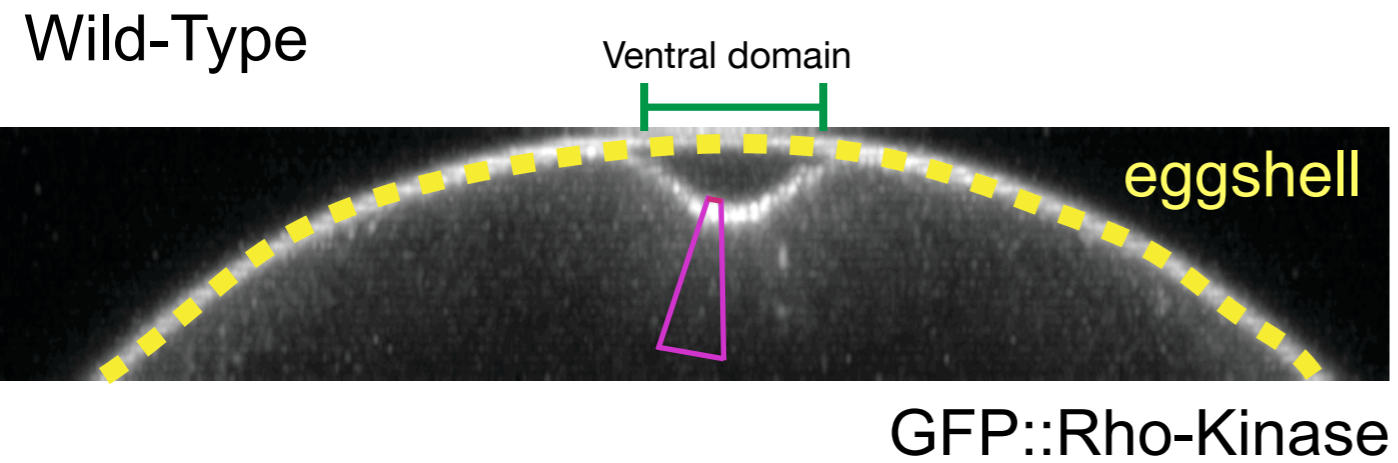
ROCK is polarized within the apical domain



Frank Mason, Ph.D.

Detailed description: A small, square portrait of a man with short brown hair, glasses, and a beard, wearing a light-colored checkered shirt. He is smiling slightly. The background is a plain, light color.

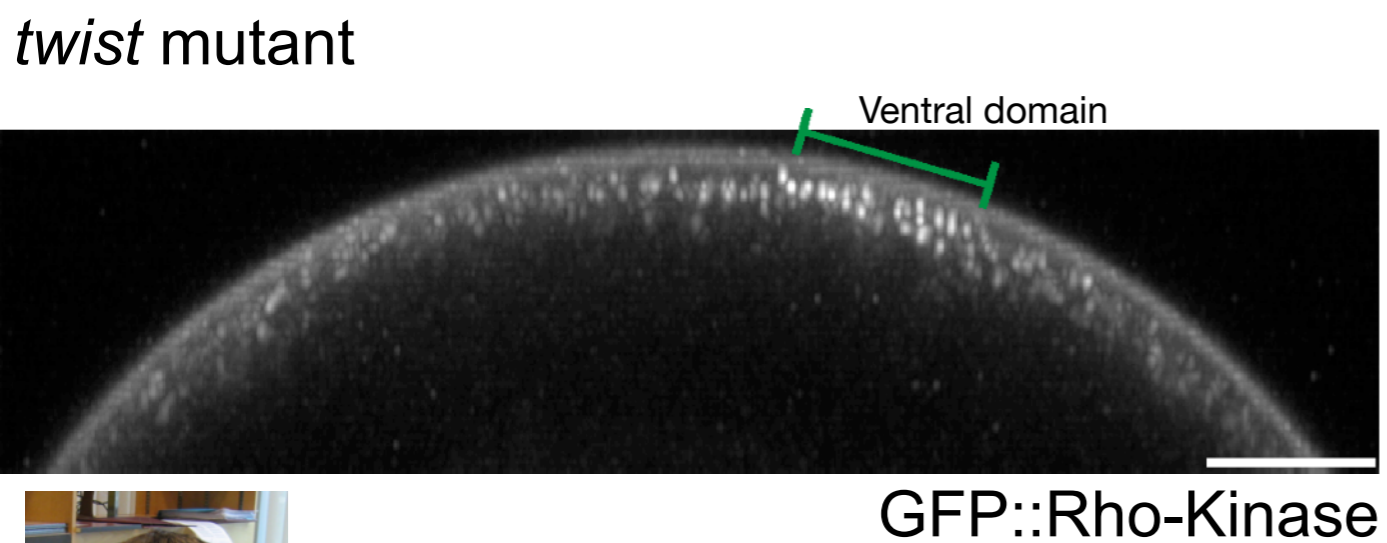
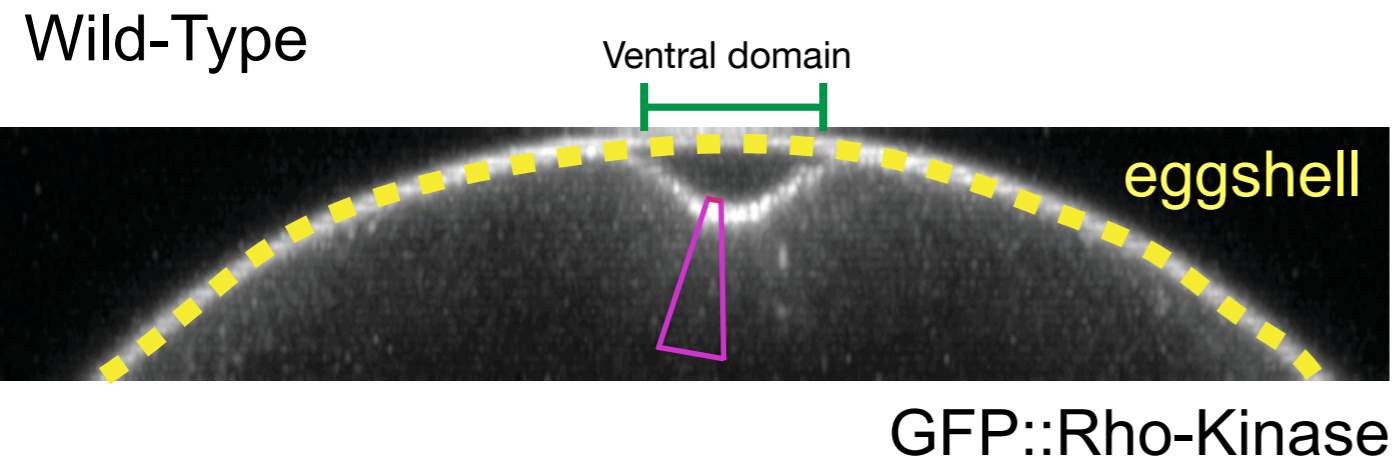
ROCK is polarized within the apical domain



Frank Mason, Ph.D.

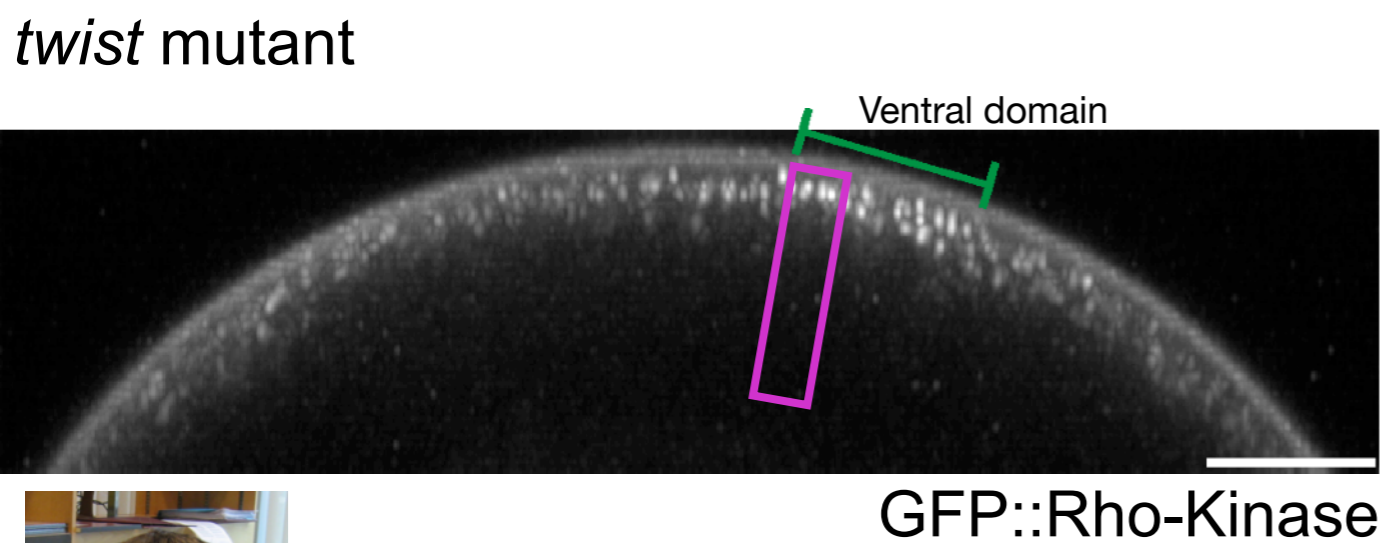
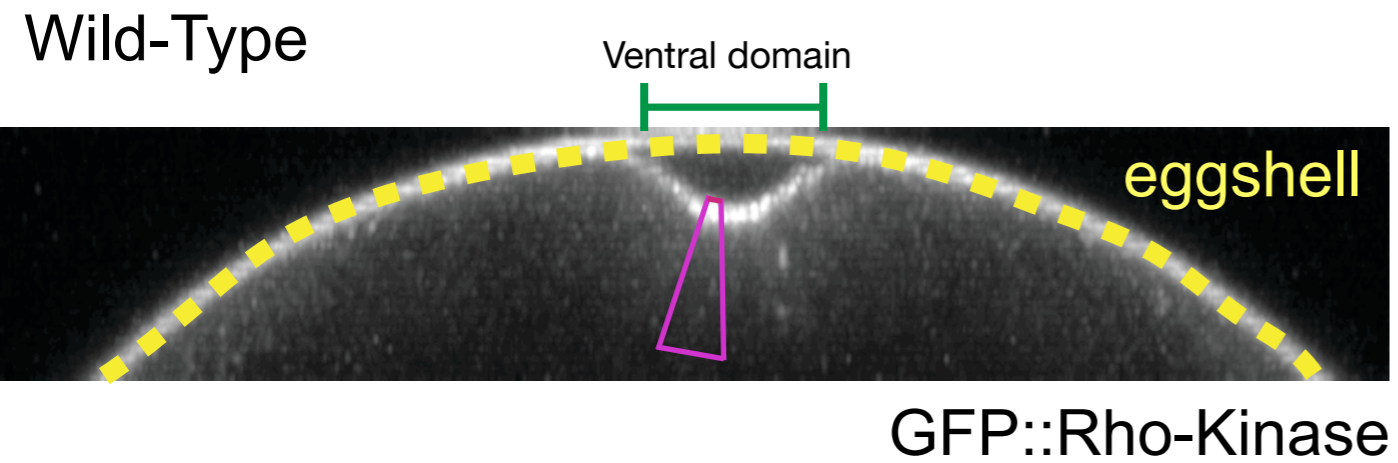
Detailed description: A small portrait of Frank Mason, Ph.D., is located in the bottom left corner of the slide. He is a man with glasses and a beard, wearing a light-colored shirt.

ROCK is polarized within the apical domain



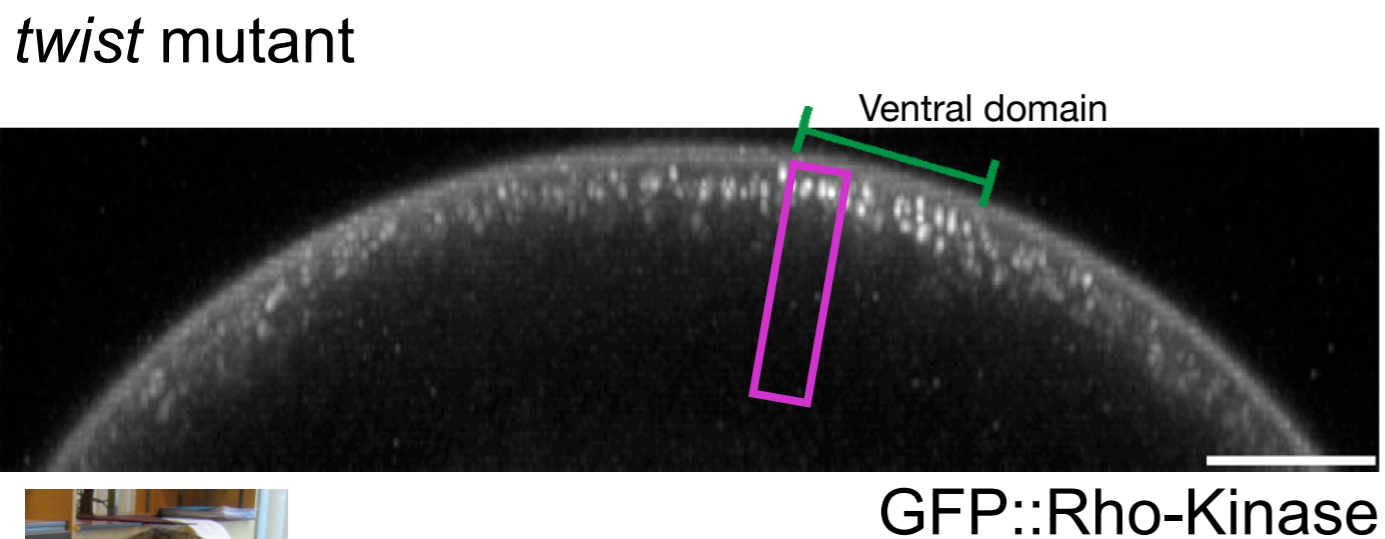
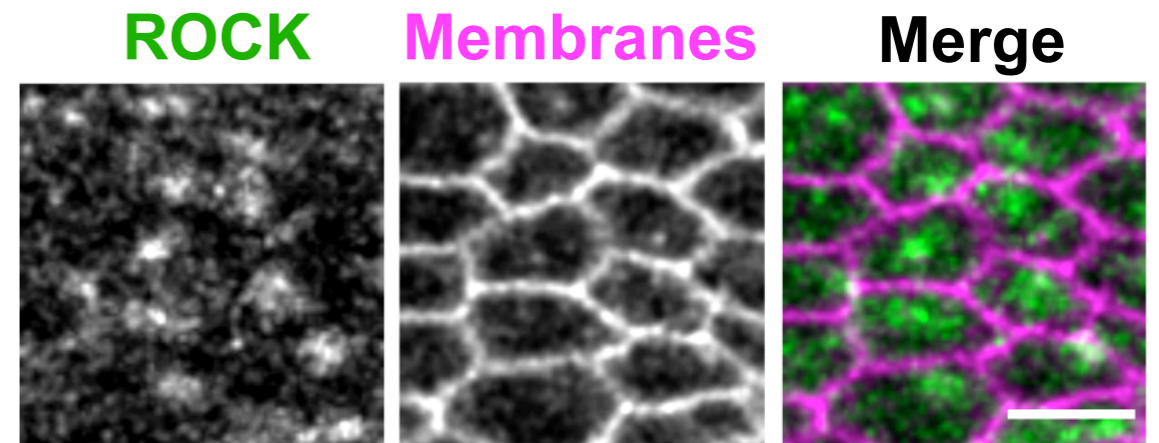
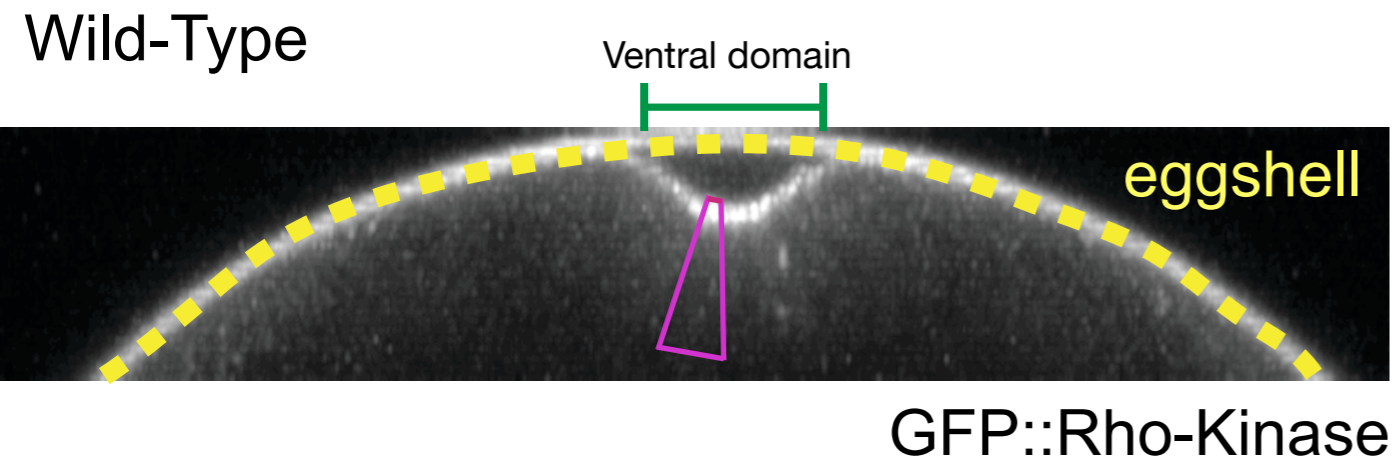
Frank Mason, Ph.D.

ROCK is polarized within the apical domain



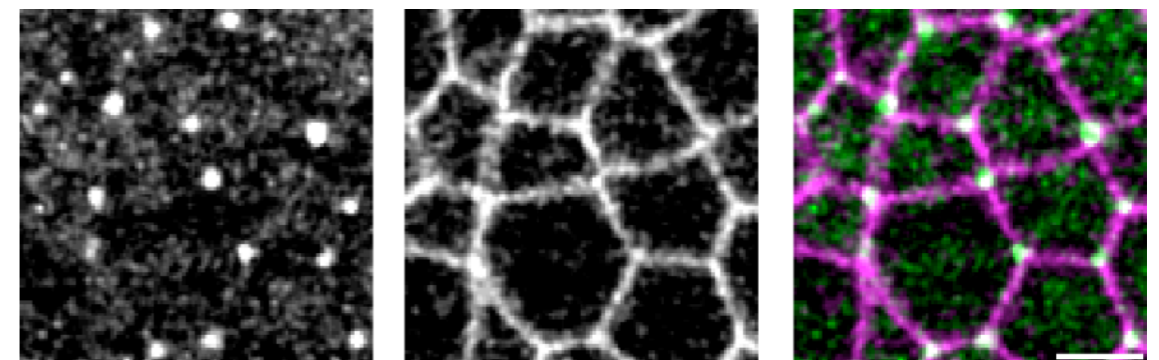
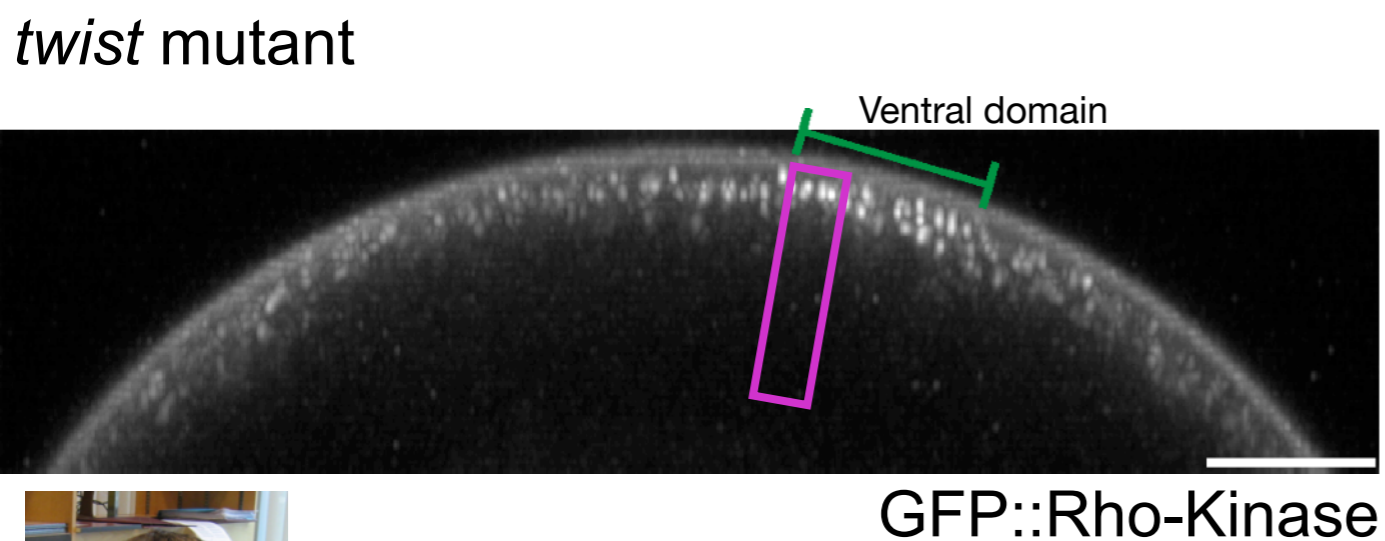
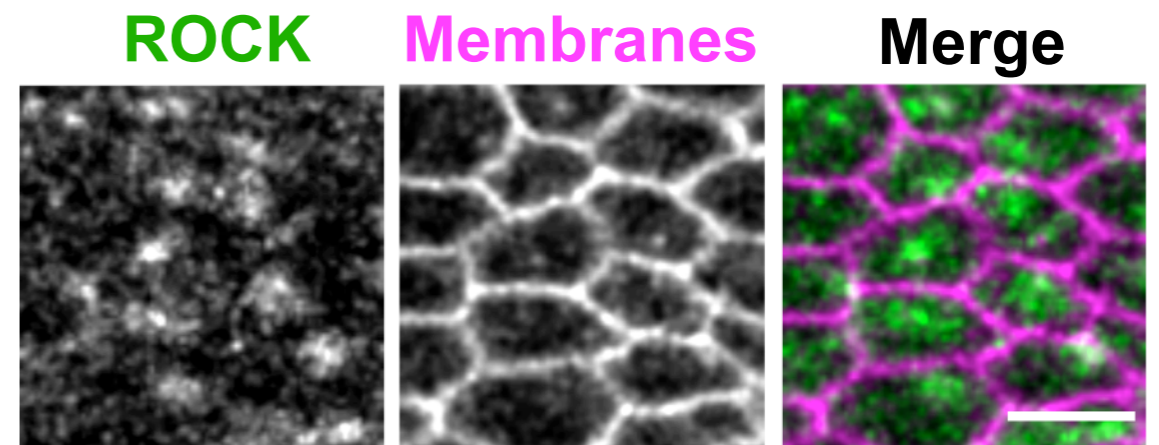
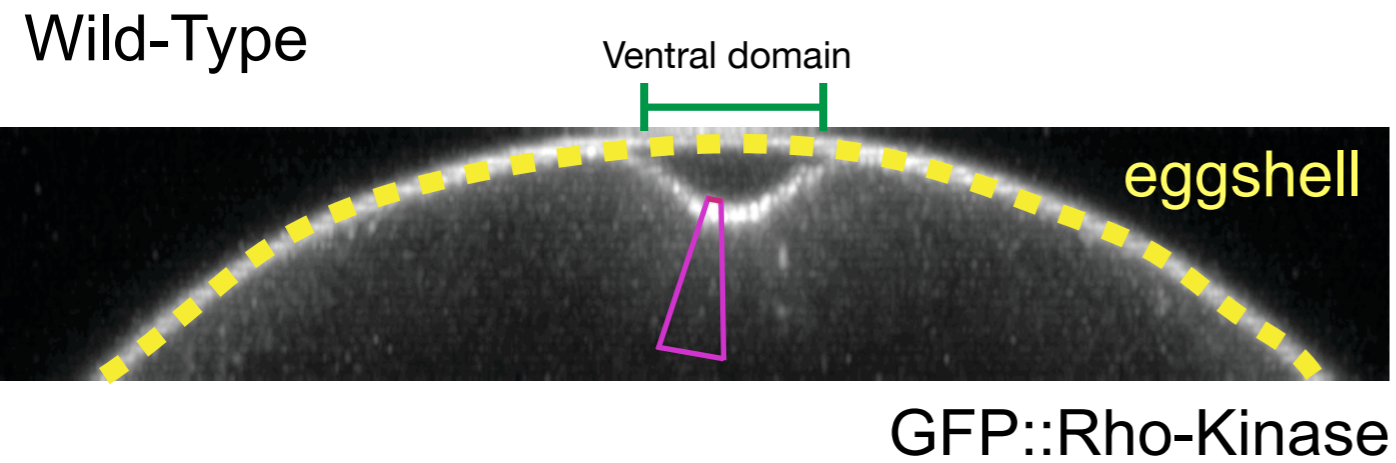
Frank Mason, Ph.D.

ROCK is polarized within the apical domain



Frank Mason, Ph.D.

ROCK is polarized within the apical domain

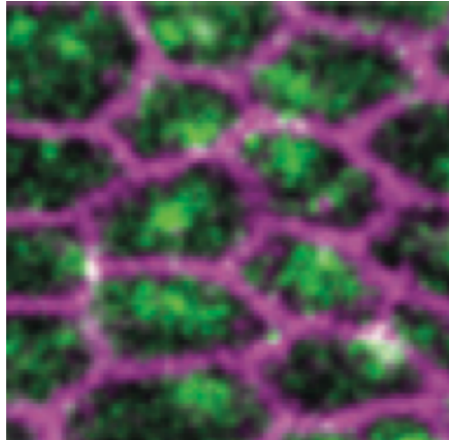


Frank Mason, Ph.D.

RhoA signaling is polarized within apical domain

GFP::ROCK

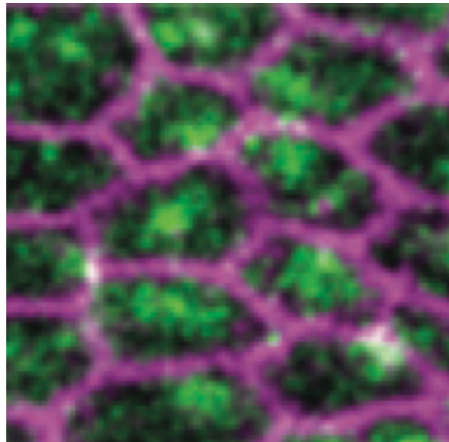
Surface view



RhoA signaling is polarized within apical domain

GFP::ROCK

Surface view

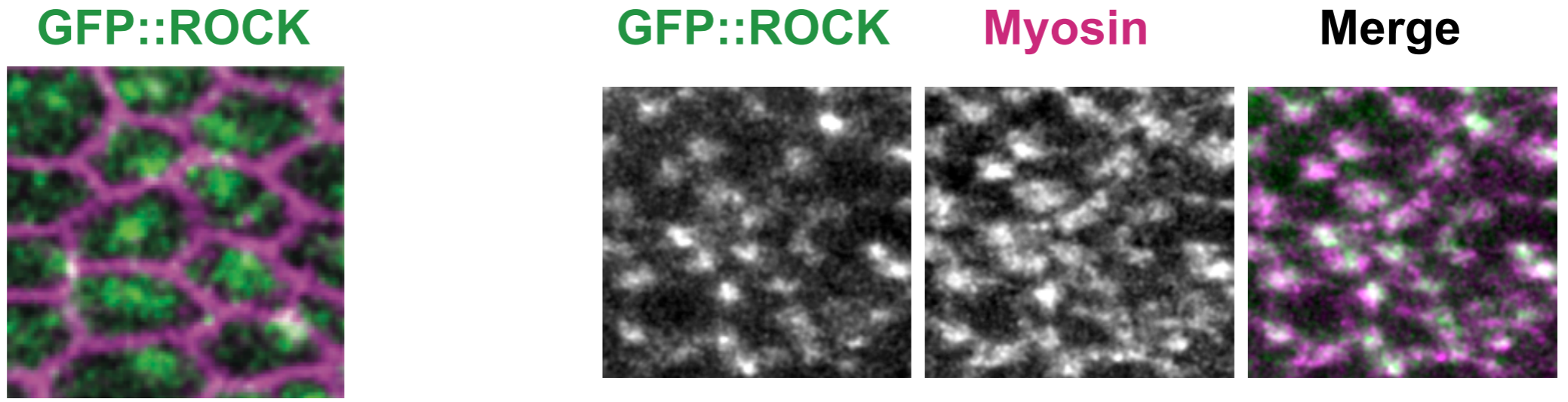


Depends on RhoA activity

Active RhoA is also medioapically polarized

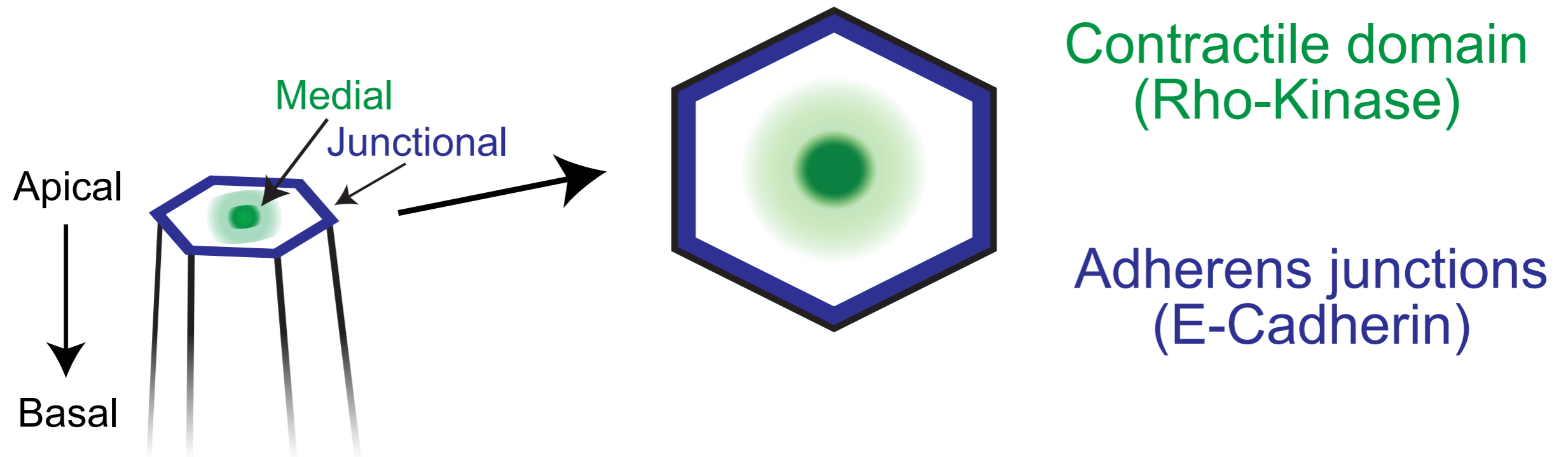
RhoA signaling is polarized within apical domain

Surface view



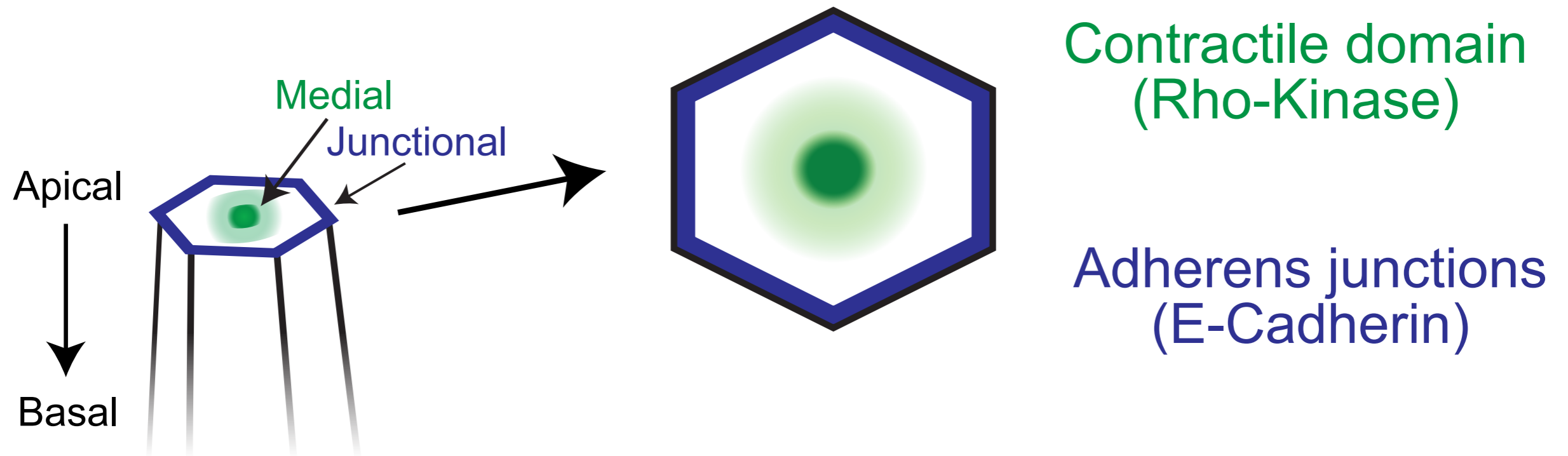
Depends on RhoA activity
Active RhoA is also medioapically polarized

Constricting cell apex has a spatial pattern of ROCK/myosin



Mason FM, Tworoger M, Martin AC. *Nat Cell Biol.* (2013).
Mason et al., *J. Cell Biol.* (2016).

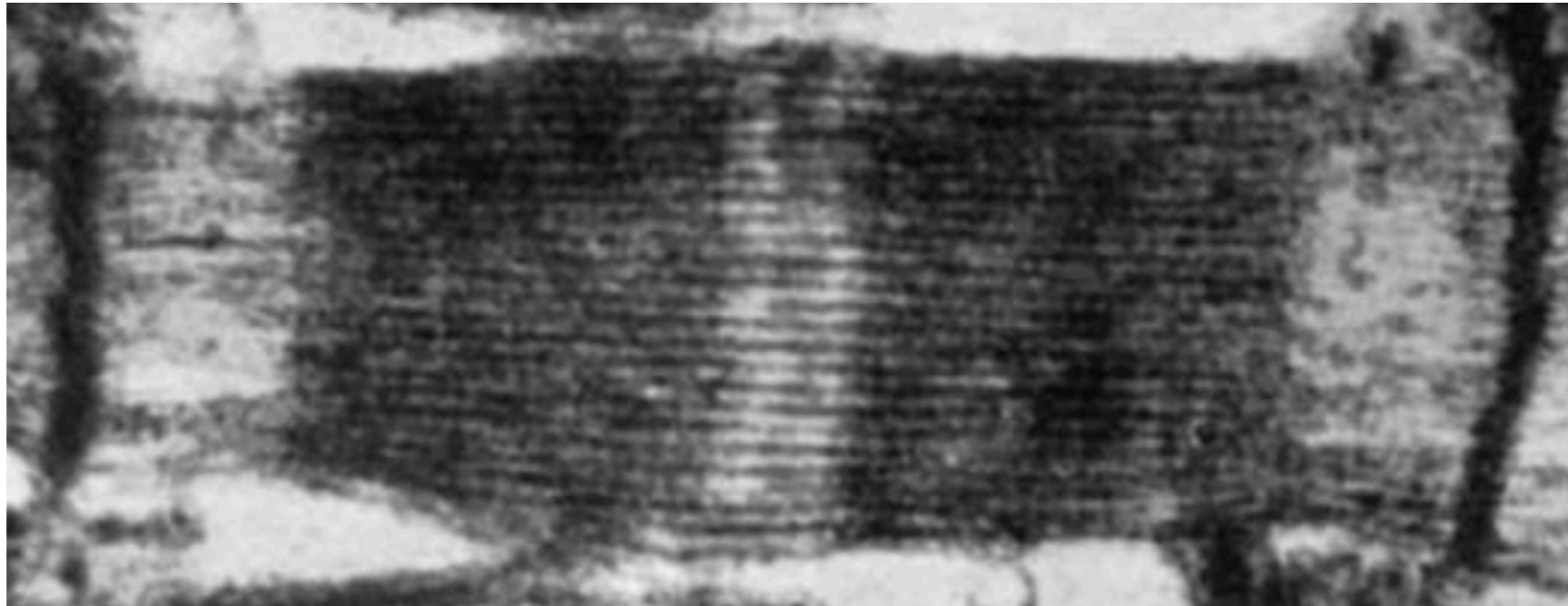
Constricting cell apex has a spatial pattern of ROCK/myosin



Why is ROCK/myosin in the middle?

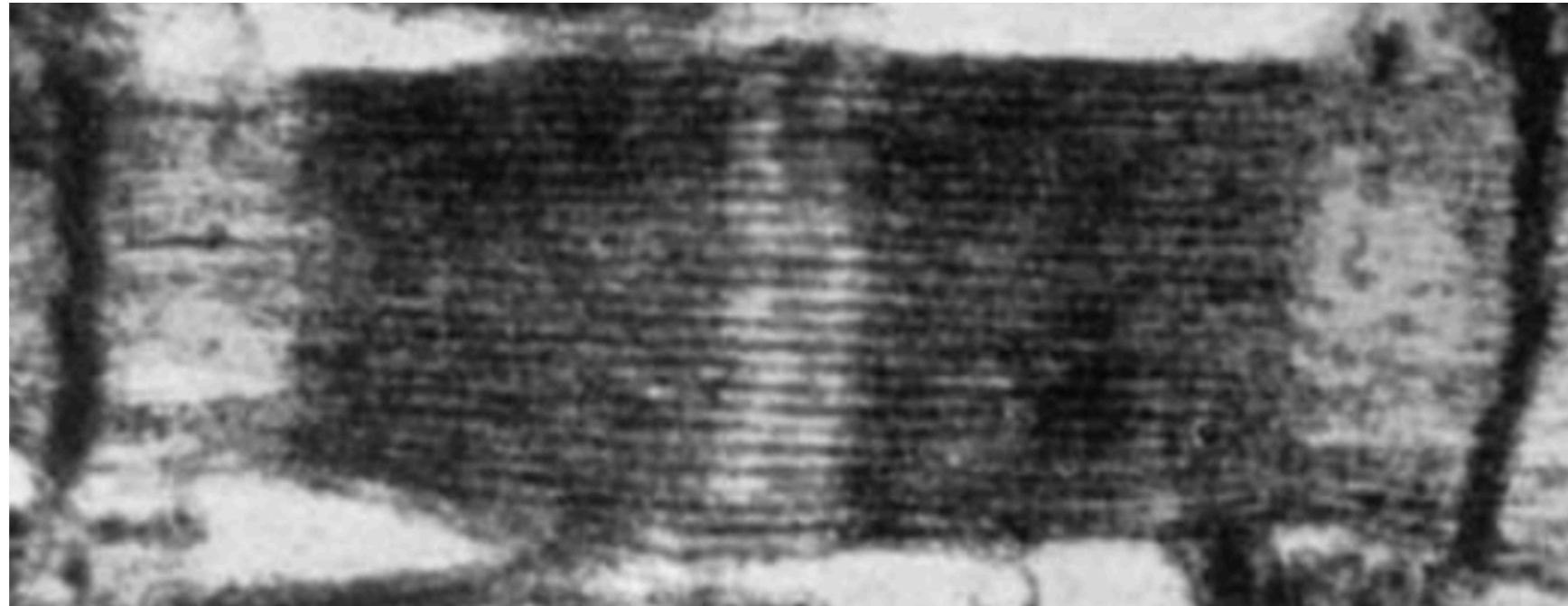
Mason FM, Tworoger M, Martin AC. *Nat Cell Biol.* (2013).
Mason et al., *J. Cell Biol.* (2016).

Sarcomere is the contractile unit of a muscle - myosin in the middle

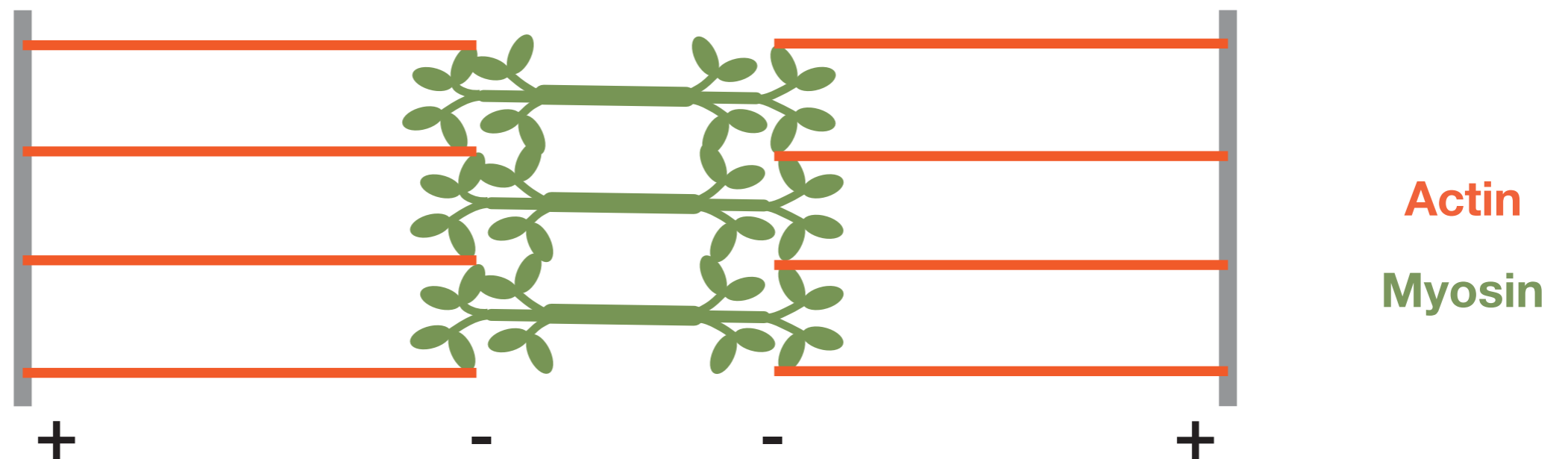


Hanson and Huxley, *Nature* (1953).

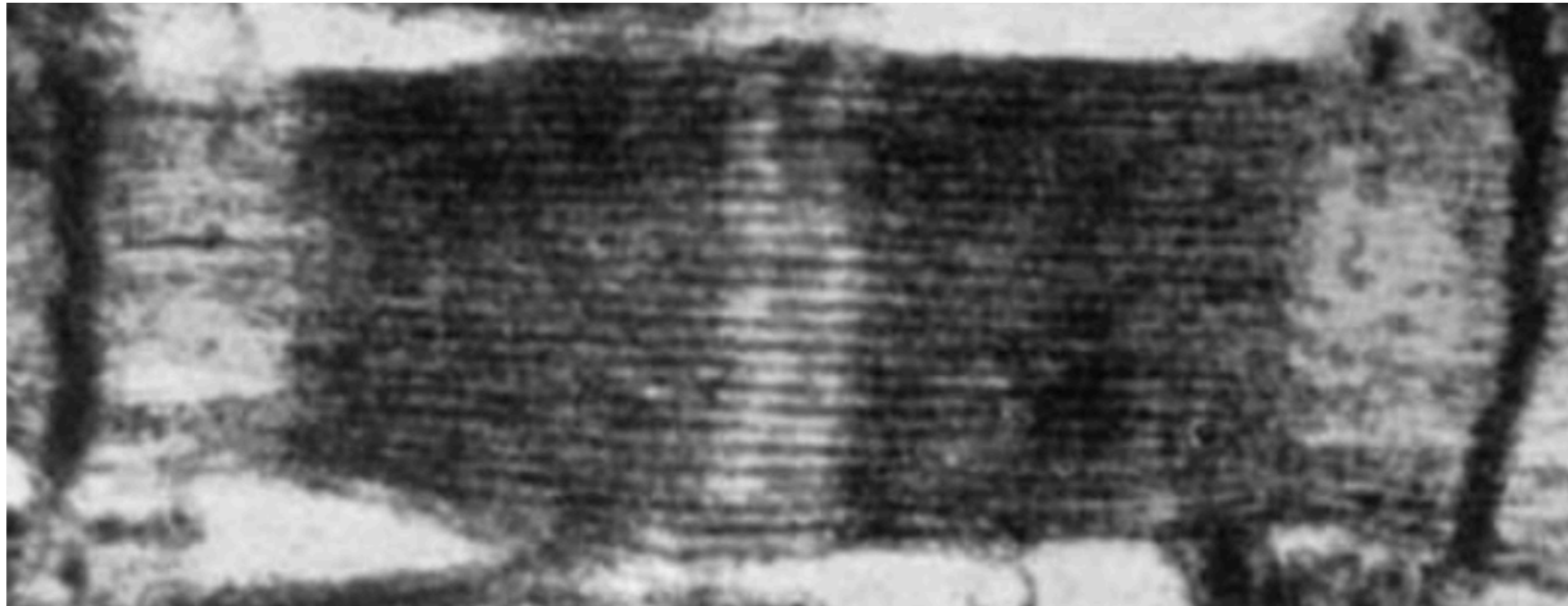
Sarcomere is the contractile unit of a muscle - myosin in the middle



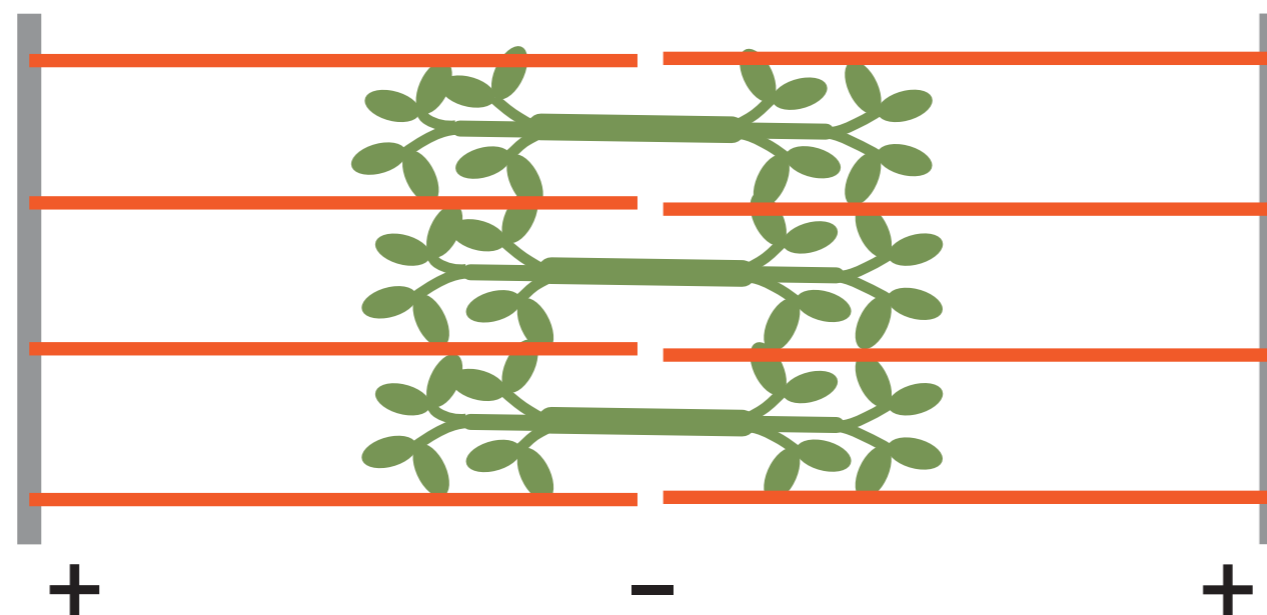
Hanson and Huxley, *Nature* (1953).



Sarcomere is the contractile unit of a muscle - myosin in the middle



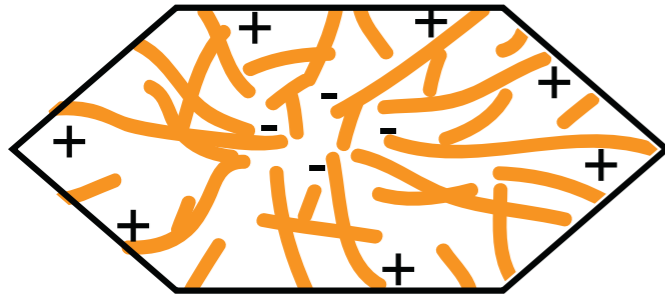
Hanson and Huxley, *Nature* (1953).



Actin
Myosin

Apical cortex polarity resembles that of sarcomere

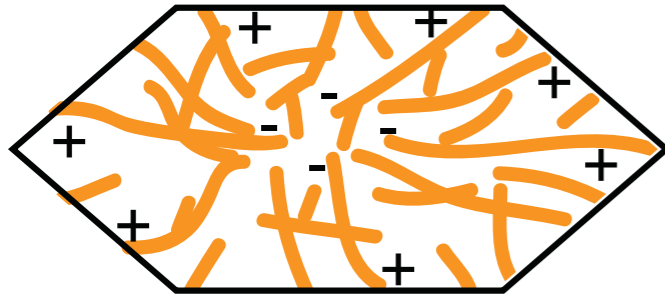
Polarized **actin**
cortex



Jonathan
Coravos, Ph.D.

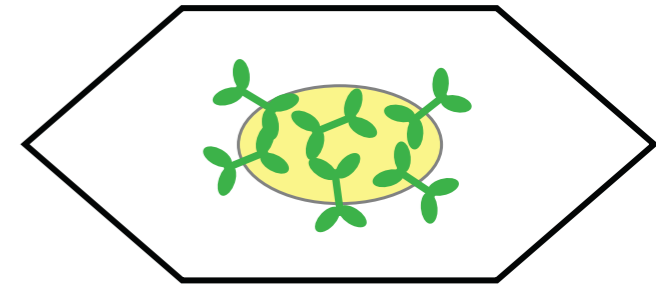
Apical cortex polarity resembles that of sarcomere

Polarized **actin**
cortex



Jonathan
Coravos, Ph.D.

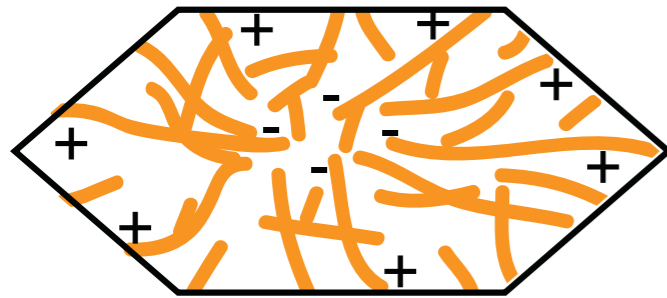
Central **ROCK** and
myosin II



Frank
Mason, Ph.D.

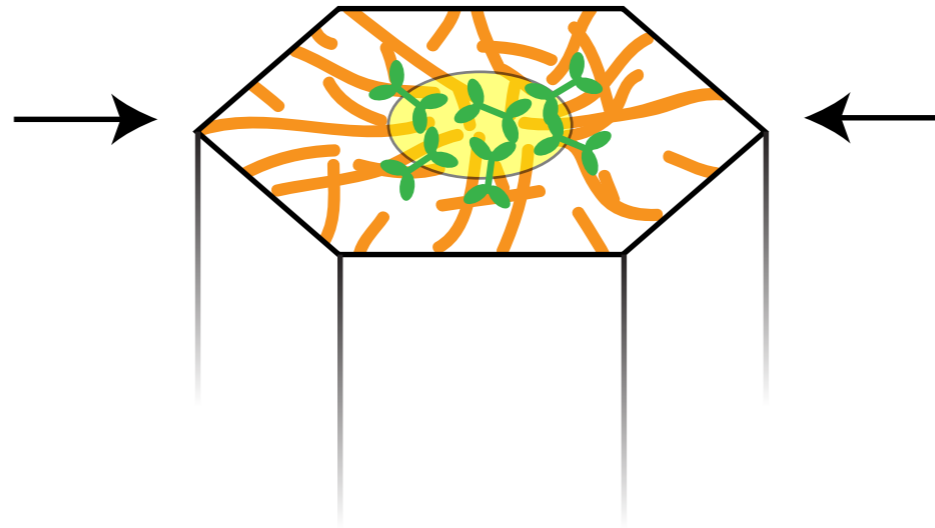
Apical cortex polarity resembles that of sarcomere

Polarized **actin**
cortex

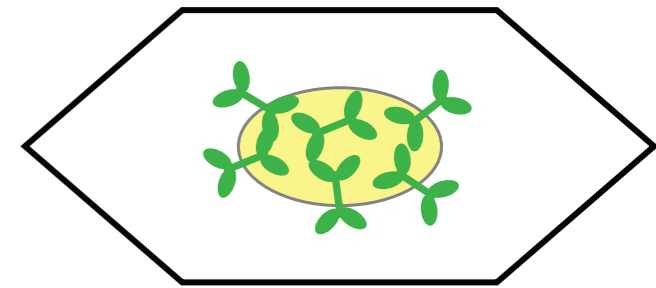


Jonathan
Coravos, Ph.D.

“Radial Sarcomere”



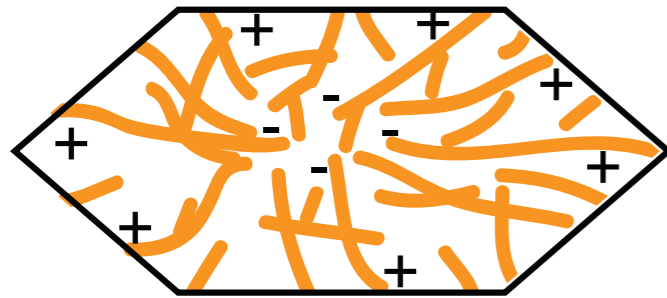
Central **ROCK** and
myosin II



Frank
Mason, Ph.D.

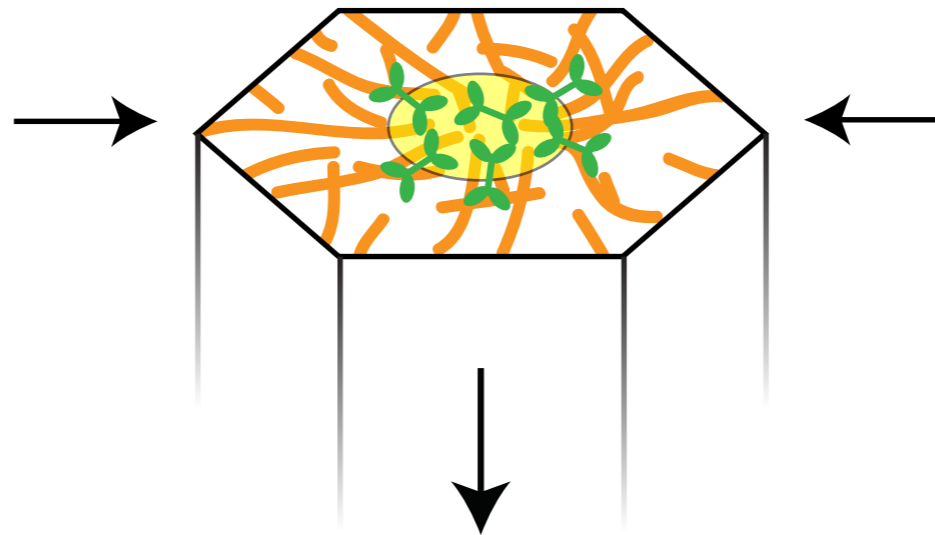
Apical cortex polarity resembles that of sarcomere

Polarized **actin**
cortex



Jonathan
Coravos, Ph.D.

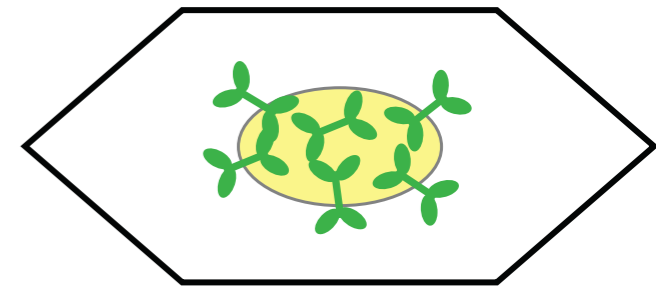
“Radial Sarcomere”



Apical **Constriction**

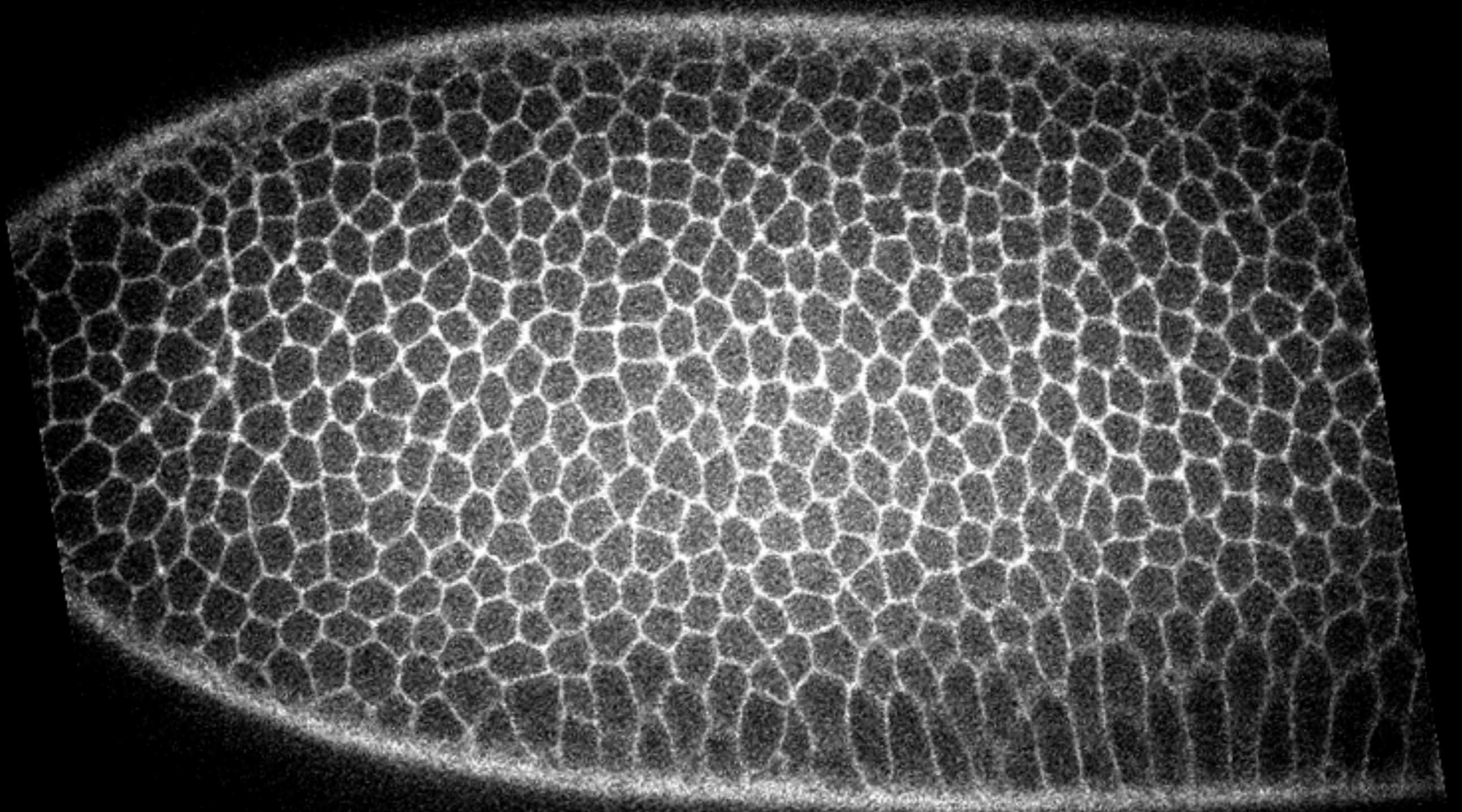


Central **ROCK** and
myosin II

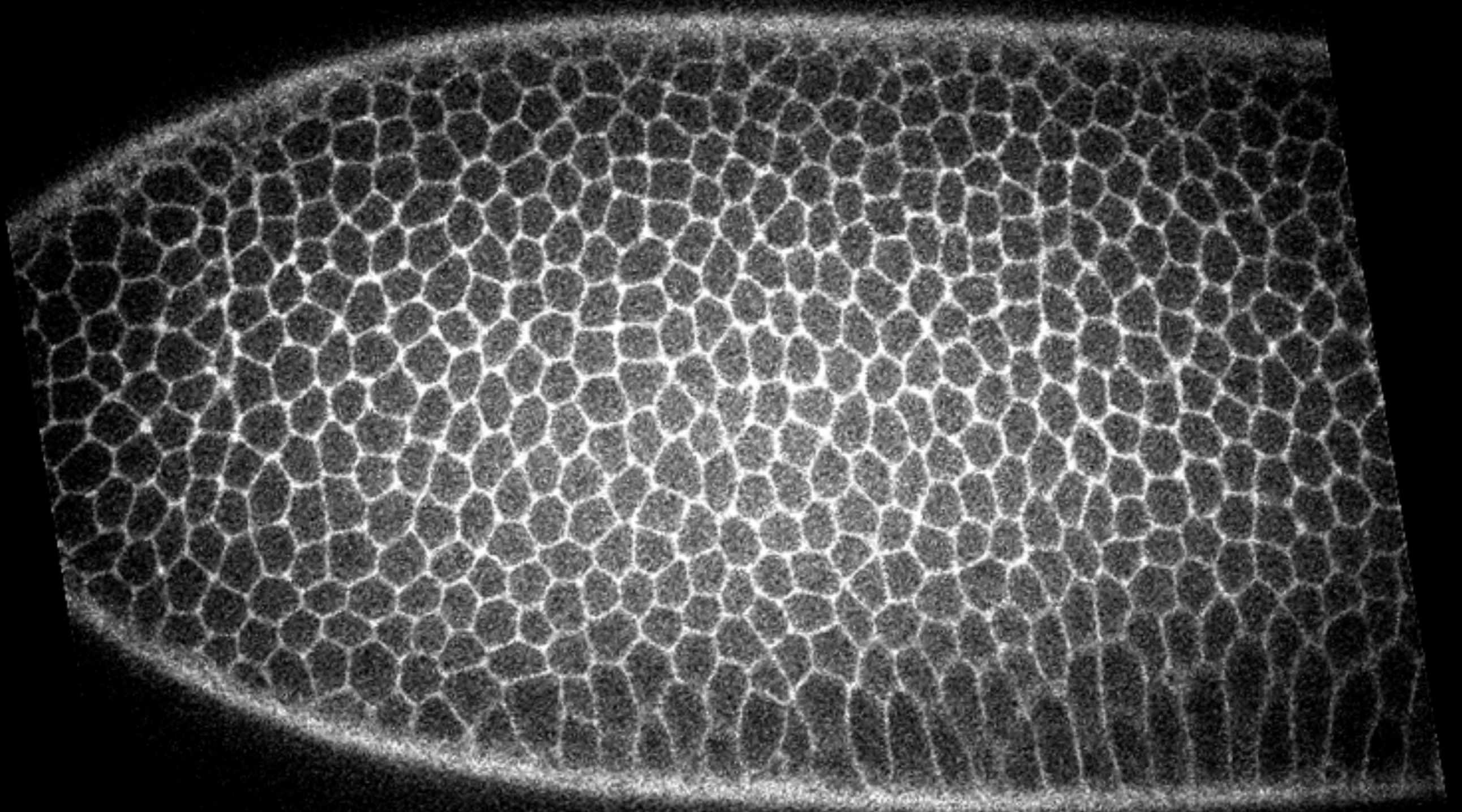


Frank
Mason, Ph.D.

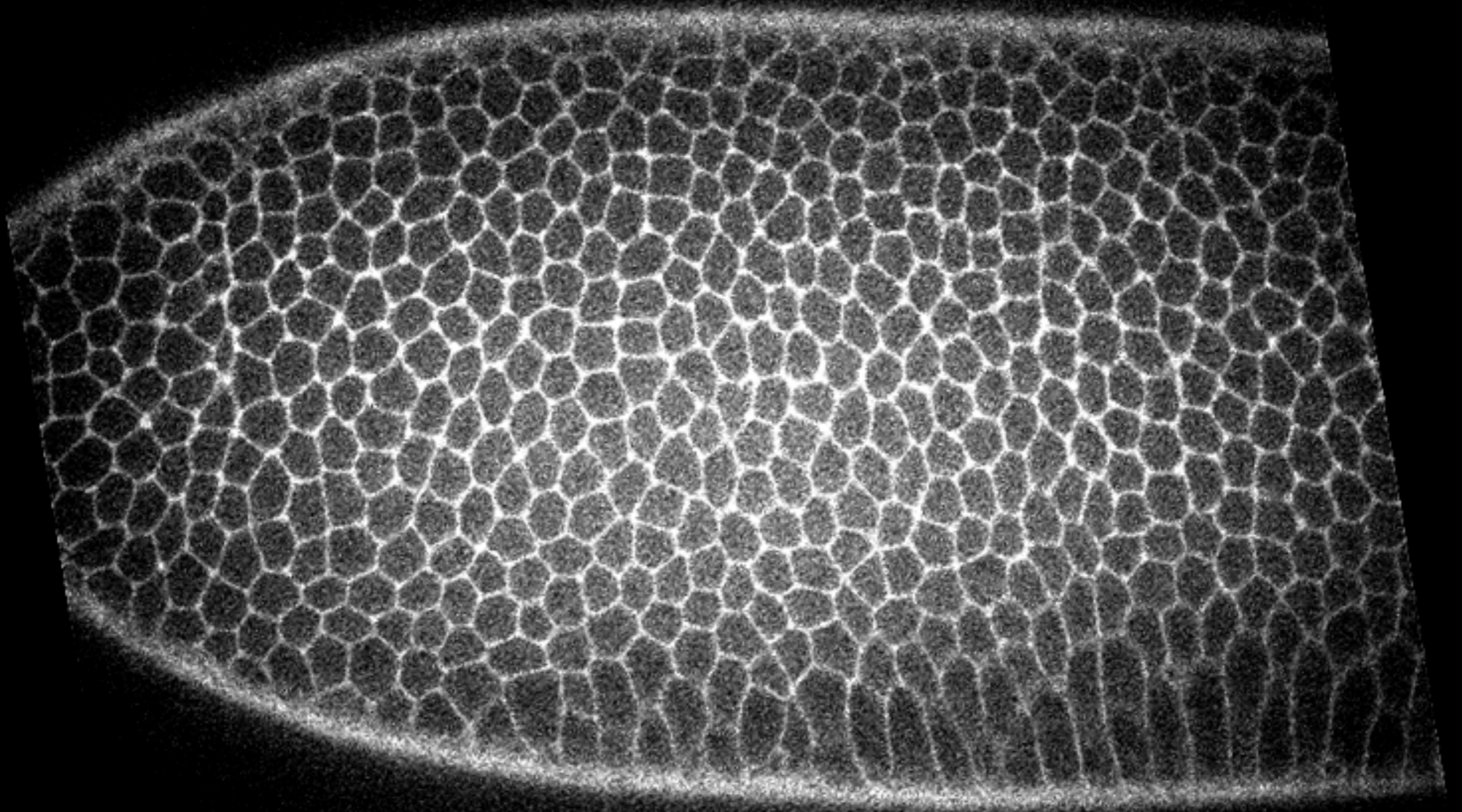
Tissue extension in Drosophila



Tissue extension in Drosophila

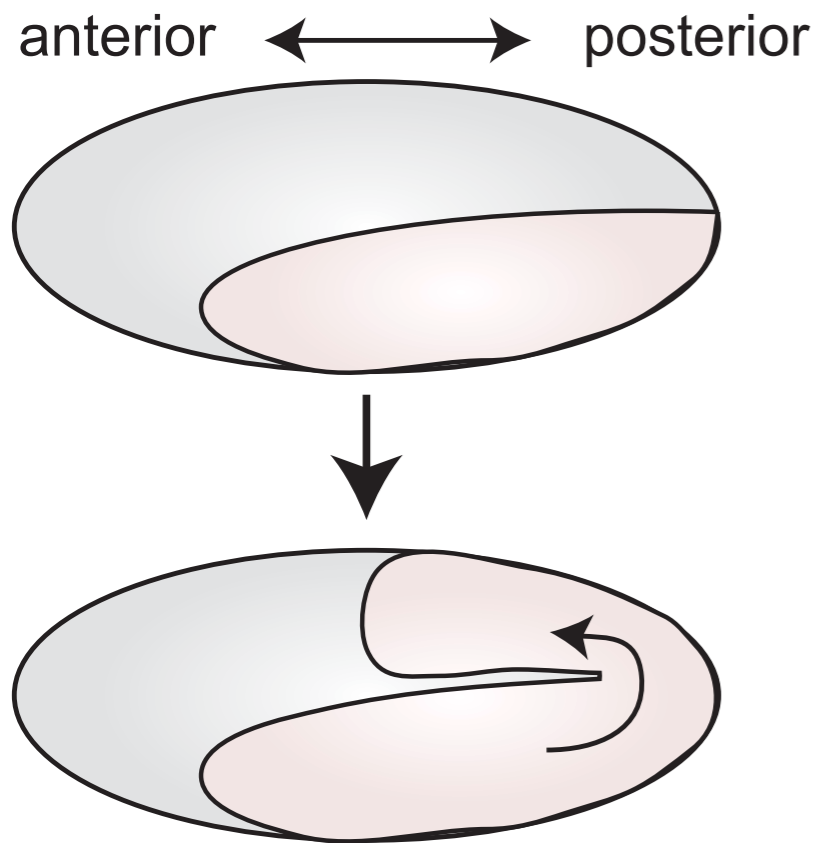


Tissue extension in Drosophila



Ordered exchange of intercellular contacts can drive extension

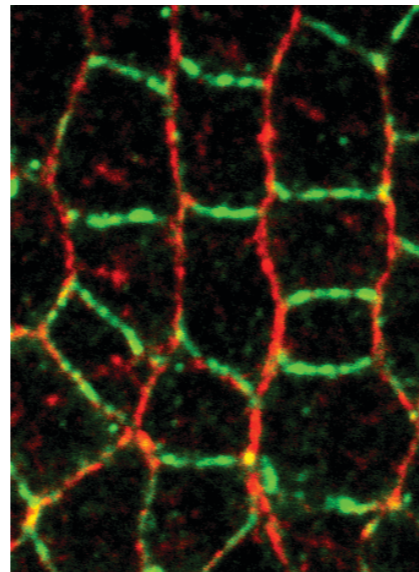
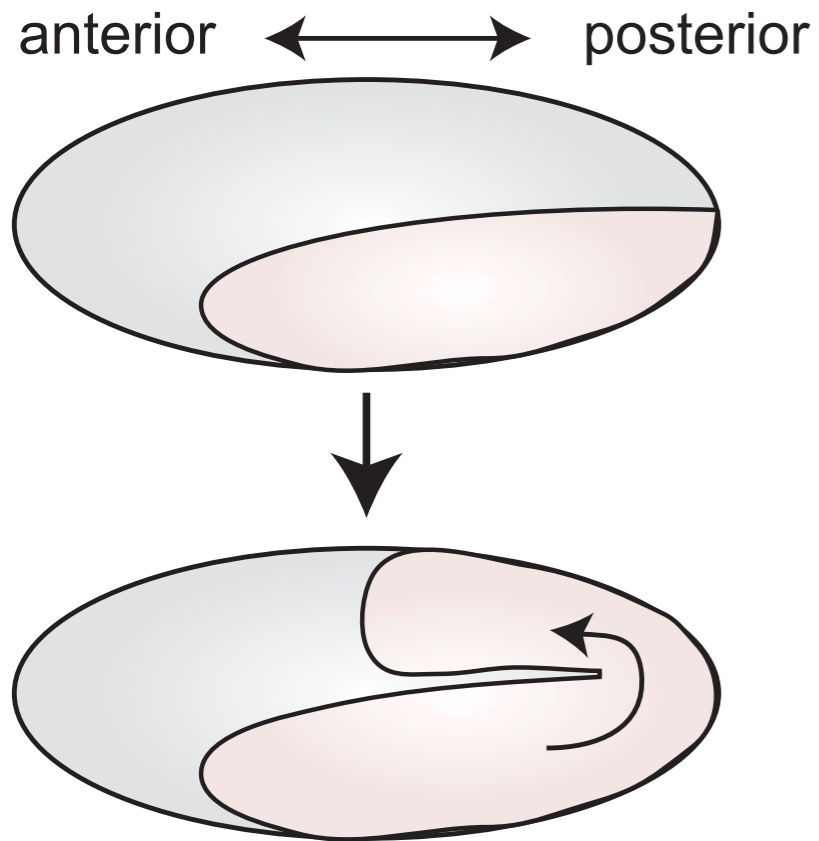
1



Mason & Martin, 2011

Ordered exchange of intercellular contacts can drive extension

1

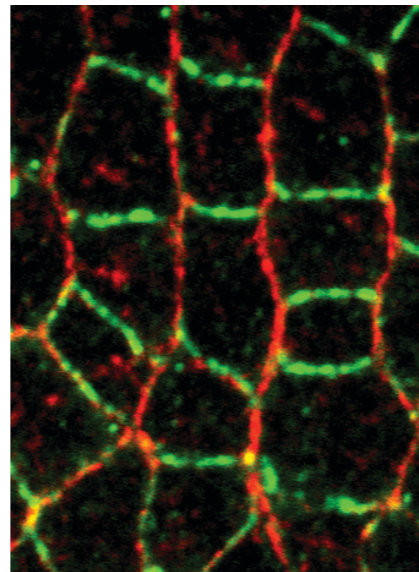
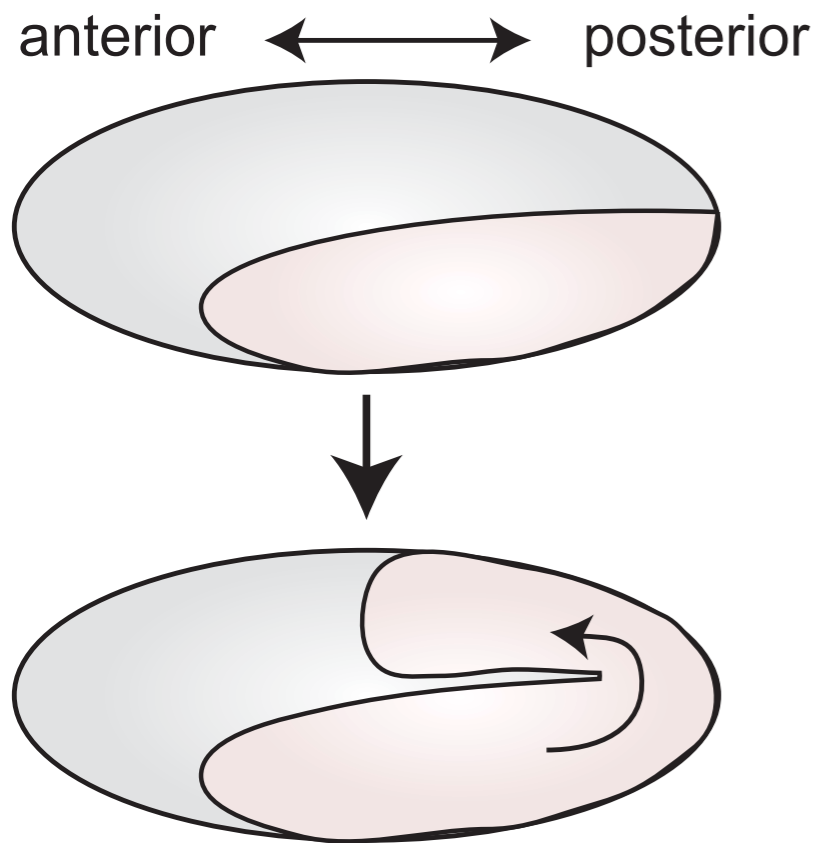


Myo-II

Baz/Par-3

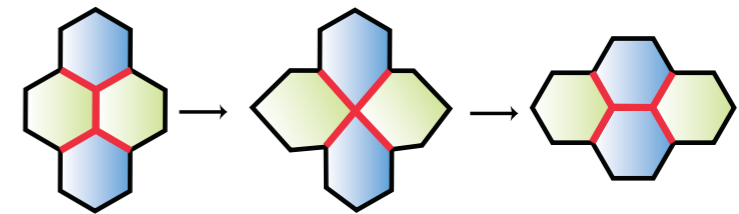
Mason & Martin, 2011

Ordered exchange of intercellular contacts can drive extension

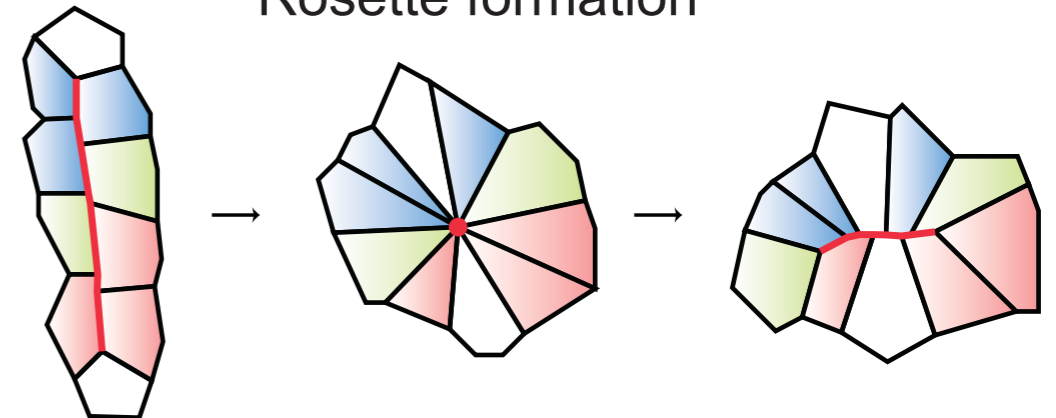


Myo-II
Baz/Par-3

Type 1 transition



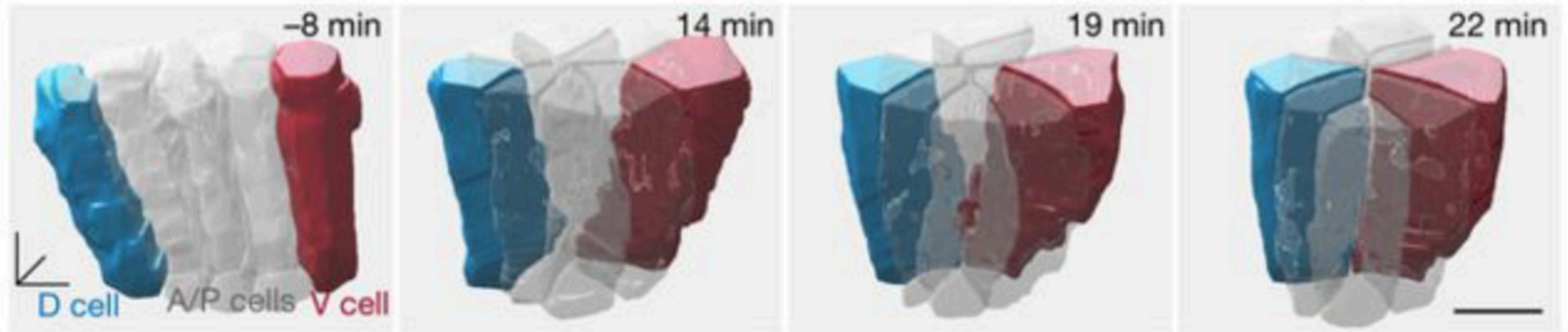
Rosette formation



anterior \longleftrightarrow posterior

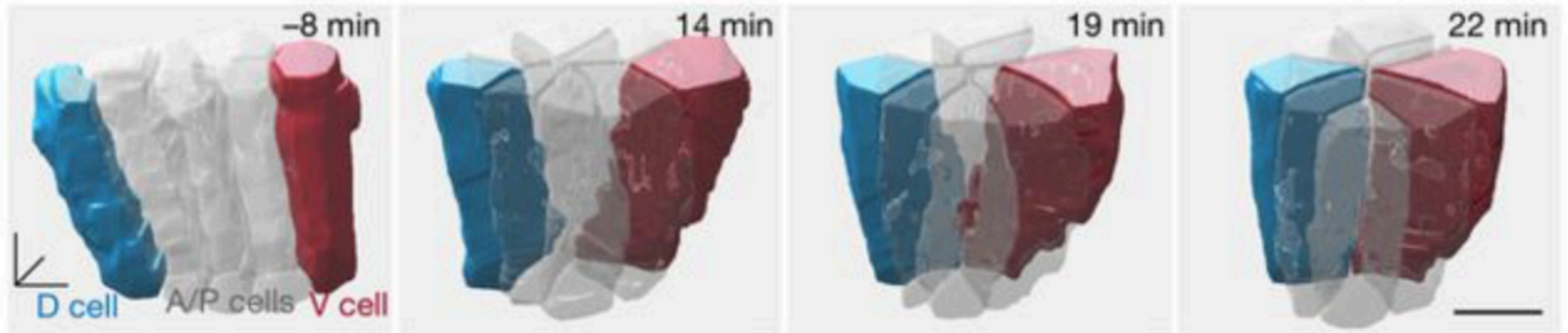
Mason & Martin, 2011

Cell protrusion is also important for convergent extension

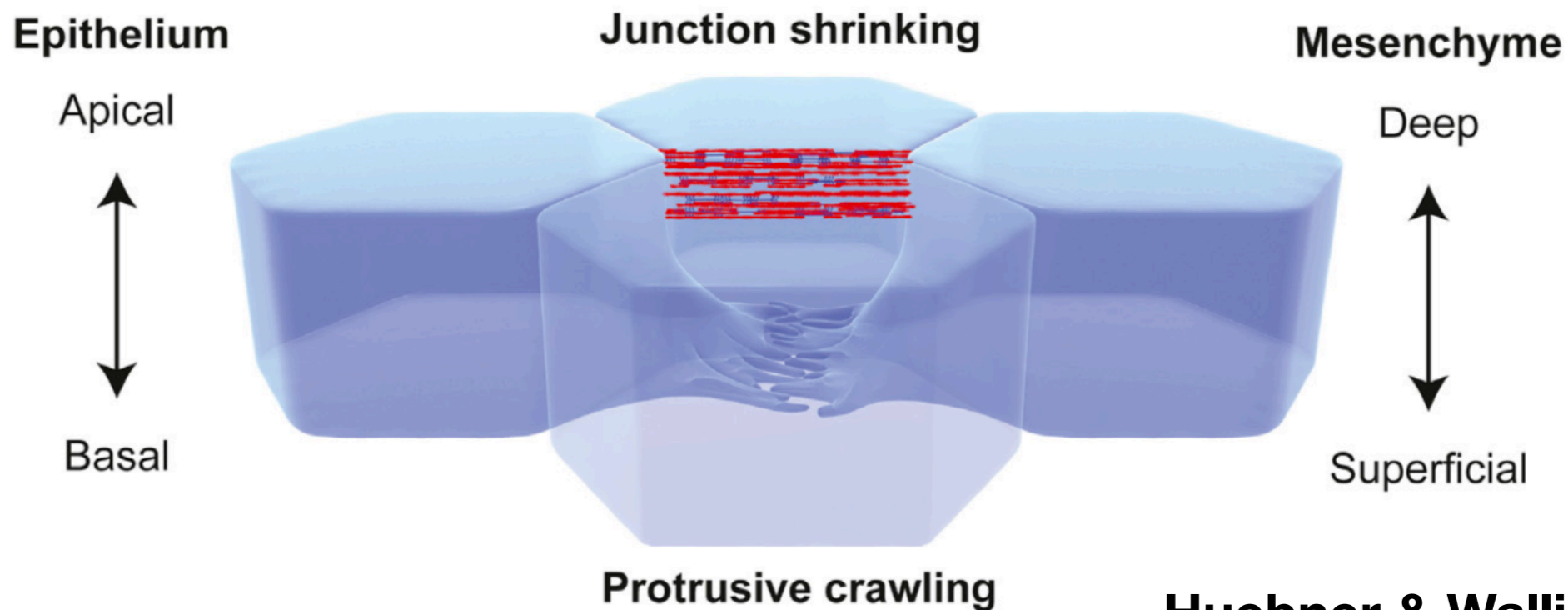


Sun et al., 2017

Cell protrusion is also important for convergent extension



Sun et al., 2017



Huebner & Wallingford, 2018

How do we know whether myosin functions as a 'motor'?

ATPase Activity of Myosin Correlated with Speed of Muscle Shortening

MICHAEL BÁRÁNY

1967

From the Institute for Muscle Disease, Inc., New York

How do we know whether myosin functions as a 'motor'?

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Claudia Vasquez



ATPase rate scales with muscle contraction velocity

TABLE II
RELATIONSHIP BETWEEN CONTRACTION TIME AND
ATPASE ACTIVITY OF MYOSIN IN MUSCLES OF CAT AND SLOTH

Muscle	Contraction time*	ATPase activity‡ in the presence of				ATP sensitivity‡
		Actin	Ca ⁺⁺ +0.05 M KCl	Ca ⁺⁺ +0.5 M KCl	EDTA	
		$\mu\text{moles } P_i / \text{mg/min}$	$\mu\text{mole } P_i / \text{mg/min}$	$\mu\text{mole } P_i / \text{mg/min}$	$\mu\text{mole } P_i / \text{mg/min}$	
	<i>msec</i>					%
Cat extensor digitorum longus	19–19.5	1.46	0.67	0.45	0.61	131
Sloth extensor digitorum longus . .	122–135	0.26	0.18	0.12	0.17	100
Cat gastrocnemius medialis	22.5–27	1.41	0.68	0.39	0.58	128
Sloth gastrocnemius medialis	109	0.25	0.20	0.12	0.19	122

* 37–38°C and 34–35°C for muscles of cat and sloth, respectively. References for contraction times: cat extensor digitorum longus, Gordon and Phillips (13, 14); cat gastrocnemius, Wills (15) and Buller et al. (11); sloth muscles, Goffart et al. (12).

‡ 37.5°C and 34.5°C for myosin of muscles of cat and sloth, respectively.

ATPase rate scales with muscle contraction velocity



TABLE II

CONTRACTION TIME AND
IN MUSCLES OF CAT AND SLOTH

		ATPase activity‡ in the presence of				ATP sensitivity‡
		Actin	Ca ⁺⁺ +0.05	Ca ⁺⁺ +0.5	EDTA	
			m KCl	m KCl		
		$\mu\text{moles } P_i / \text{mg/min}$	$\mu\text{mole } P_i / \text{mg/min}$	$\mu\text{mole } P_i / \text{mg/min}$	$\mu\text{mole } P_i / \text{mg/min}$	%
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ATPase rate scales with muscle contraction velocity



TABLE II

CONTRACTION TIME AND
IN MUSCLES OF CAT AND SLOTH

	ATPase activity [†] in the presence of			ATP sensitivity [†]
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[†] 37.5°C and 34.5°C for myosin of muscles of cat and sloth, respectively.

ATPase rate scales with muscle contraction velocity



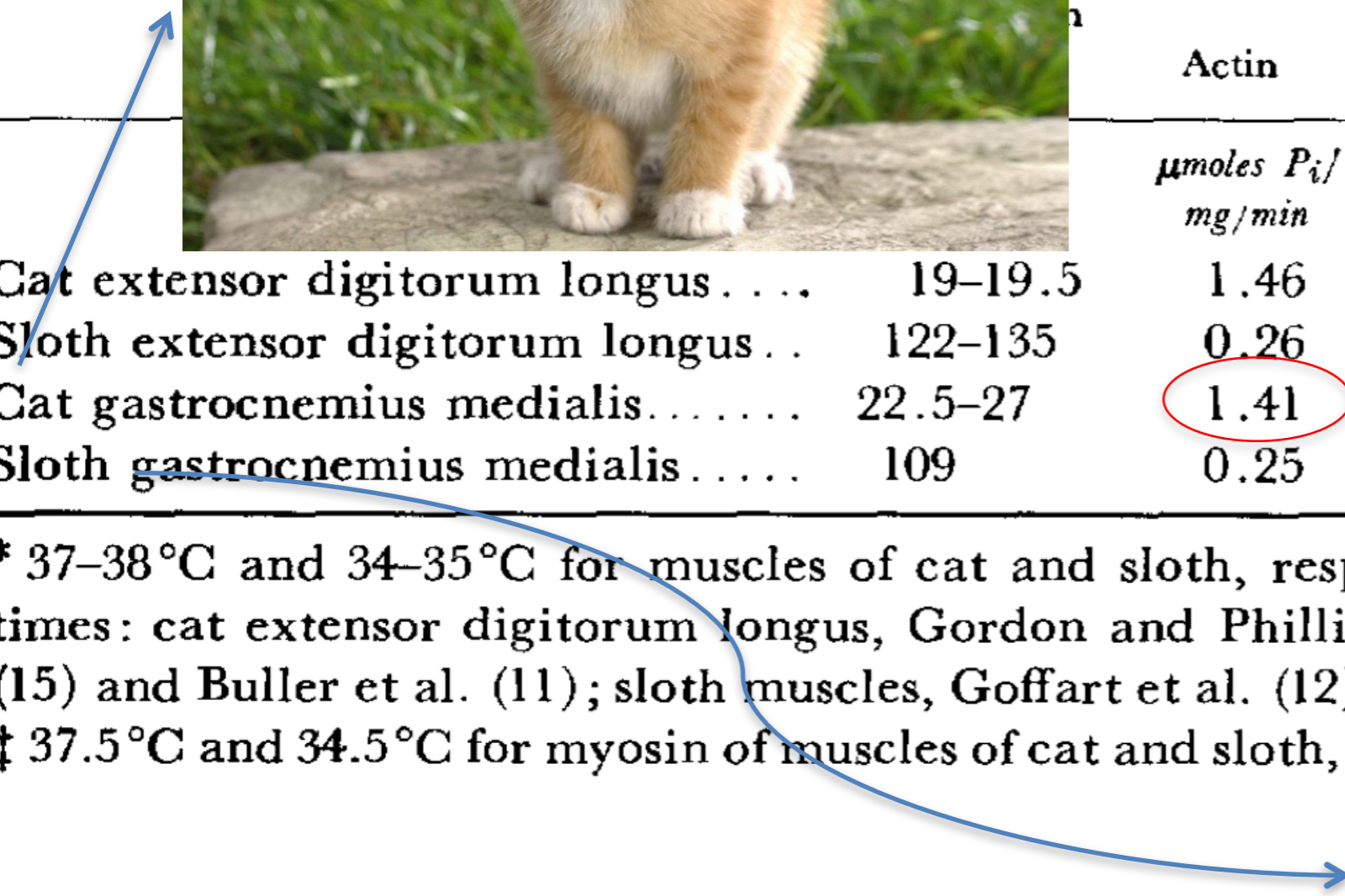
TABLE II

CONTRACTION TIME AND
IN MUSCLES OF CAT AND SLOTH

	Contraction time sec	ATPase activity‡ in the presence of			ATP sensitivity†
		Ca ⁺⁺ +0.05 μ KCl	Ca ⁺⁺ +0.5 μ KCl	EDTA	
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ATPase rate scales with muscle contraction velocity



TABLE II

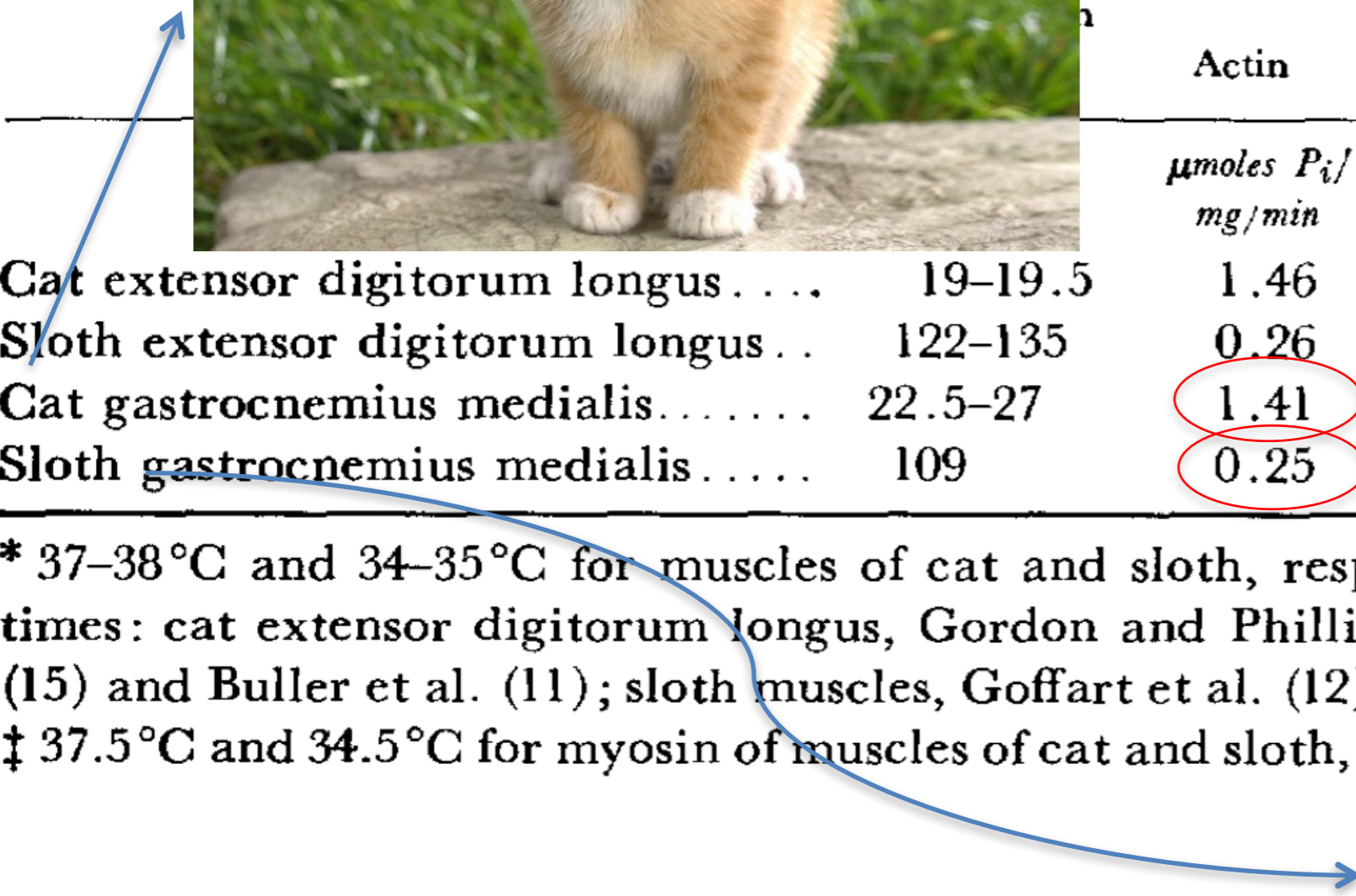
CONTRACTION TIME AND
IN MUSCLES OF CAT AND SLOTH

	Contraction time sec	ATPase activity‡ in the presence of		
		Ca ⁺⁺ +0.05 μ KCl	Ca ⁺⁺ +0.5 μ KCl	EDTA
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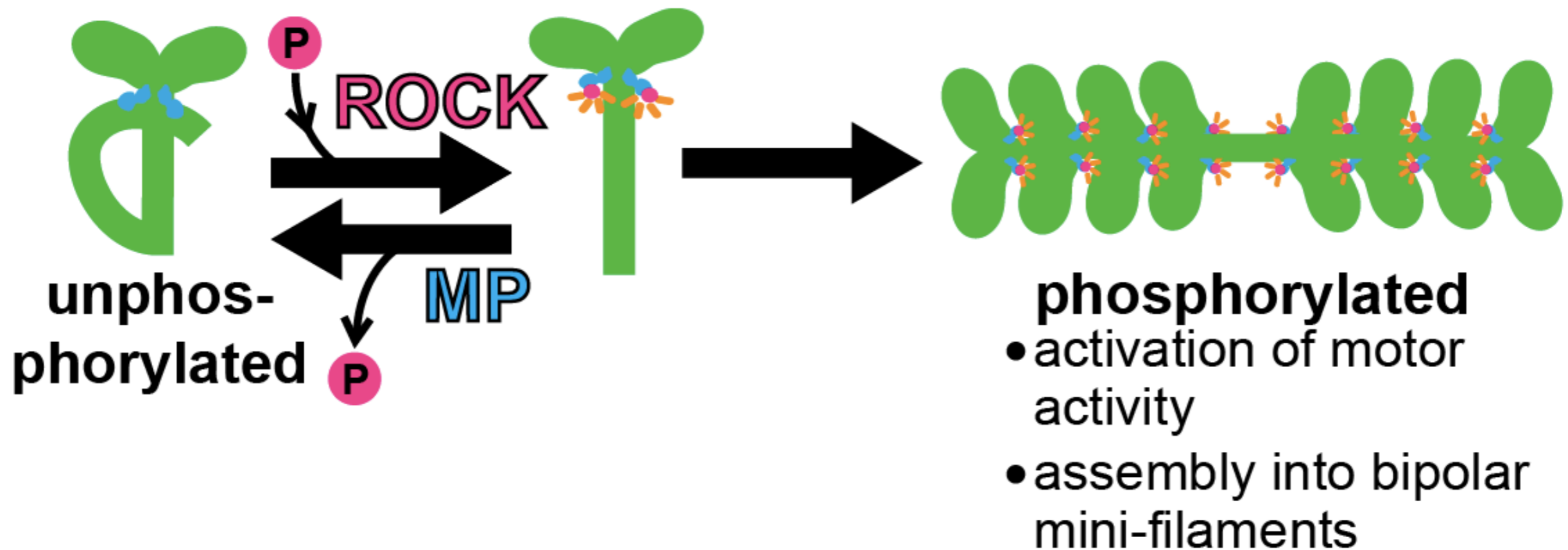
μmoles P_i/
mg/min



* 37-38°C and 34-35°C for muscles of cat and sloth, respectively: cat extensor digitorum longus, Gordon and Phillip (15) and Buller et al. (11); sloth muscles, Goffart et al. (12).
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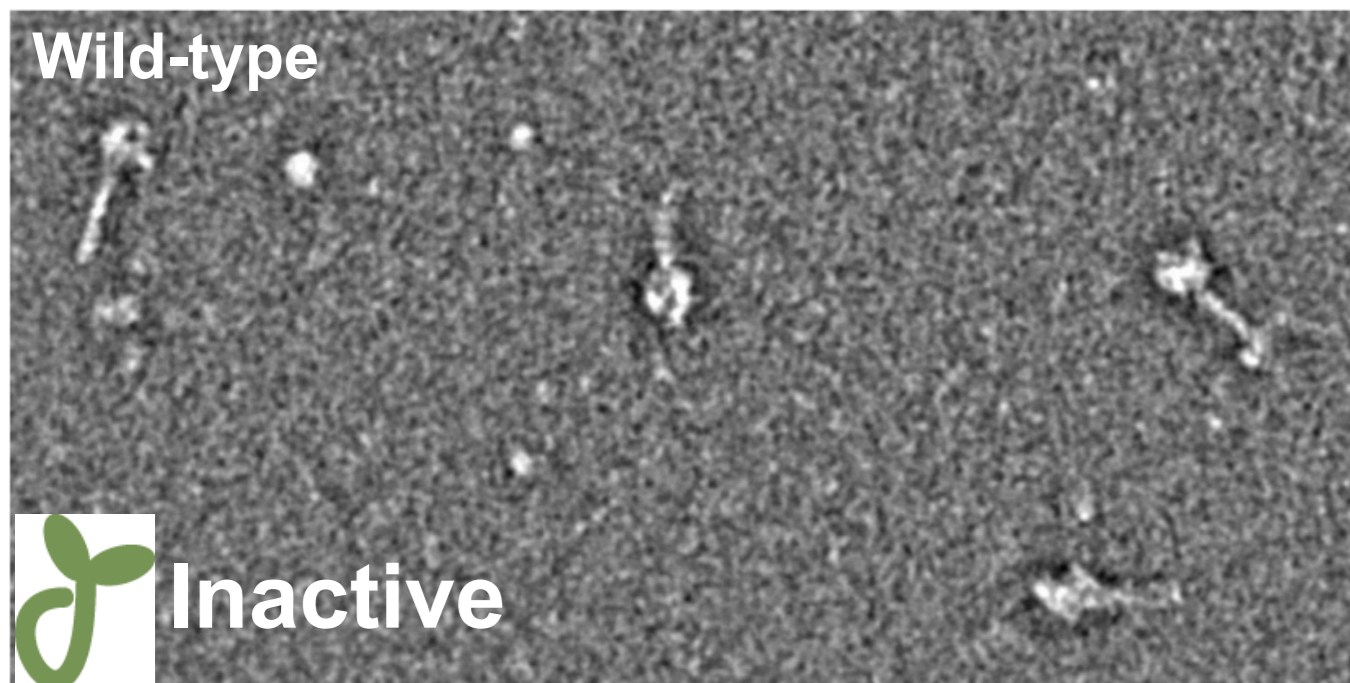


Myosin activity and assembly is regulated by phosphorylation



Drosophila myosin 2 is regulated by phosphorylation similar to mammalian myosin 2

- Phosphorylation

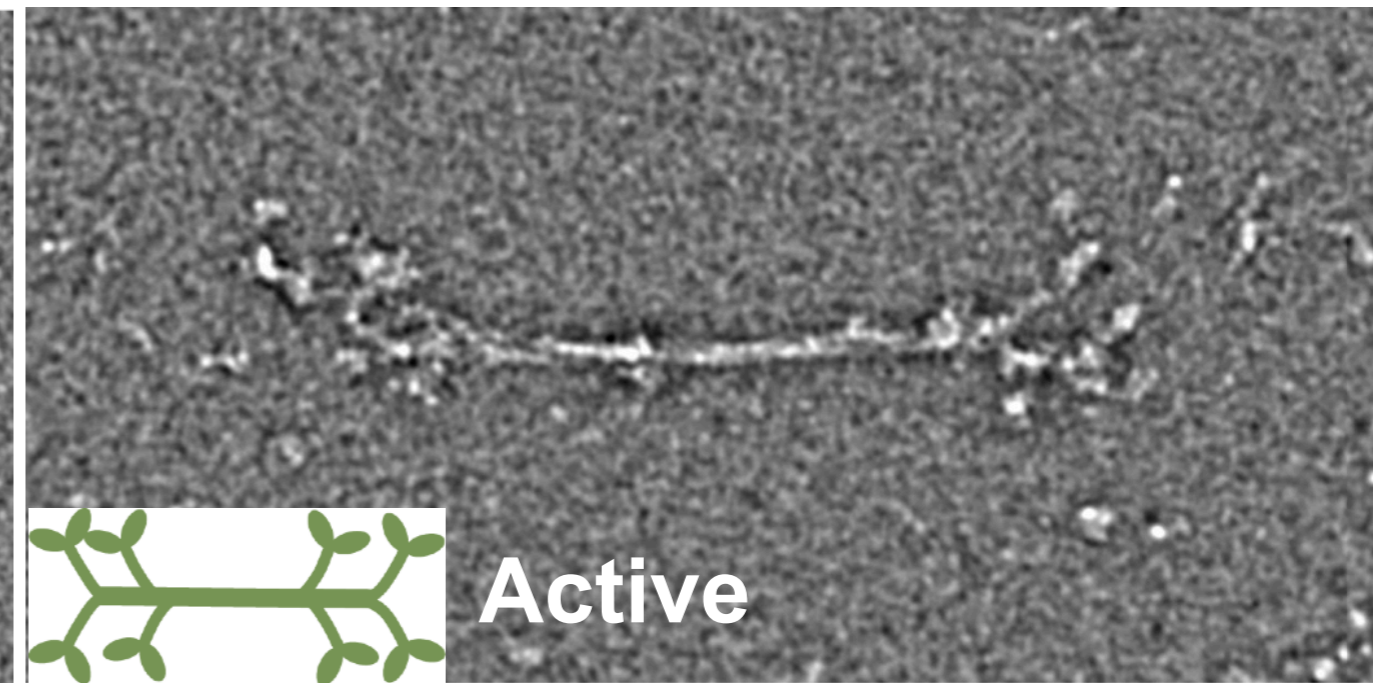
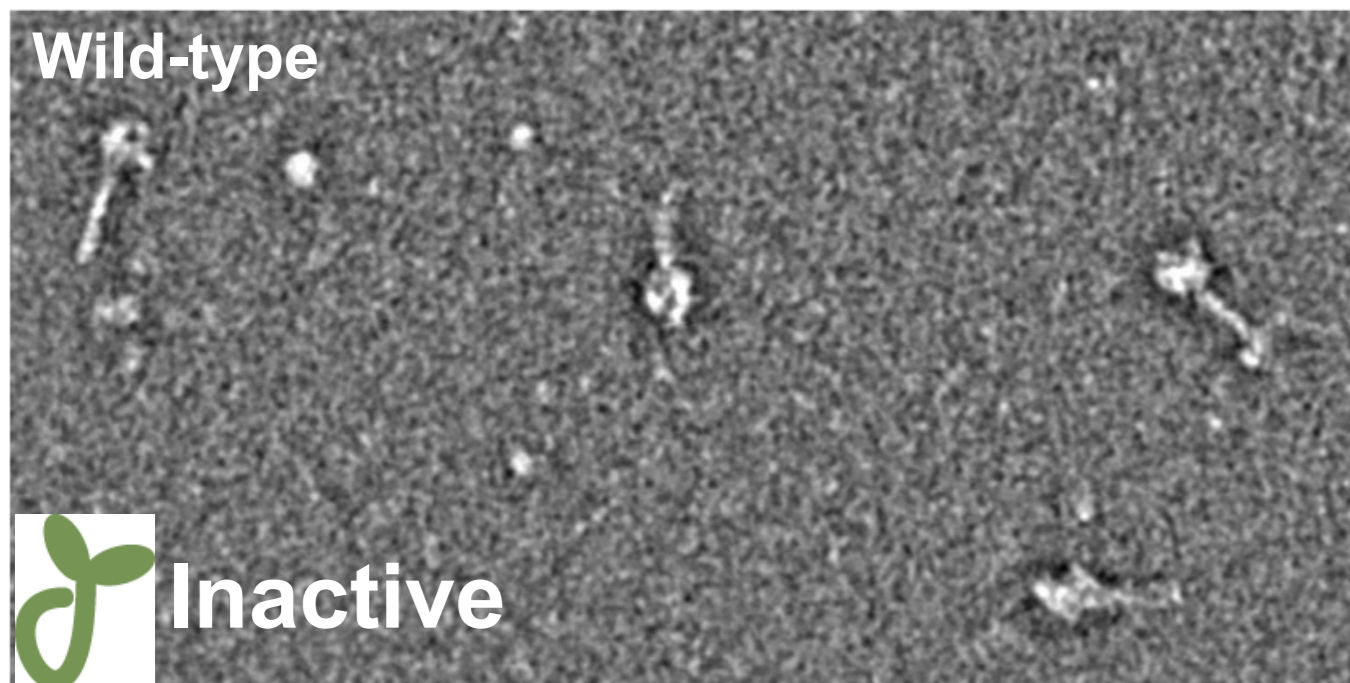


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Drosophila myosin 2 is regulated by phosphorylation similar to mammalian myosin 2

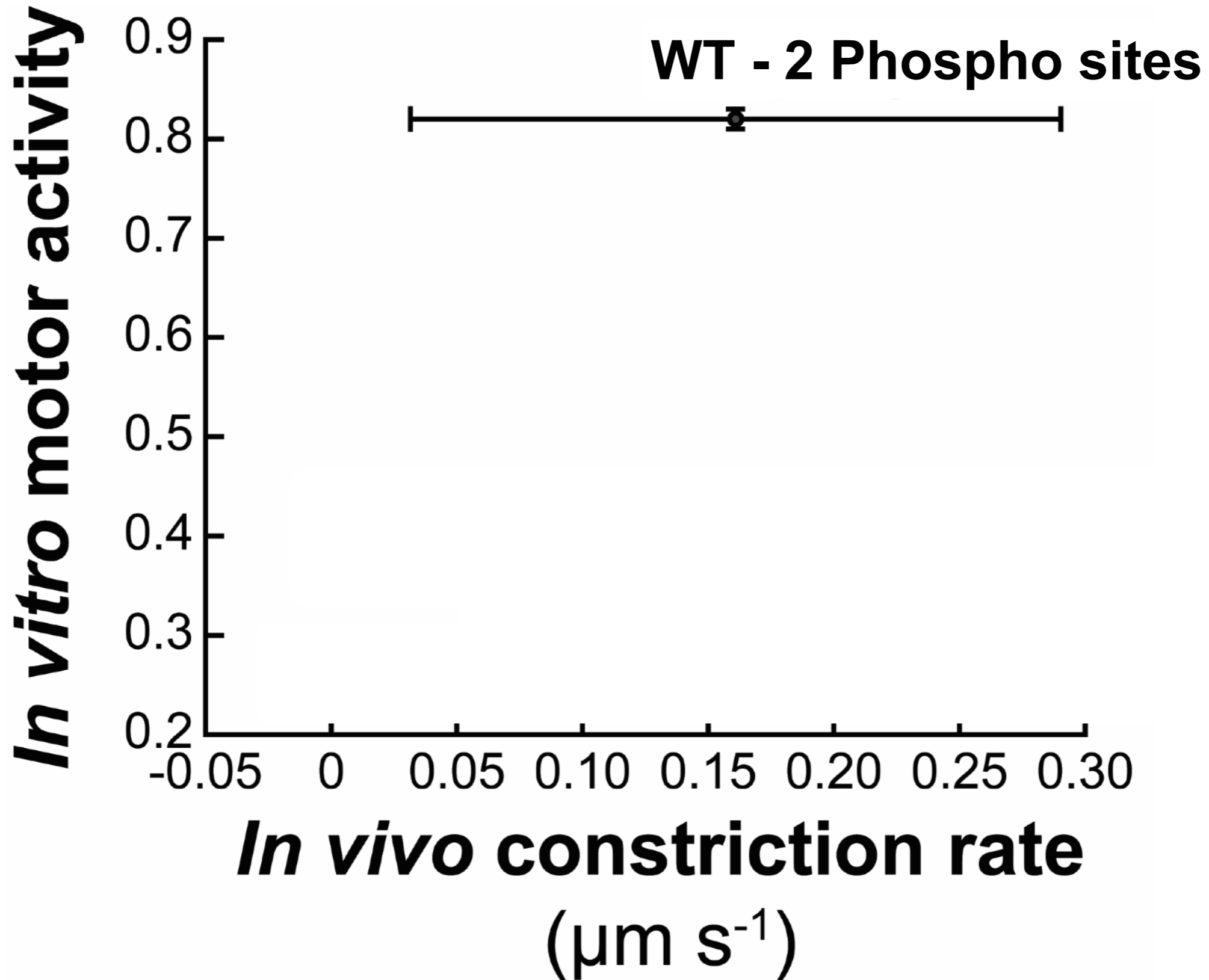
- Phosphorylation

+ Phosphorylation

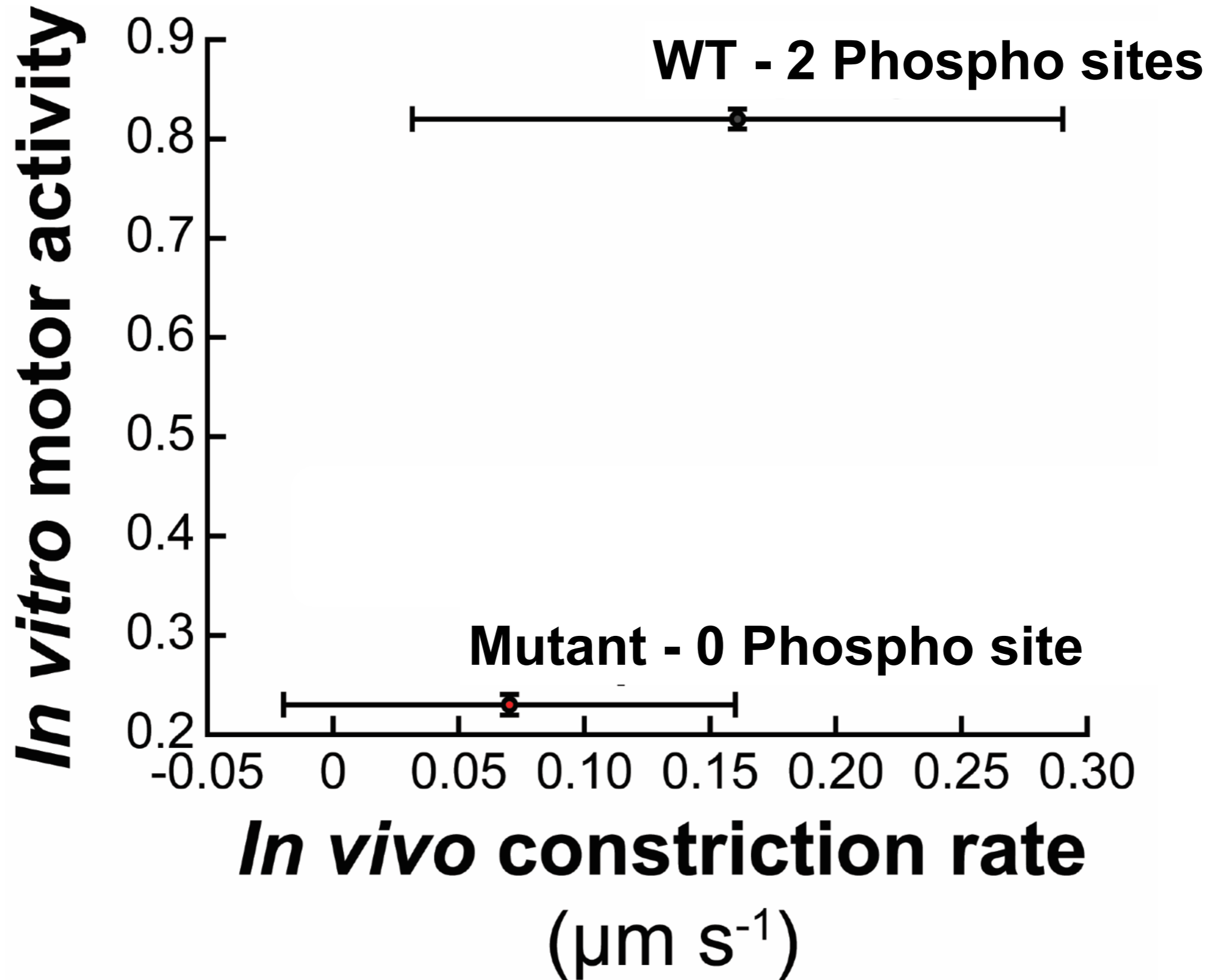


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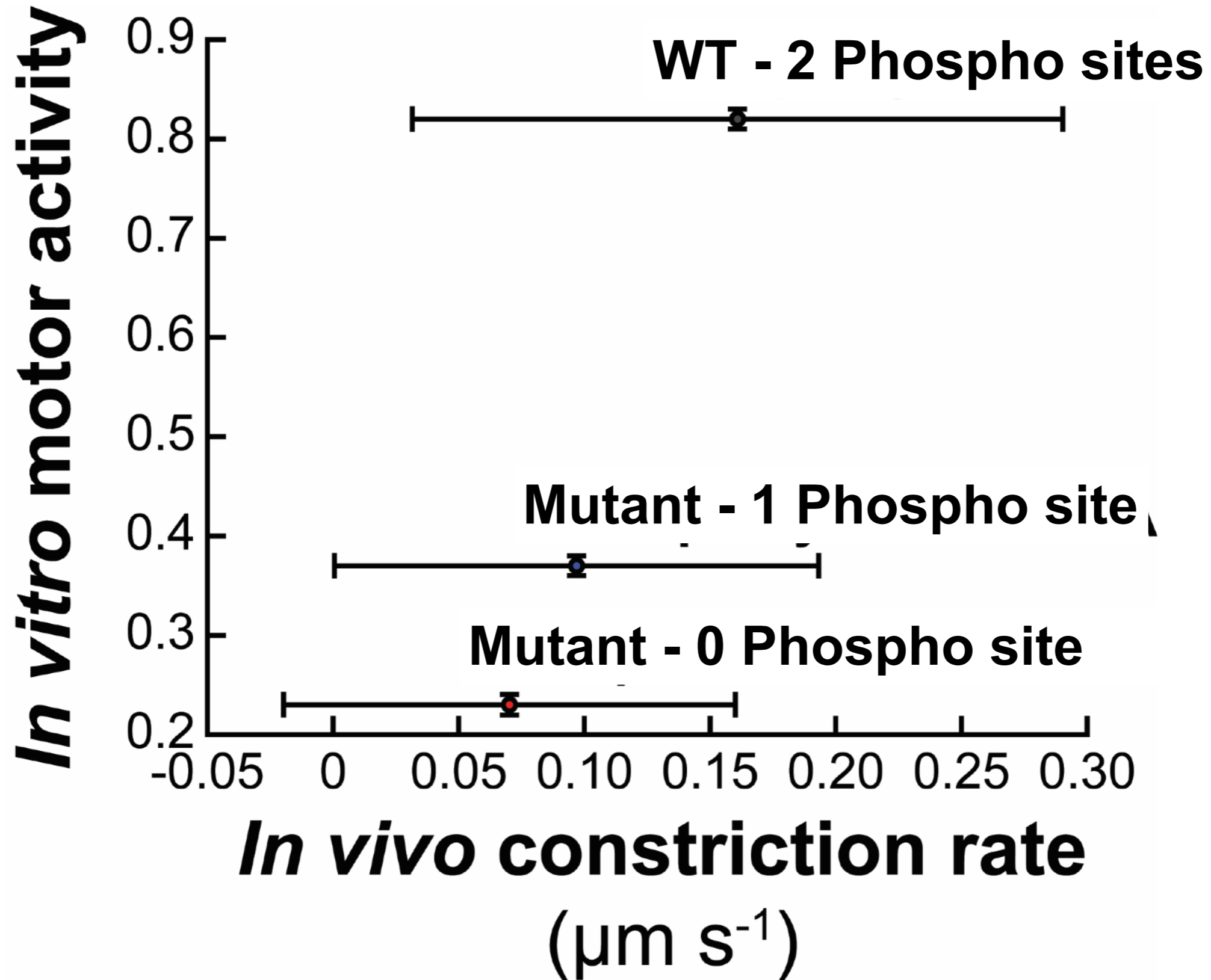
Myosin motor activity scales with contraction rate



Myosin motor activity scales with contraction rate



Myosin motor activity scales with contraction rate



Myosin motor activity scales with contraction rate

