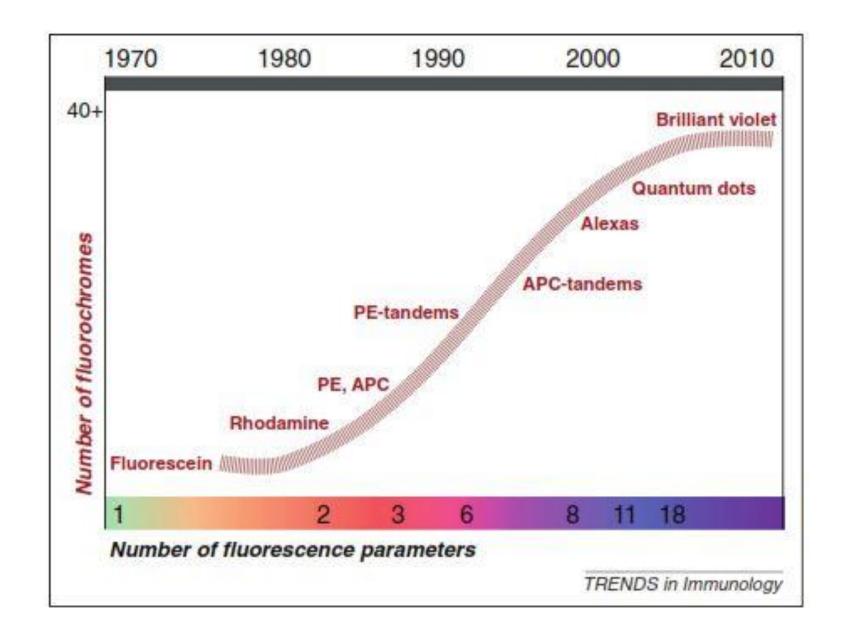
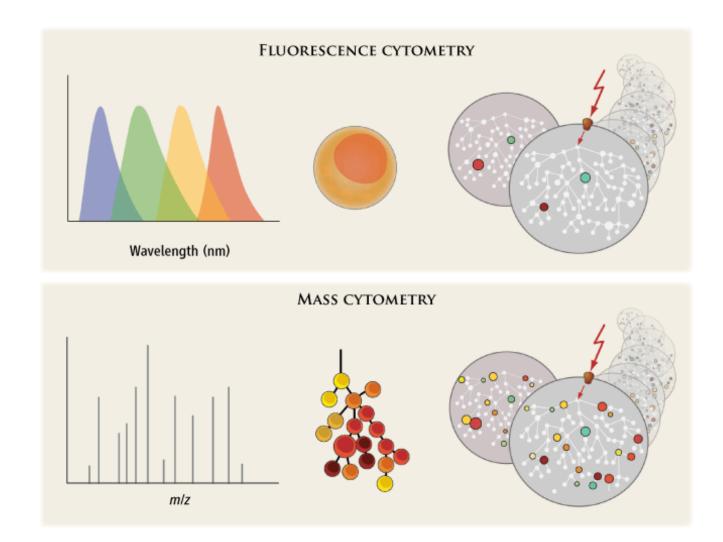
## Single-Cell Mass Cytometry of Differential Immune and Drug Responses Across a Human Hematopoietic Continuum

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**Science** vol 332 (6 May 2011) page 687

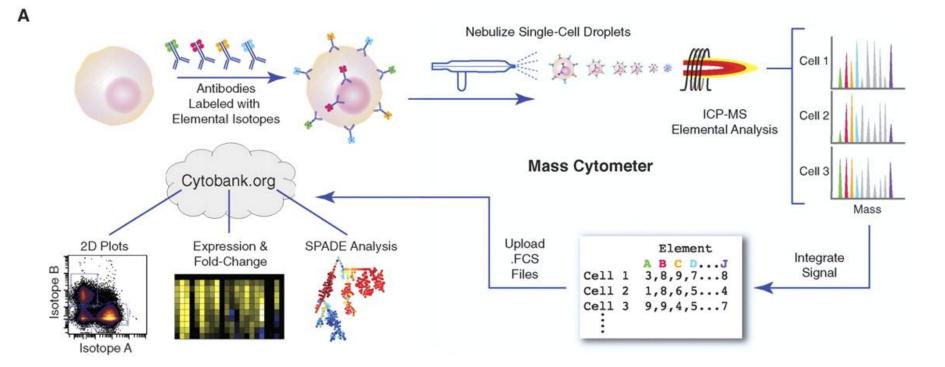


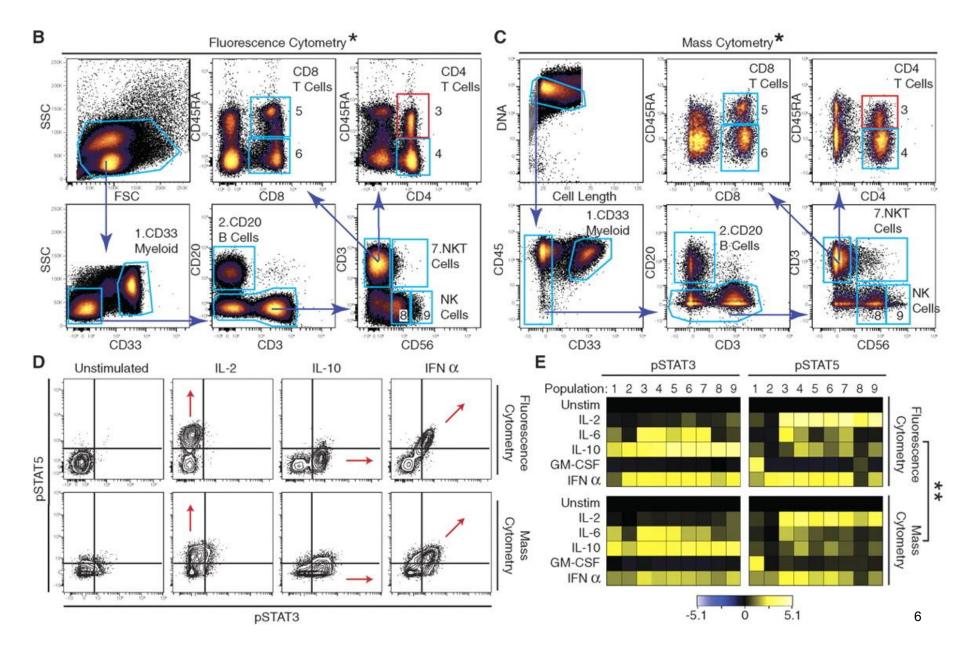
## Bendall et al. Trends in Immunology 33: 323 (2012)



## Random samples

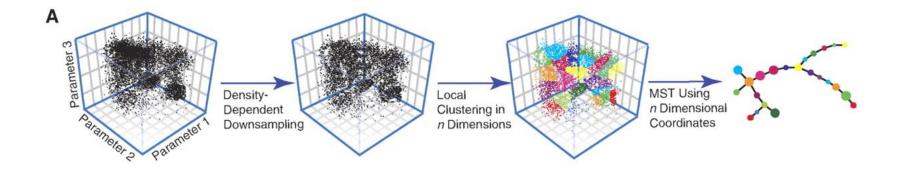
- transition element isotopes not normally found in biological systems as chelated antibody tags
- this technology can reasonably allow for a as many as 100 "tags" per cell
- currently TOF sampling resolution enables up to 1000 cells per second (~400,000 events per run)
- workflow for mass cytometry is comparable with that of fluorescence flow cytometry



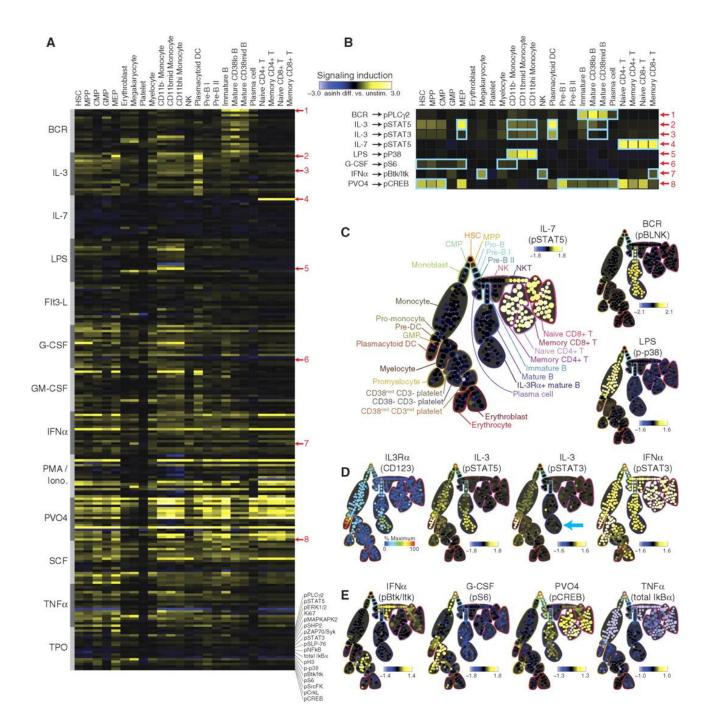


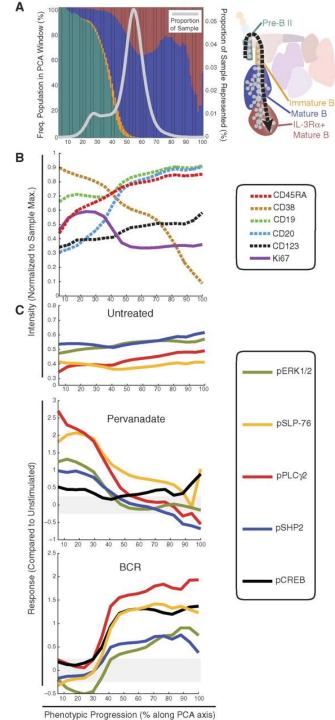
## Two panels

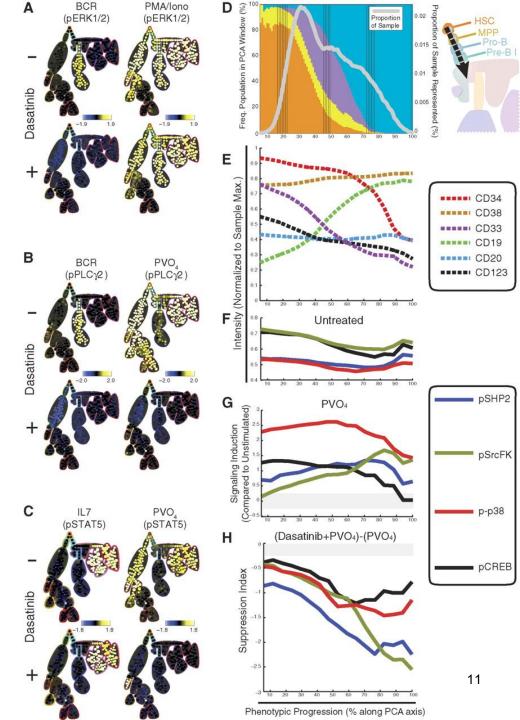
- An "immunophenotyping" panel was designed that monitored 13 "core" surface markers and 18 subset-specific cell-surface markers to allow identification of human hematologic cell types.
- A "functional" panel contained the 13 core surface markers and also 18 intracellular epitopes that reflect intracellular signaling states, such as phosphorylation status of kinase substrates.
- Analyses by clustering, heatmaps and "spanning-tree progression analysis of density-normalized events"



 The unsupervised organization of phenotypically related cell types into adjacent branches, such as CD4 and CD8 T cells (Fig. 2C), mature and immature B cells (Fig. 2D), and different clusters of myeloid cells (Fig. 2E) collectively illustrates that the algorithmic ordering of surface marker similarity can objectively organize cell types into physiologically relevant compartments.







- "A central dogma of immunology is that cells at different stages of maturation can be characterized by the expression of unique sets of proteins on the cell surface."
- "The number of nodes and ultimately their boundaries is driven by a user-definable value (21)."