

# The mechanics of cell growth

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# Outline

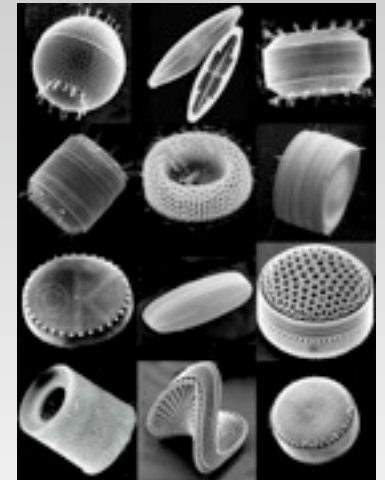
- Introduction: an atlas of shape
- Fission yeast: Minc et al. 2009–
- Plant cells (*Arabidopsis*): Hamant et al. 2008–

# Introduction

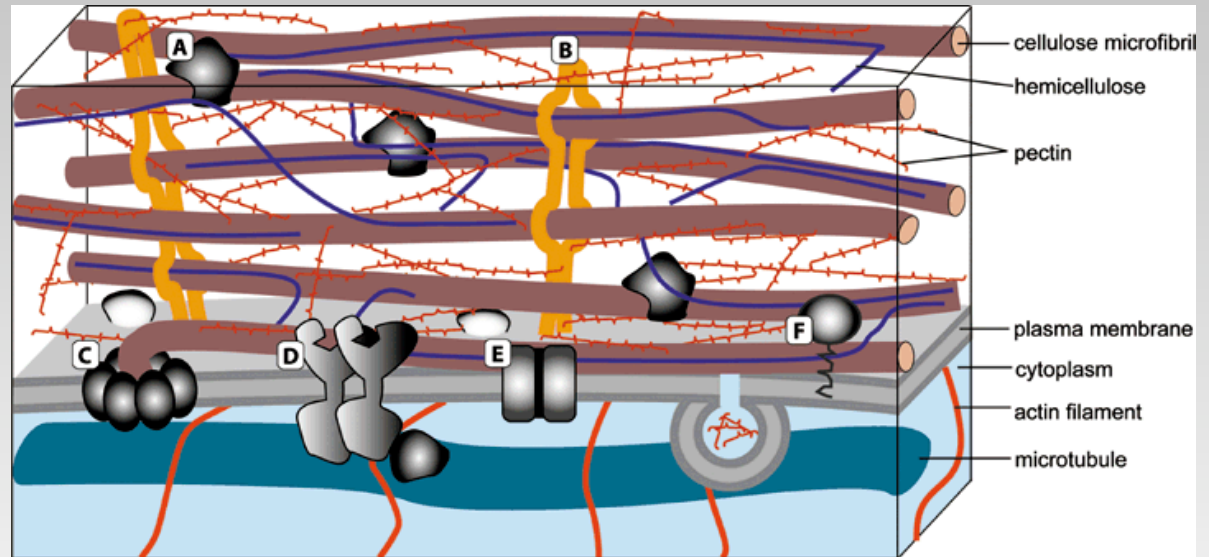
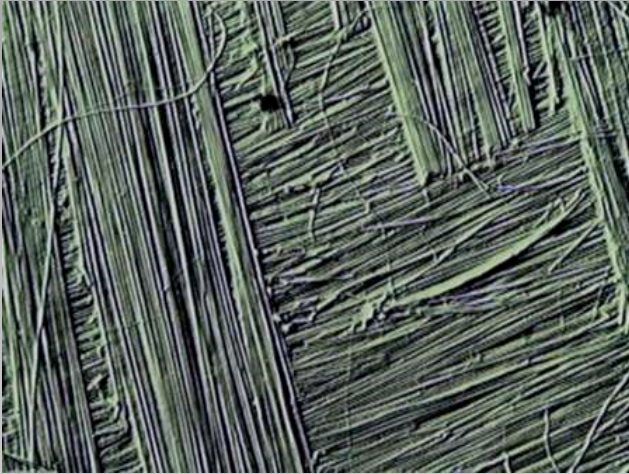
## ★ Non walled cells

## ★ Walled cells - A stiff casing

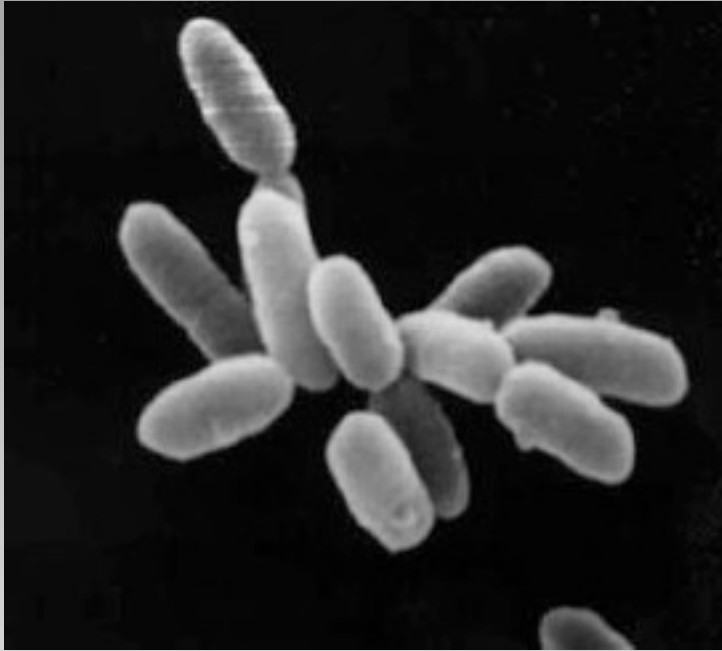
- Bacteria *peptidoglycan*
- Archea *various polymers (aminoacids + sugars)*
- Fungi *chitin*
- Plants and green algae *cellulose*
- Diatoms (not relevant here) *silica*



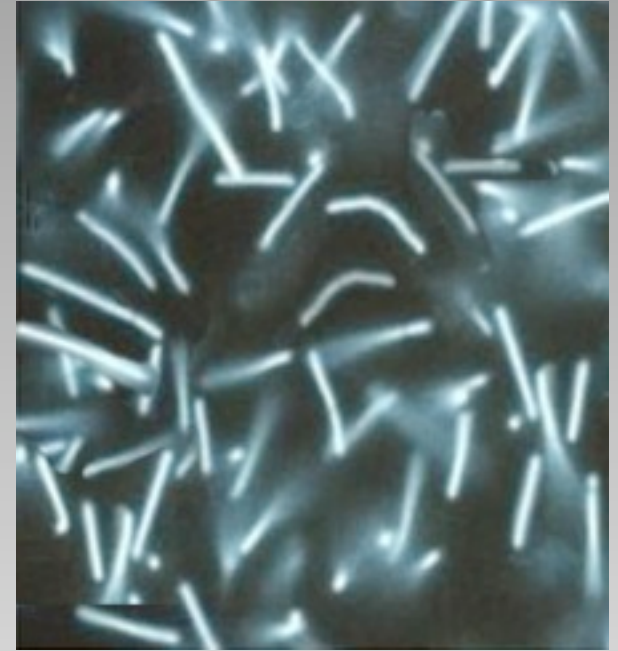
# Introduction



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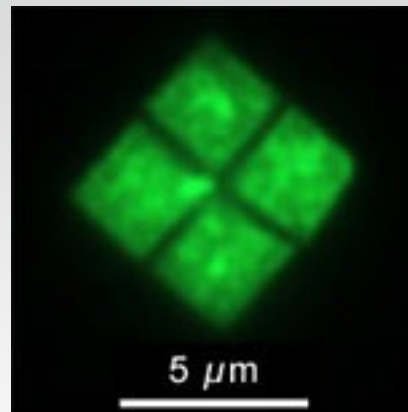


*Halobacteria*

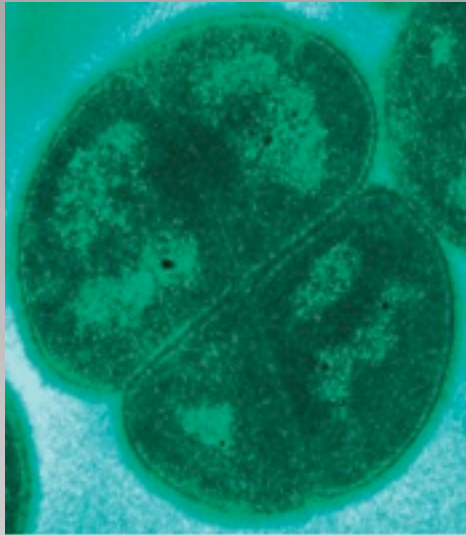


*Haloquadra walsbyi*

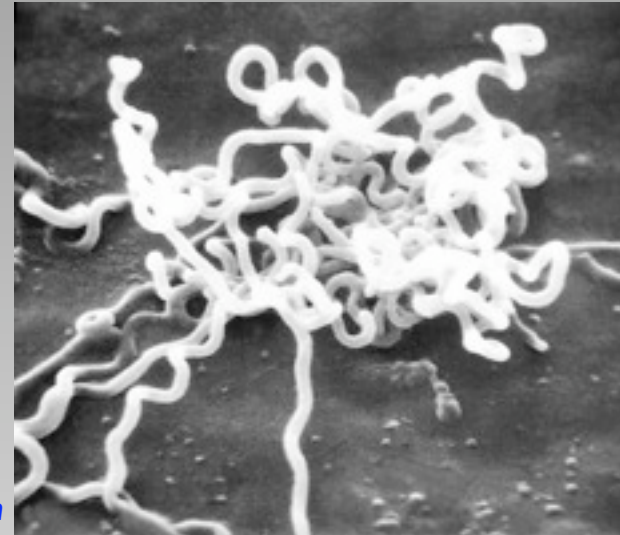
*Methanopyrus kandleri*



# Introduction



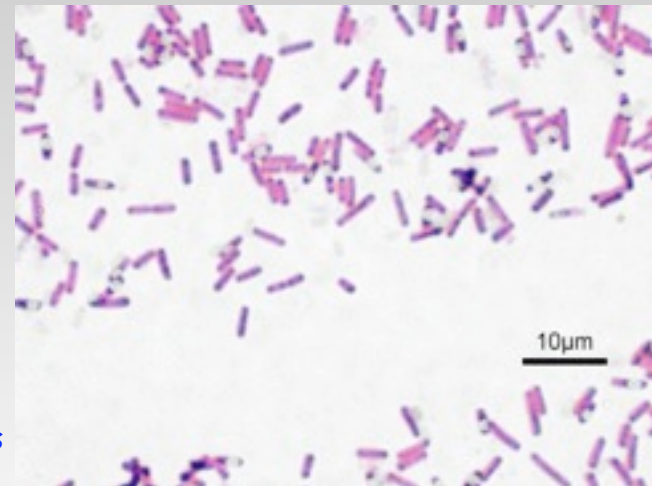
*Deinococcus radiodurans*



*Treponema pallidum*

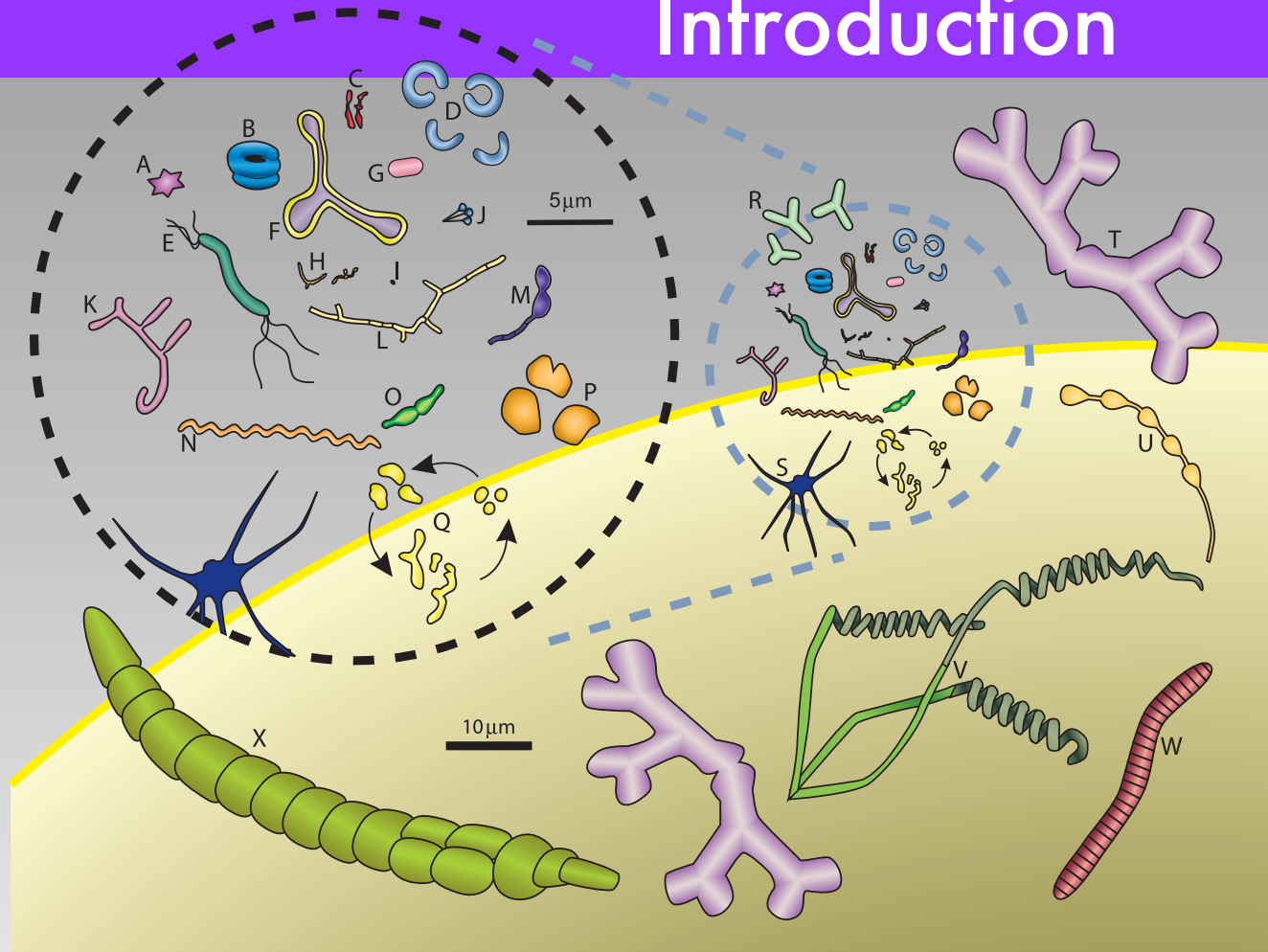


*E. coli*



*B. subtilis*

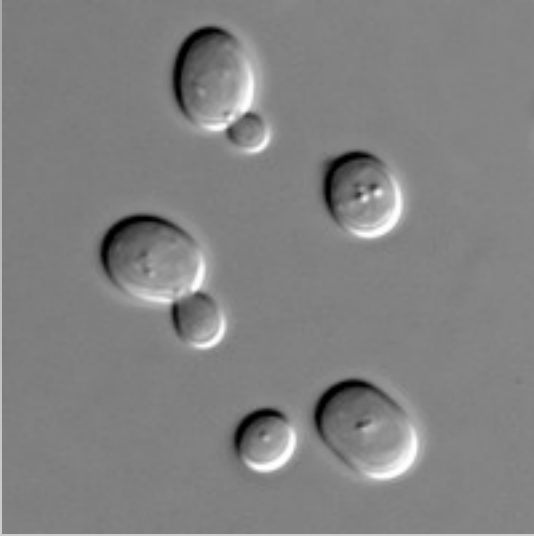
# Introduction



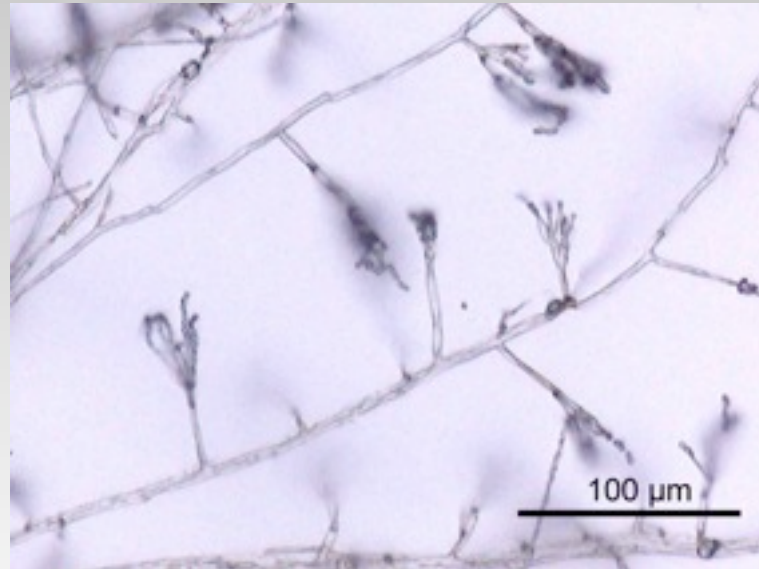
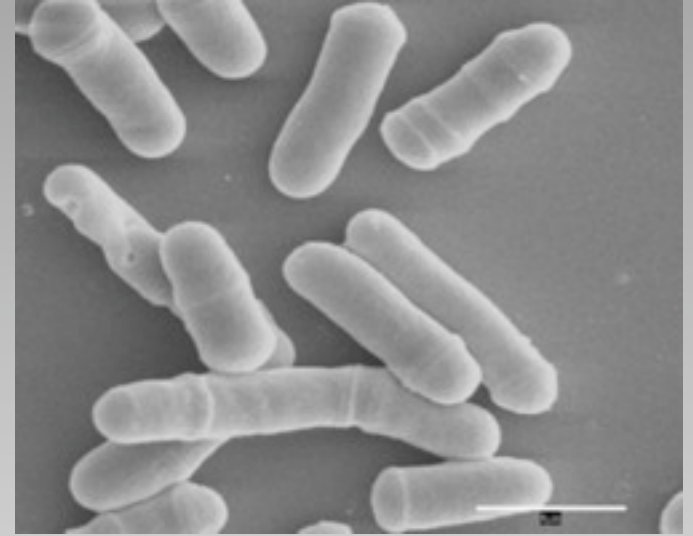
K.D. Young 2006

FIG. 1. Variety of prokaryotic shapes. This collage of different cells, unless otherwise stated, is constructed from descriptions and illustrations given by Starr et al. (313) or by Zinder and Dworkin (380). The cells are drawn to scale. Those in the dashed black circle are drawn relative to the 5- $\mu\text{m}$  line. These same cells are included in smaller form in the dashed blue circle to compare their sizes to those of larger bacteria, which are drawn relative to the 10- $\mu\text{m}$  line. (A) *Stella* strain IFAM1312 (380); (B) *Microcyclus* (a genus since renamed *Ancylobacter*) *flavus* (367); (C) *Bifidobacterium bifidum*; (D) *Clostridium cocleatum*; (E) *Aquaspirillum autotrophicum*; (F) *Pyroditium abyssii* (380); (G) *Escherichia coli*; (H) *Bifidobacterium* sp.; (I) transverse section of ratoon stunt-associated bacterium; (J) *Planctomyces* sp. (133); (K) *Nocardia opaca*; (L) Chain of ratoon stunt-associated bacteria; (M) *Caulobacter* sp. (380); (N) *Spirochaeta halophila*; (O) *Prostheco bacter fusiformis*; (P) *Methanogenium cariaci*; (Q) *Arthrobacter globiformis* growth cycle; (R) gram-negative *Alphaproteobacteria* from marine sponges (240); (S) *Ancalomicrobium* sp. (380); (T) *Nevskia ramosa* (133); (U) *Rhodomicrobium vannielii*; (V) *Streptomyces* sp.; (W) *Caryophanon latum*; (X) *Calothrix* sp. The yellow-lined background orb represents a slice of the giant bacterium *Thiomargarita namibiensis* (290), which is represented to scale with the other organisms.

# Introduction



*Penicillium*



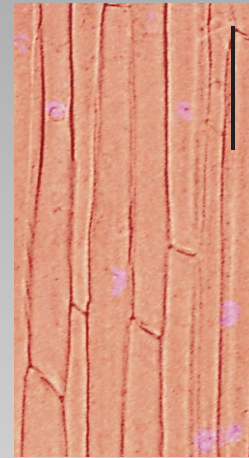


# Introduction



Courtesy J. Dumais

Ditengou et al. 2008



*Chlamydomonas reinhardtii*



# Introduction

★ A wide occurrence of rod-like shape

★ Constraints?

- Nutrients => surface/volume
- Adhesion to substrate
- Resistance (motile)
- Exploration of space (non motile)
- Partition of material between daughter cells

# Introduction

★ Stiff casing

★ Growth into rod-like shape

- Turgor pressure

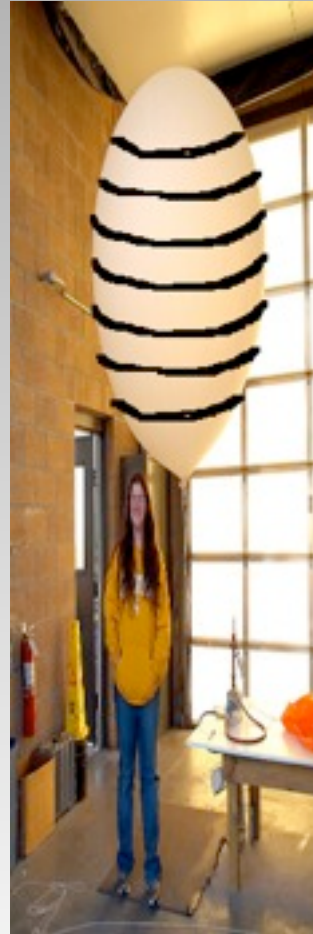
# Introduction – Structure



# Introduction – Structure



# Introduction – Structure



# Introduction – Structure

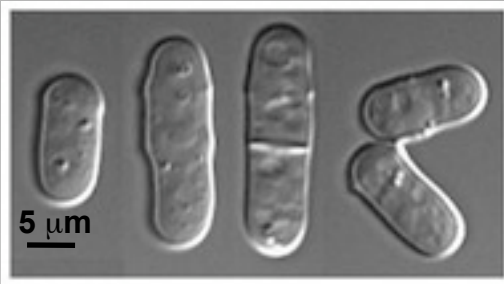
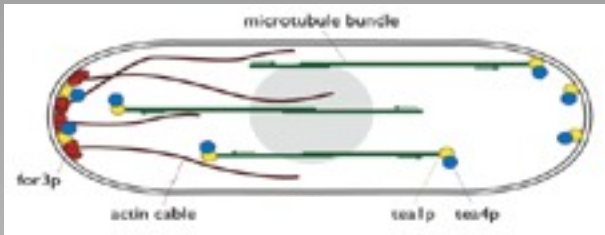


# Outline

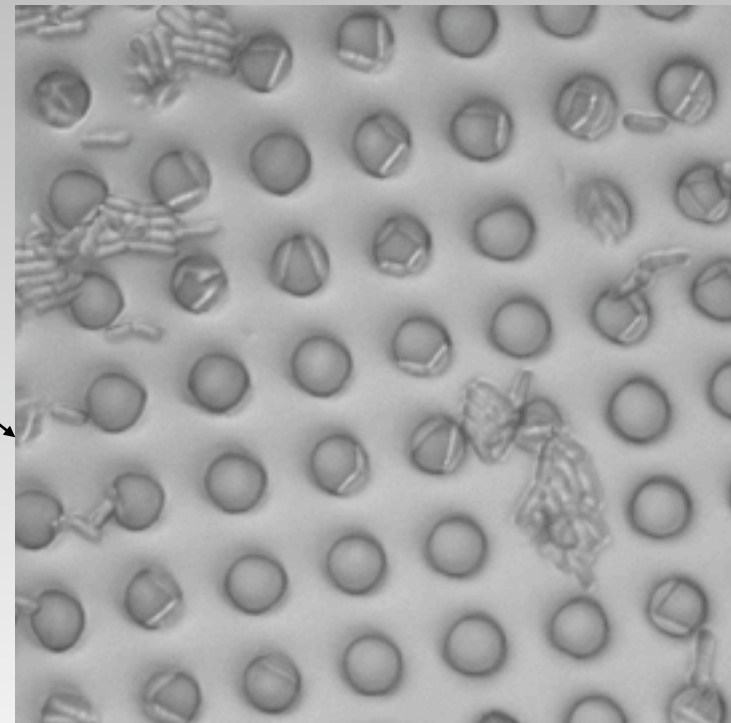
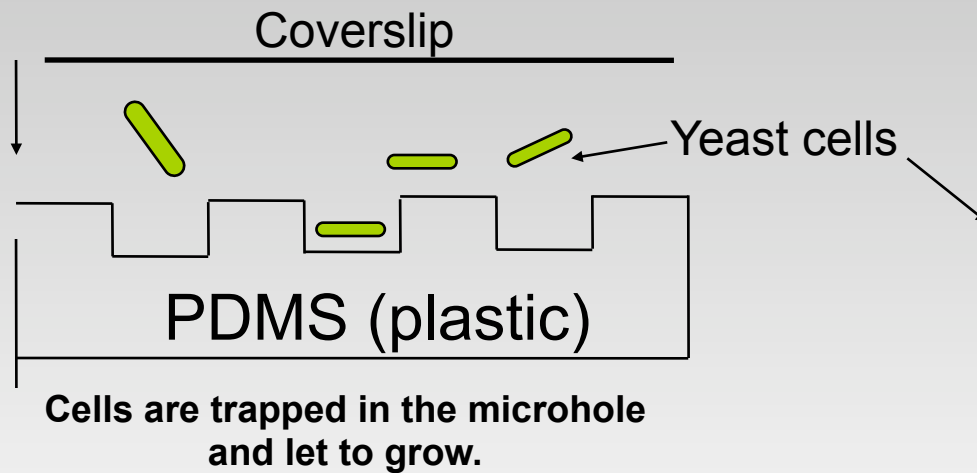
- Fission yeast: Minc et al. 2009—  
What are the forces involved?
- Plant cells (*Arabidopsis*): Hamant et al. 2008—  
How is anisotropic growth controlled?



# Fission yeast

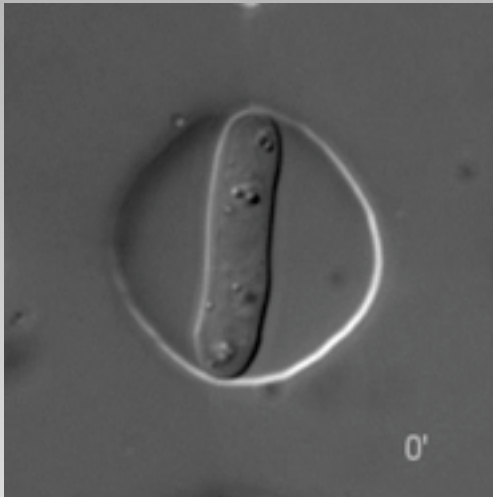


The fission yeast *Schizosaccharomyces pombe*, is a model system for studying mechanisms of polarized growth. Cell polarity is dynamically regulated by the microtubule and actin cytoskeletons

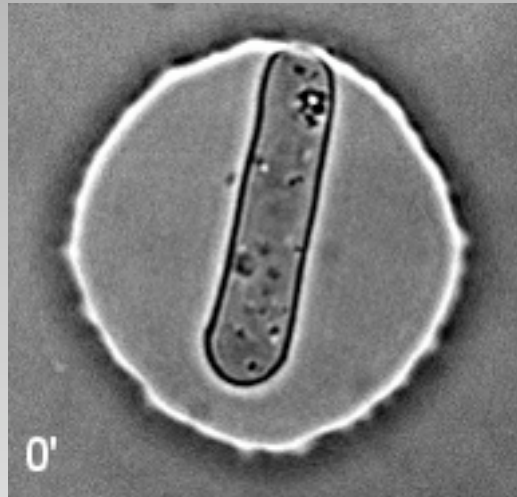
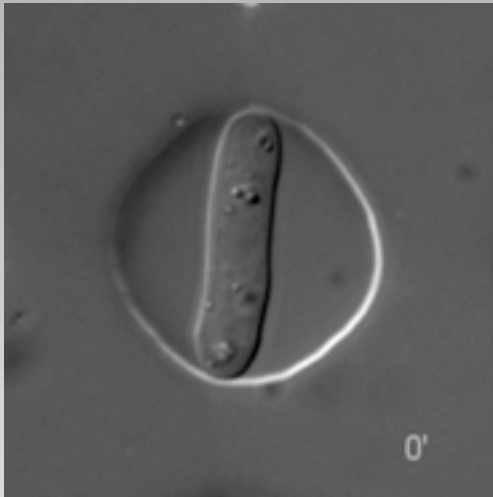


Top view of fission yeast cells trapped in the chambers

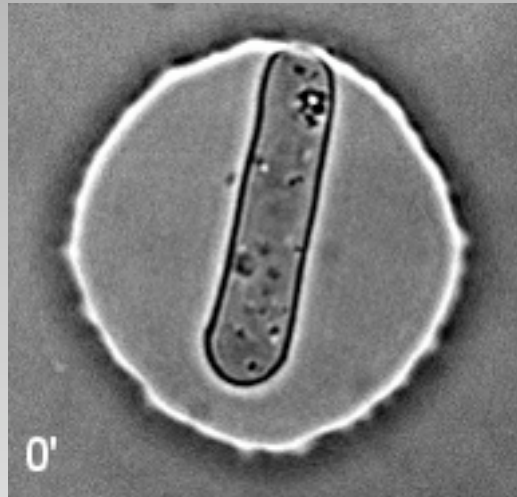
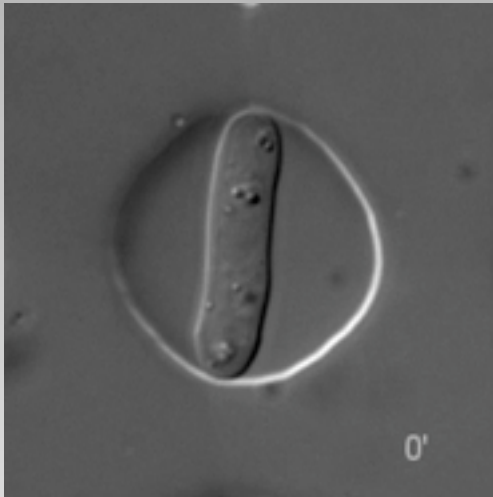
# Fission yeast



# Fission yeast



# Fission yeast



# Fission yeast



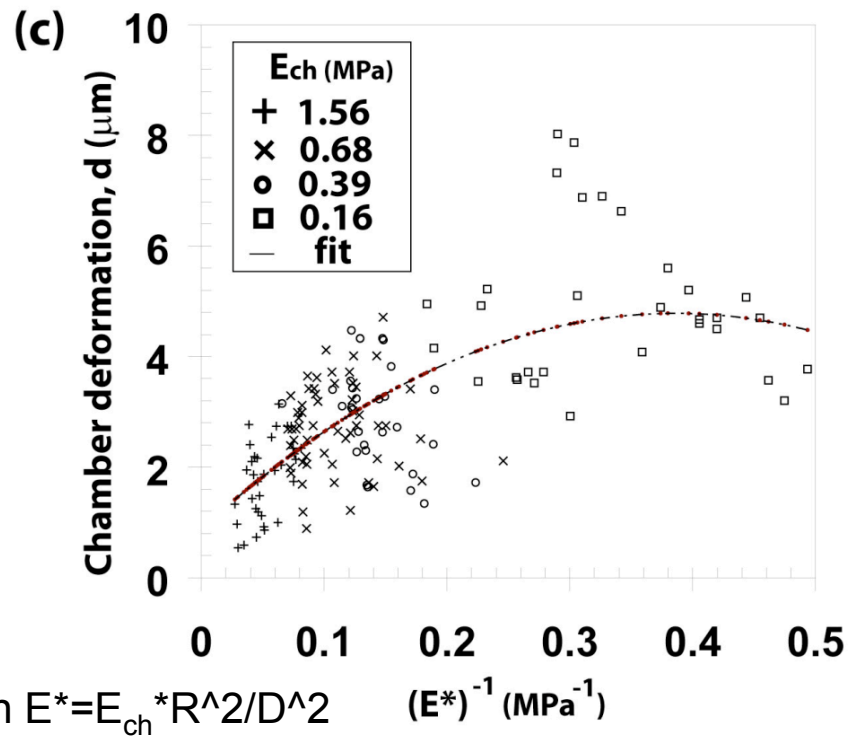
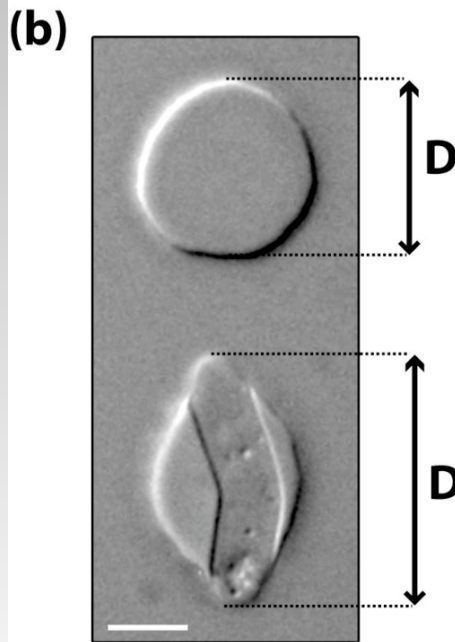
Ech (Mpa): 1.56

0.68

0.39

0.16

$$F_{ch} = \frac{8}{3} E_{ch} R d$$



$$F_B = \frac{\pi I E_{cw}}{L_T^2}$$

$$I = \pi h R^3$$

# Fission yeast

$$E_{\text{fission yeast}} = 100 \pm 30 \text{ MPa}$$

## Other fungis:

- *S. cerevisiae* : 100 Mpa (by micromanipulation) and 0.9 Mpa (by AFM)????
- *Aspergillus nidulans*: 60-100 Mpa (AFM)

## Plants :

- Root hair (*Arabidopsis*): 500 Mpa

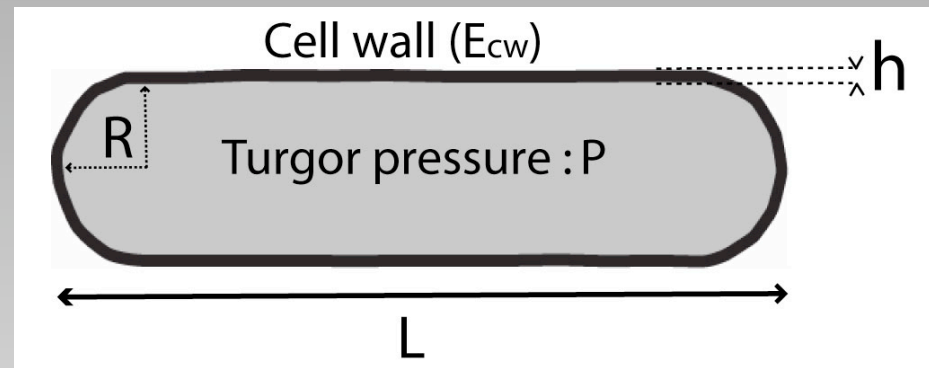
## Bacteria:

- *E.coli*: 25 Mpa
- *B subtilis*: 13-25 Mpa

# Fission yeast

$$T_P = \frac{PR}{2h}$$

$$v_0 = \frac{dL}{dt} = \frac{R}{\tau_V} \frac{T_P}{E} = \frac{P}{\tau_V} \frac{R^2}{2\sigma_{cw}}$$

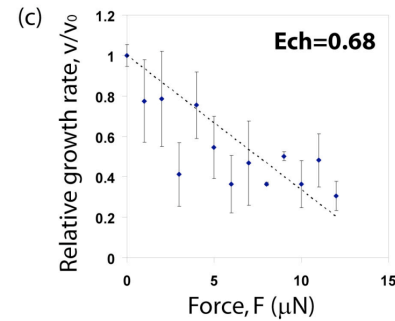
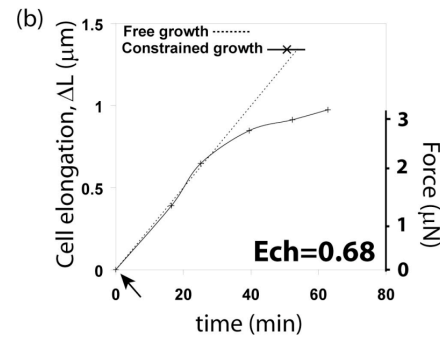
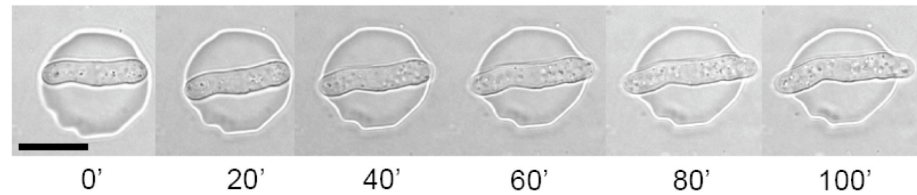


If the cell is growing under an external force  $F$ , the tension in the wall is reduced:

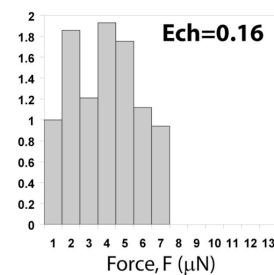
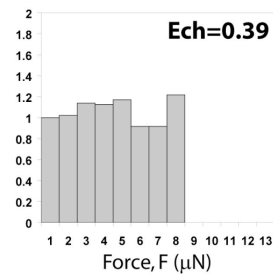
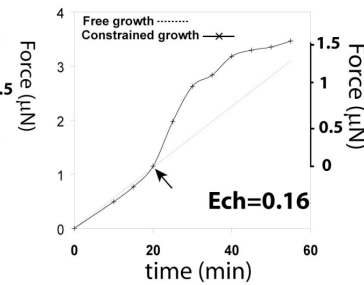
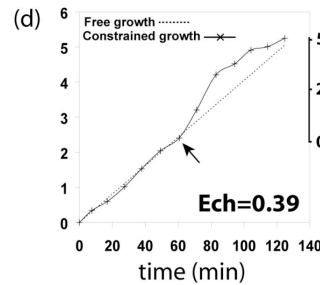
$$T = T_P - \frac{F}{2\pi R h}$$

$$v(F) = v_0 \left( \frac{P}{P_0} - \frac{F}{P_0 S} \right)$$

# Fission yeast



$$P_0 = 1.2 \pm 0.1 \text{ MPa.}$$



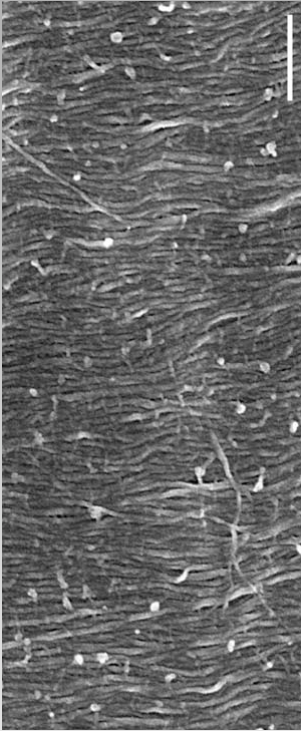


# Fission yeast

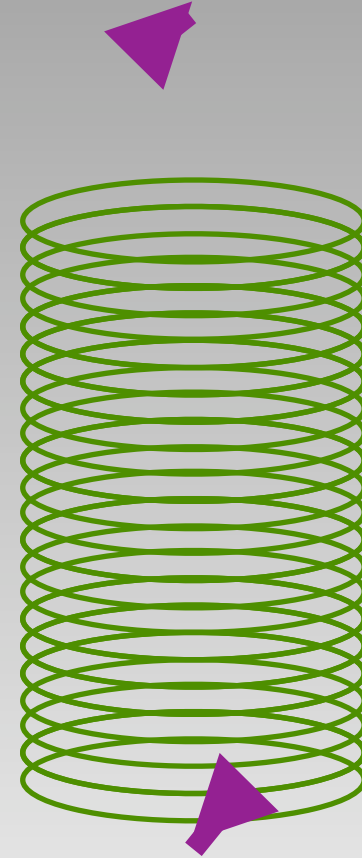
Work in progress

- Characterise mutants
- Cell division

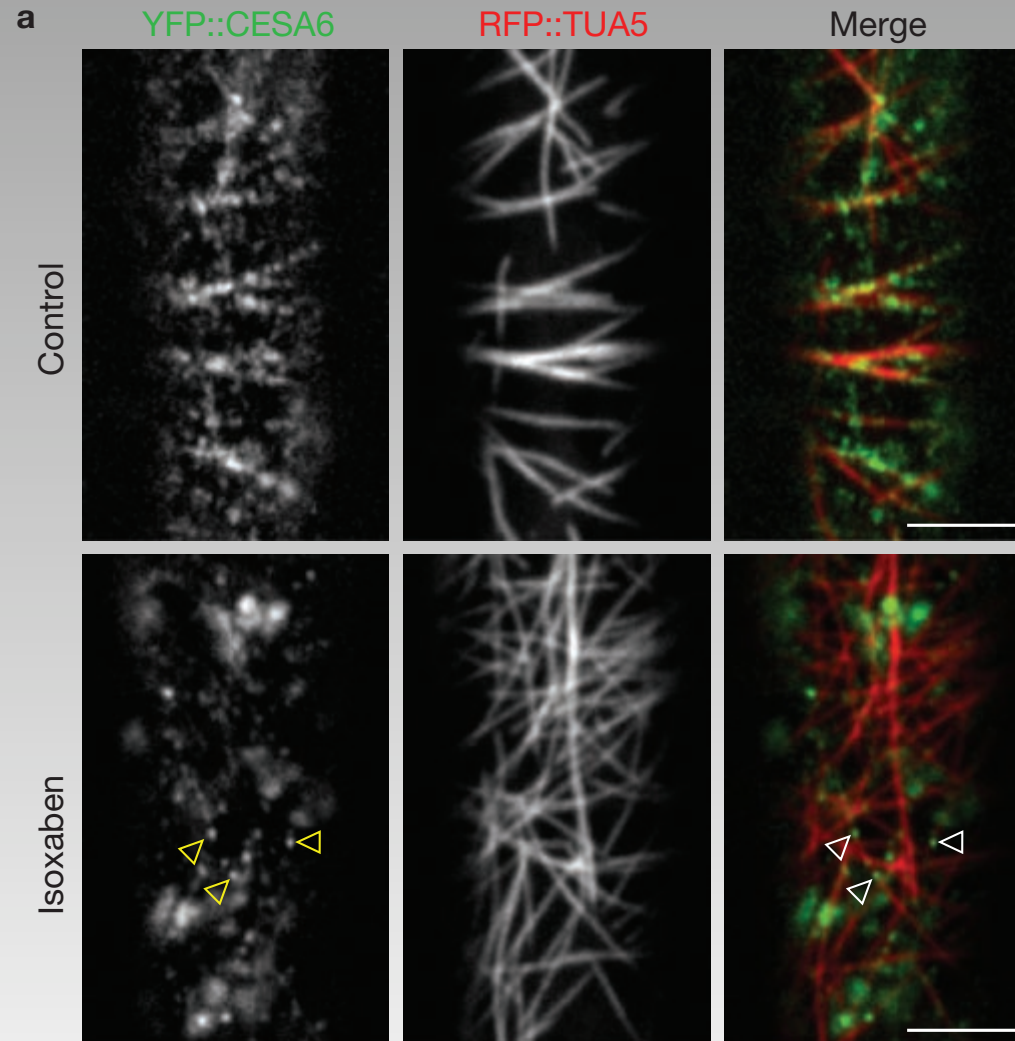
# Introduction – Structure



Marga et al. 2005



# Introduction – Building the structure

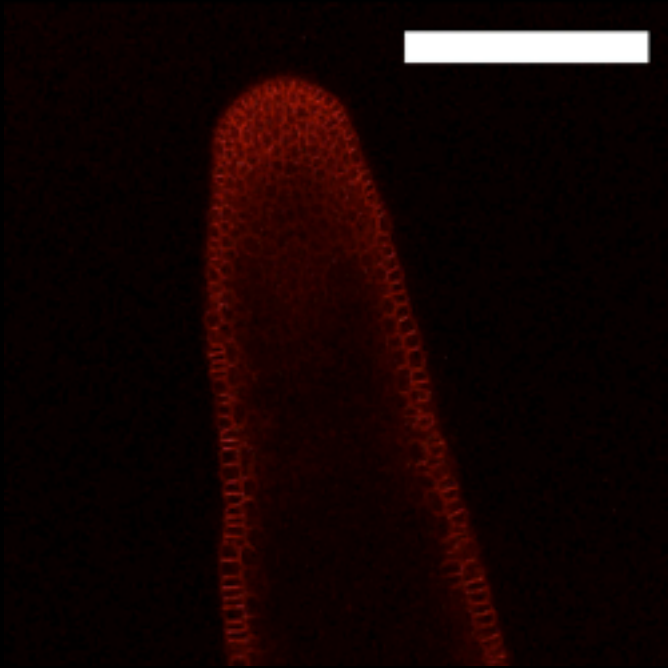


Gutierrez et al. 2009

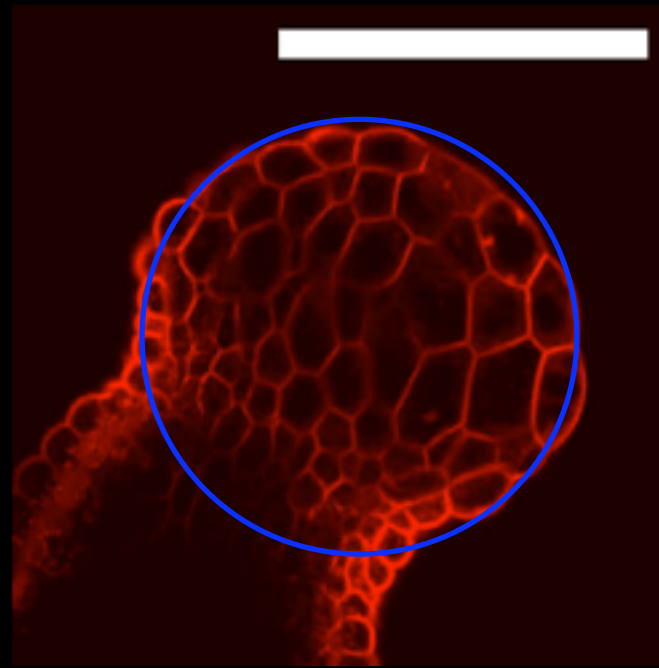
Control of CMT direction?

# Playing with CMTs

Grandjean et al. 2004

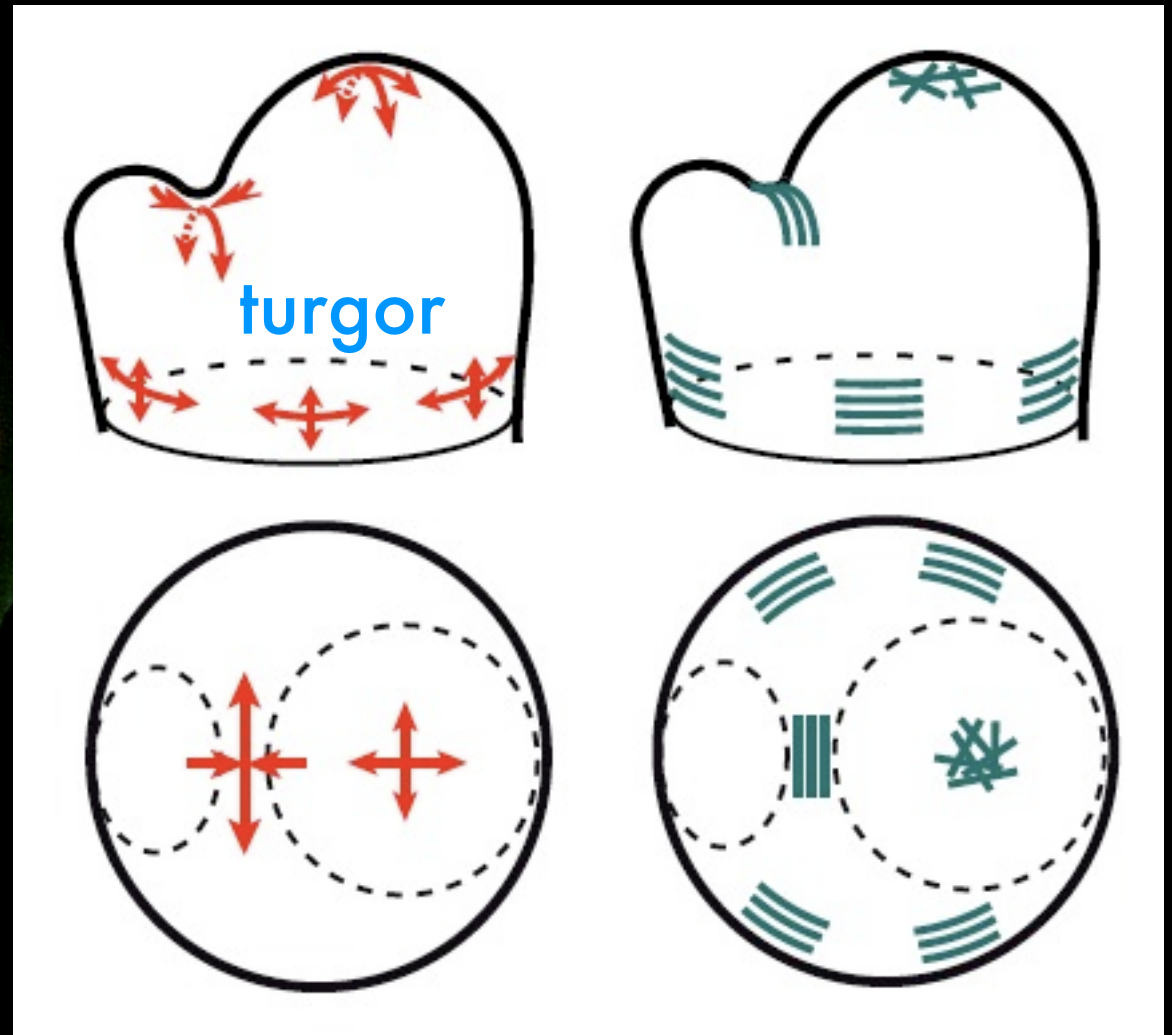
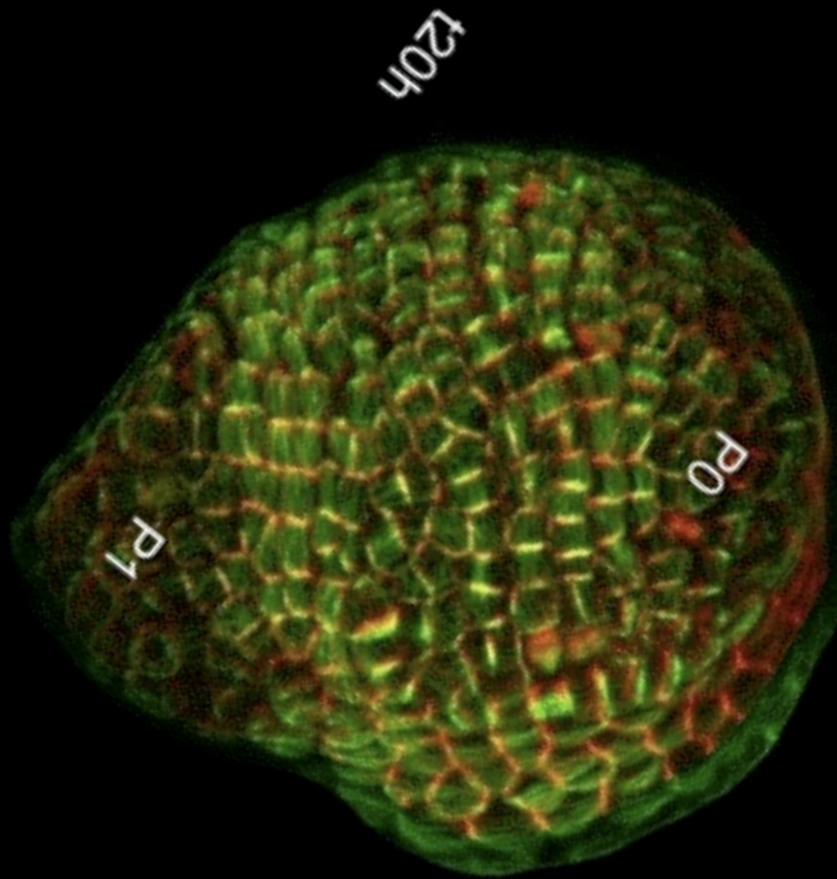


NPA



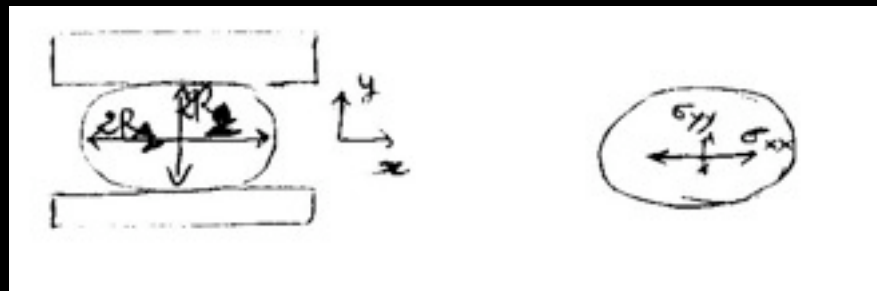
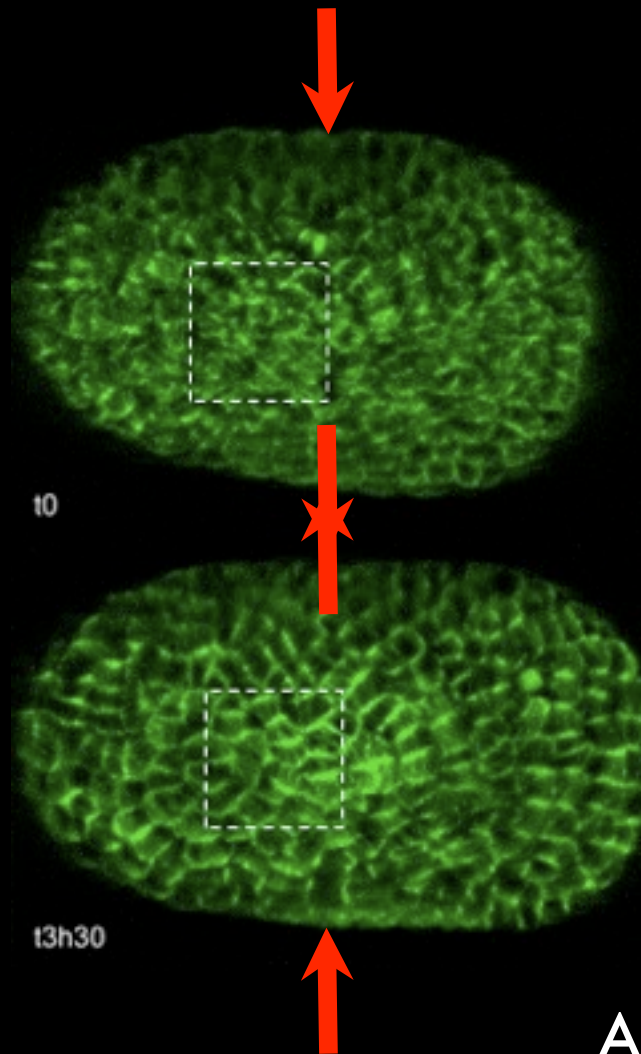
NPA + oryzalin

# Mechanical forces and CMTs patterns



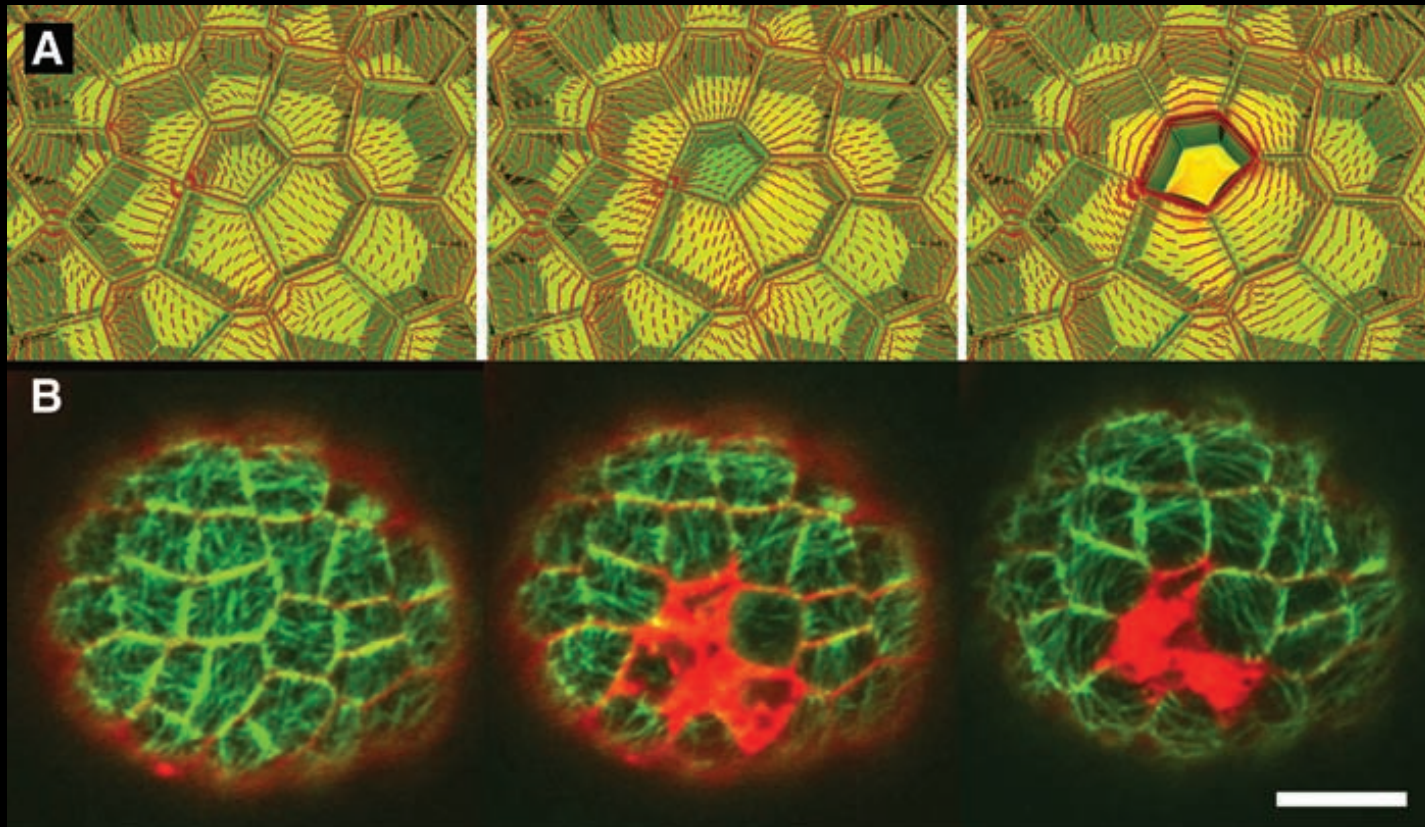
Alignment in the direction of maximal force

# Mechanical forces and CMTs patterns



Alignment in the direction of maximal force

# Mechanical forces and CMTs patterns



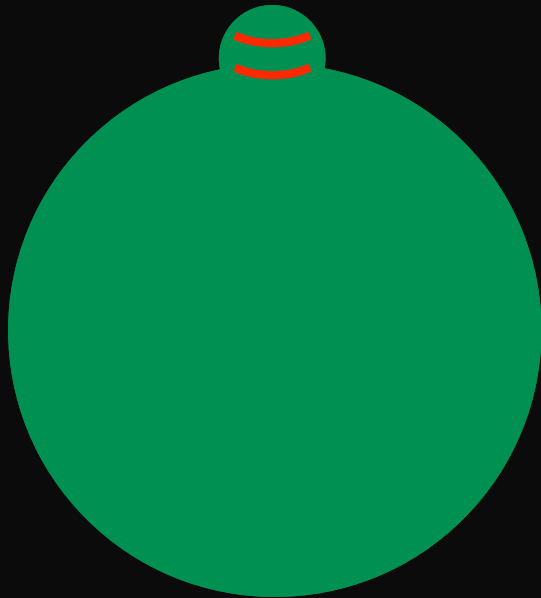
Alignment in the direction of maximal force

# Elongated organs and boundaries

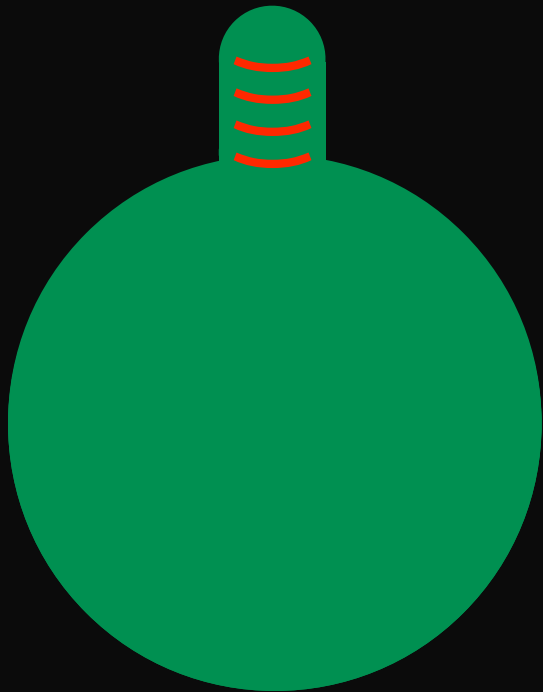




# Elongated organs and boundaries



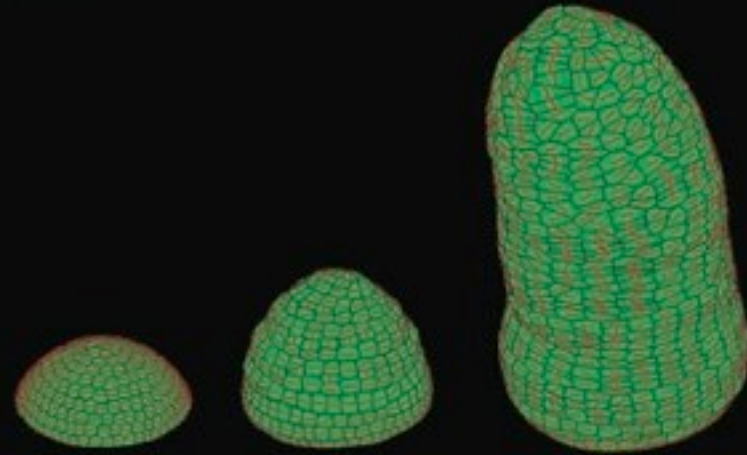
# Elongated organs and boundaries



# Elongated organs and boundaries

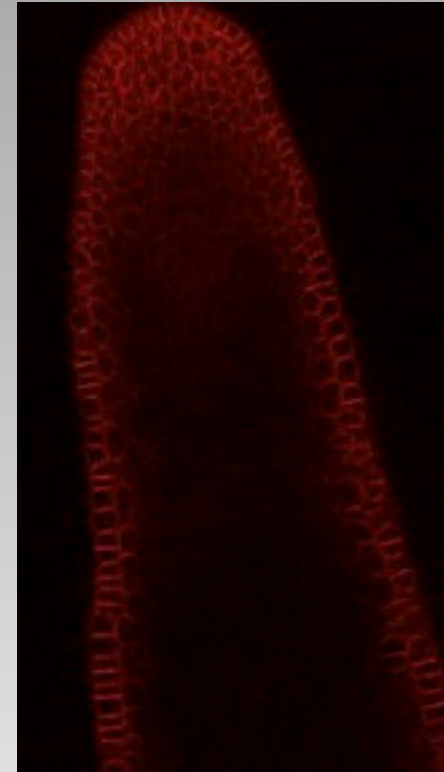
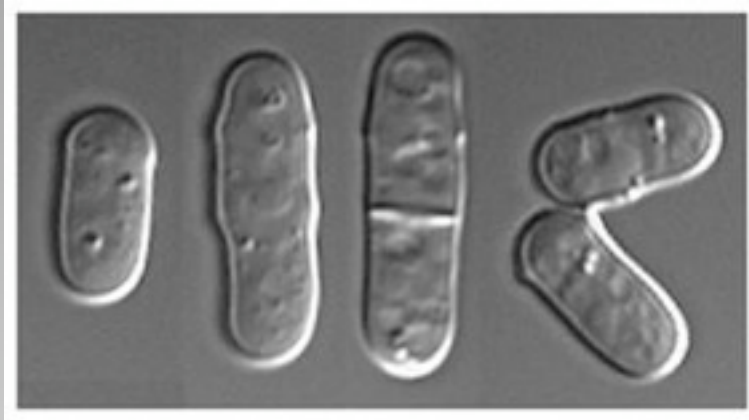


# Elongated organs and boundaries



P. Krupinski and H. Jonsson

# Conclusion



- Two strategies for anisotropic growth  
softening the tip  $\Rightarrow$  tip growth  
reinforcing
- Plant development: cellular level  $\Leftrightarrow$  morphogenesis

## Plant cells



Francis Corson  
ENS Paris, now Rockefeller University

Olivier Hamant and Jan Traas  
INRA and ENS Lyon



Yves Couder and Steffen Bohn  
Université Denis Diderot Paris



### Experimental collaborations

Elliott Meyerowitz  
Caltech

Marcus Heisler  
Caltech, now EMBL



Numerical collaborations  
Pawel Krupinski and Henrik Jonsson  
Lund University



## Fission yeast



Nicolas Minc  
Columbia University



Fred Chang  
Columbia University

*Starting collaboration with Laboratoire Joliot Curie*