

Genetic control of motor axon circuitry: a noisy computation

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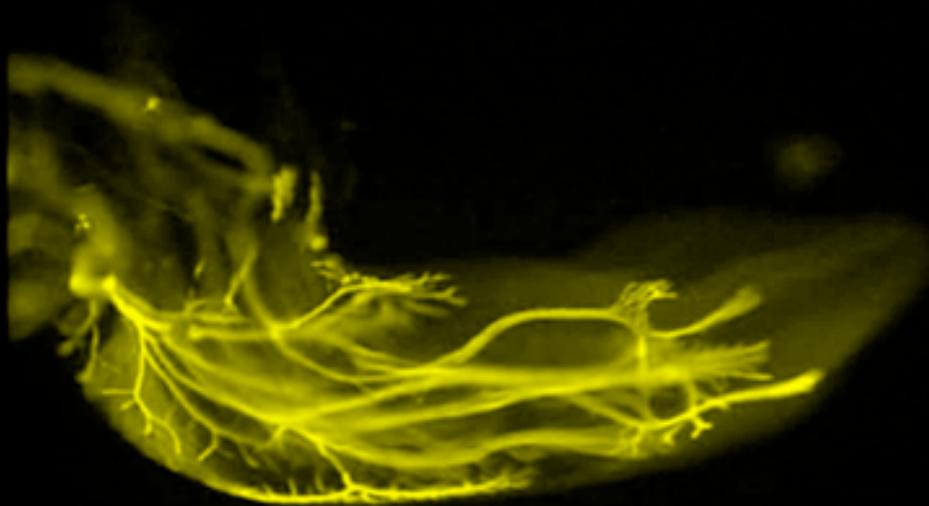
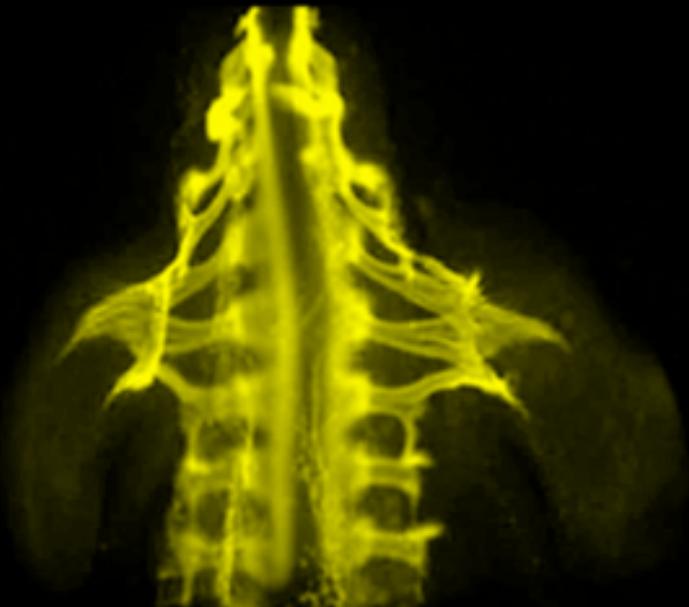
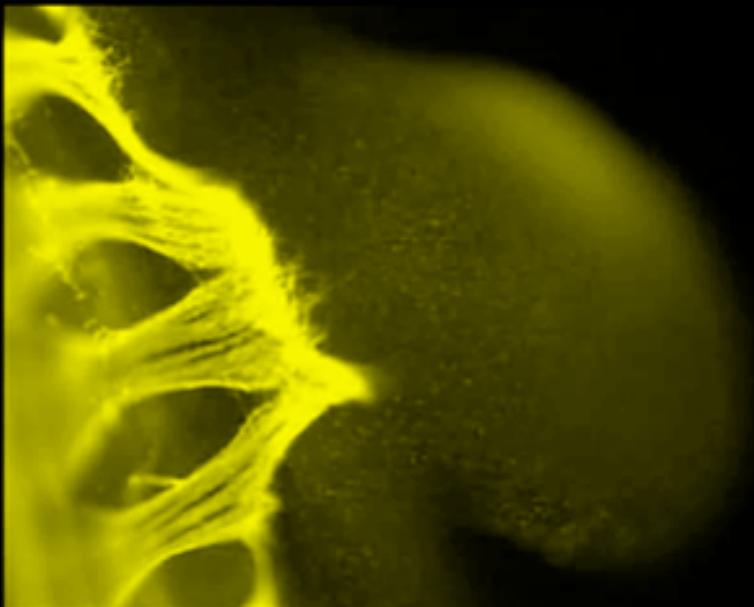
McGill & U de Montréal



Artur Kania
Dayana Krawchuk



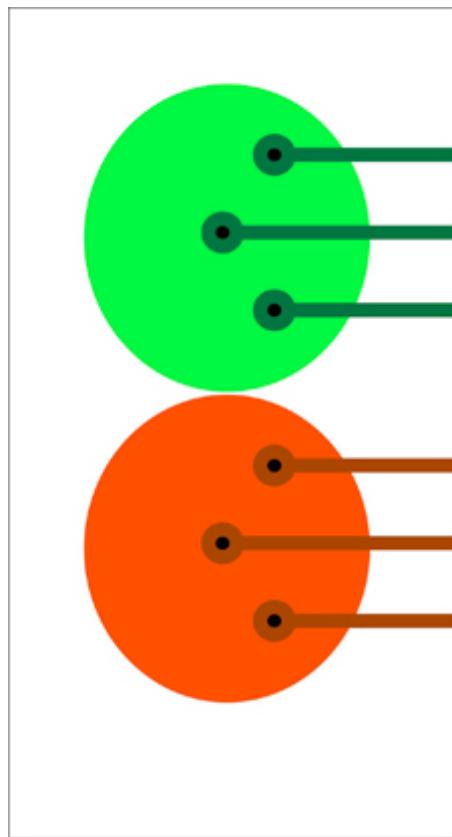
Bryan Crenshaw *Penn*
Carol Mason *Columbia*
Mark Henkemeyer *UT Southwestern*





Topographic mapping

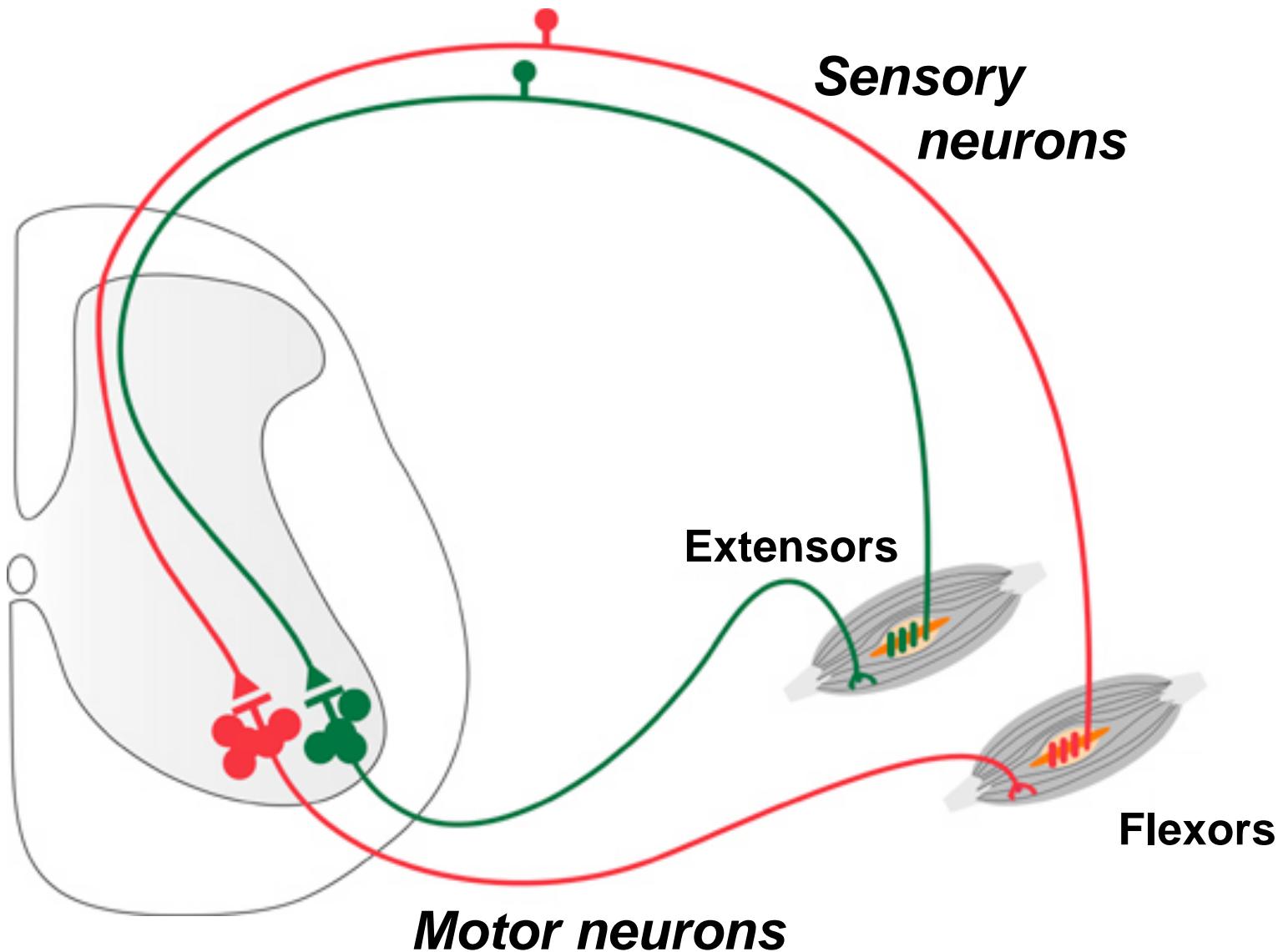
NERVOUS SYSTEM

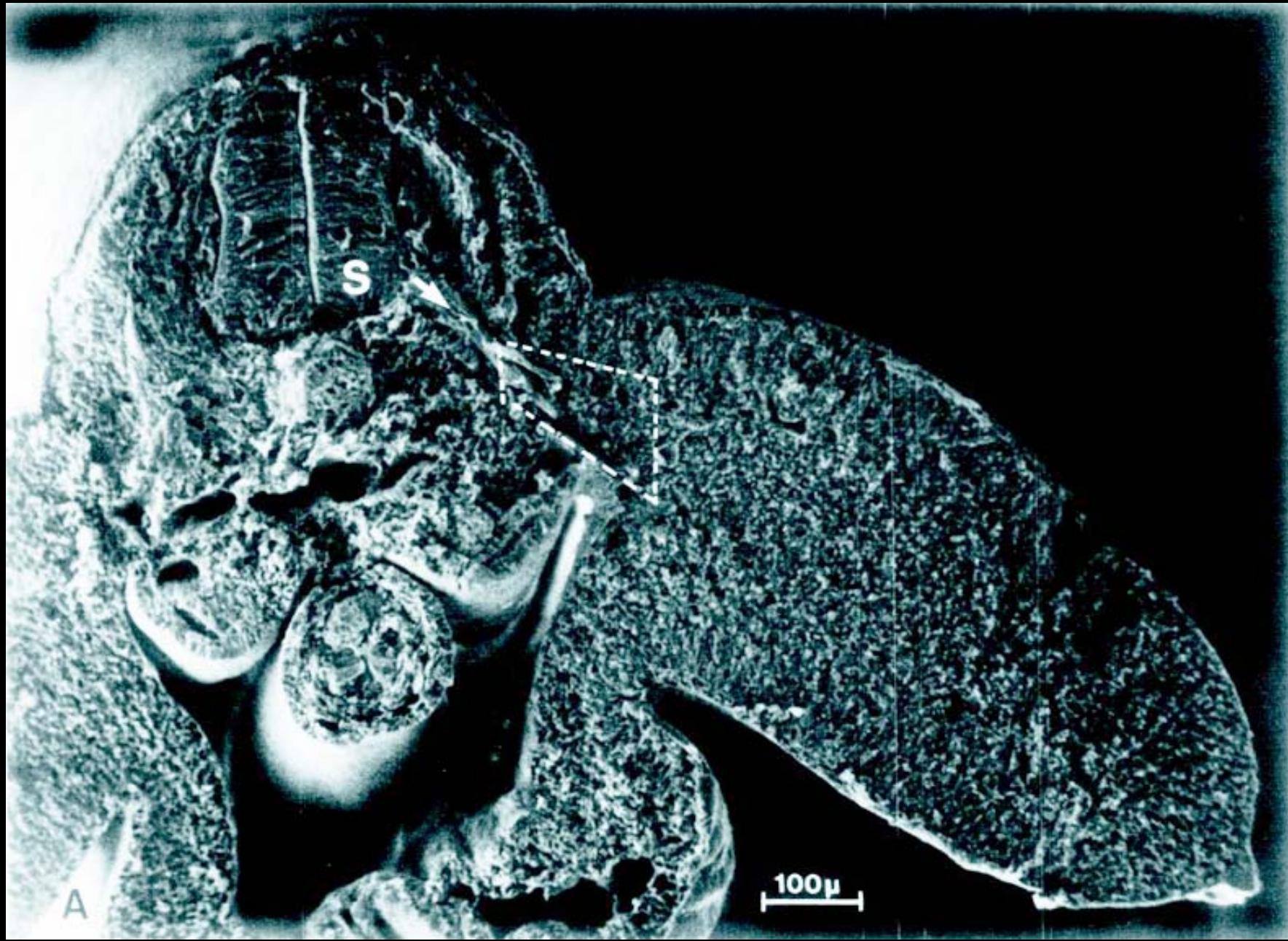


TARGET TISSUES



Motor neurons innervate target muscles



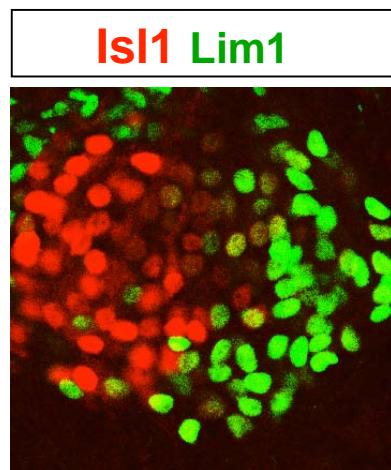


Tosney & Landmeser, 1985

Lateral Motor Column axonal projections

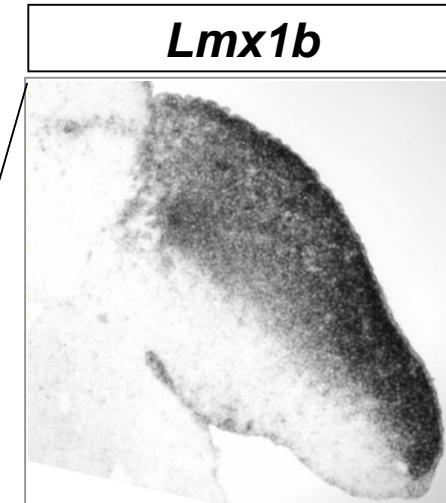
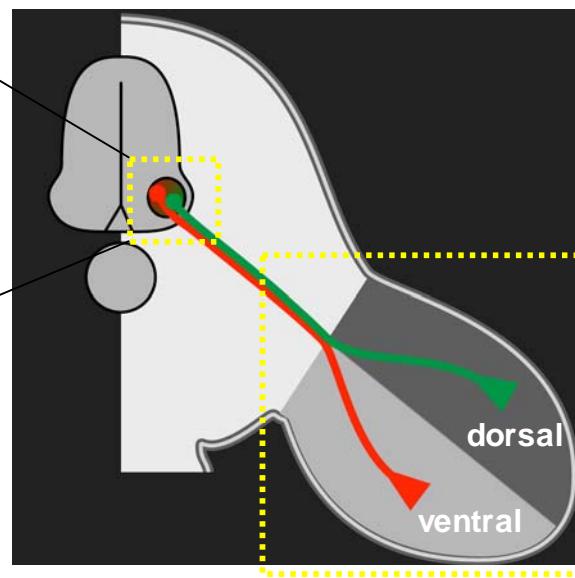
Motor column	LIM HD code
LMCI	: Lim1
LMCm	: Isl1

Limb mesenchyme	<i>Lmx1b</i>
dorsal	+
ventral	-



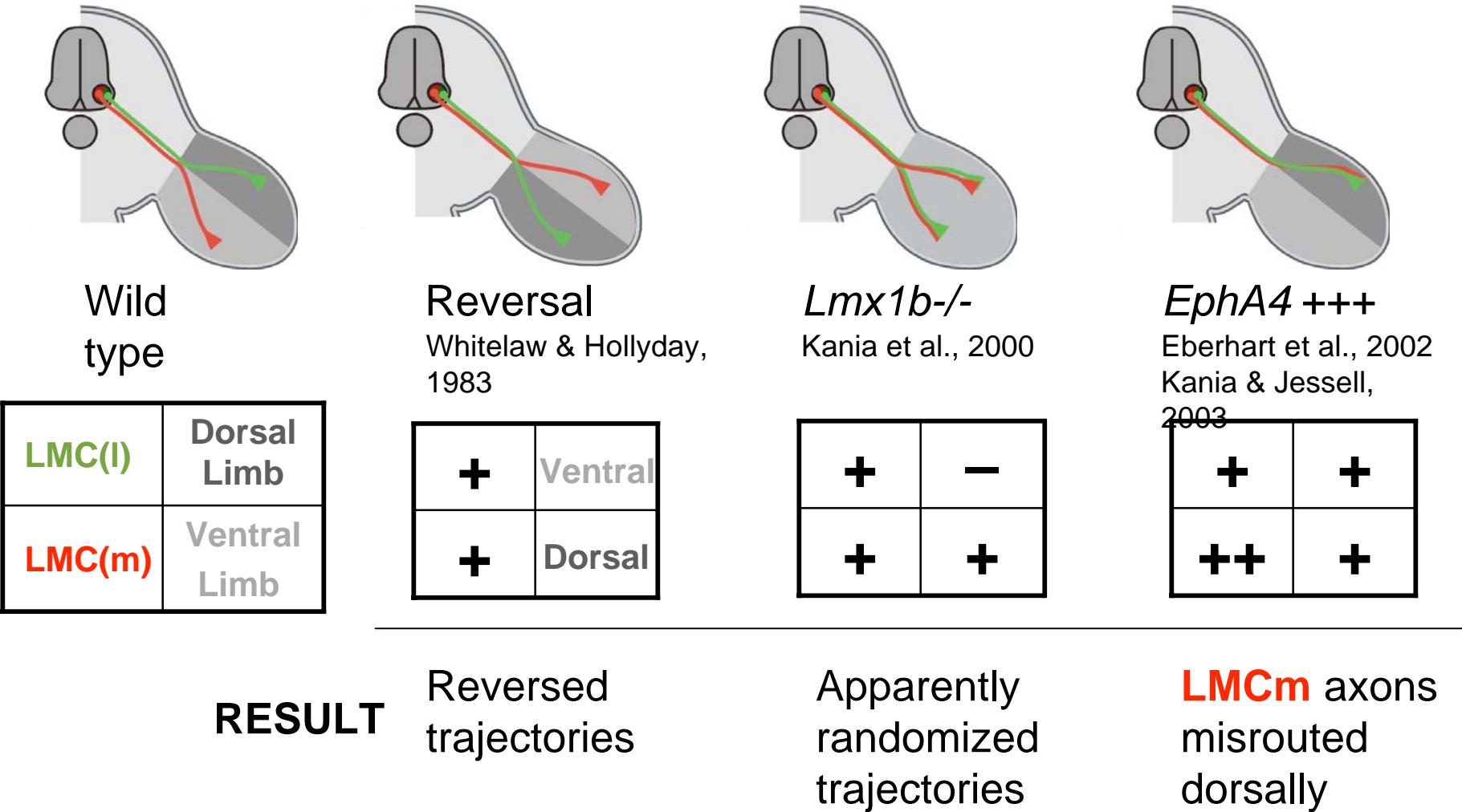
LMCm LMCI

Tsuchida et al., 1994



Riddle et al., 1995

Pathway selection is local and active

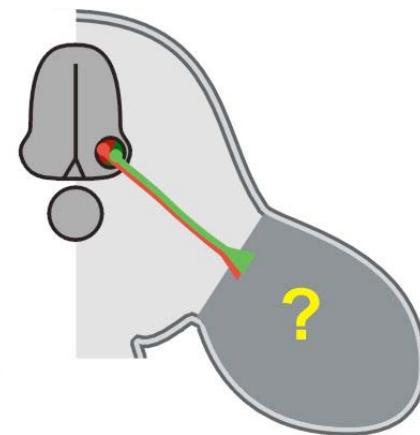
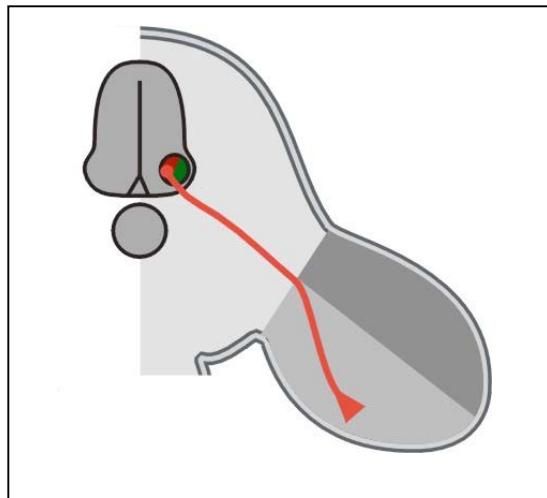


I. How are *LMCm* axons guided to the ventral limb?

II. What are the effector molecules that control *LMCm* trajectories?

III. Quantitative models, experimental predictions and tests

I. How are *LMCm* axons guided to the ventral limb?

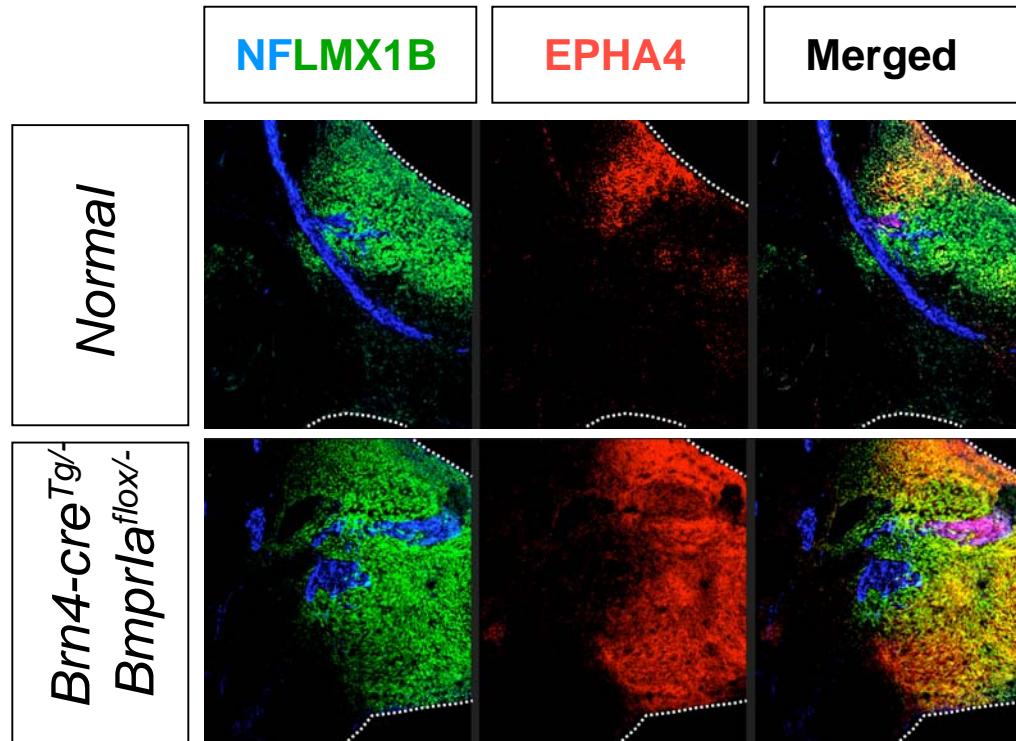


Bidorsal

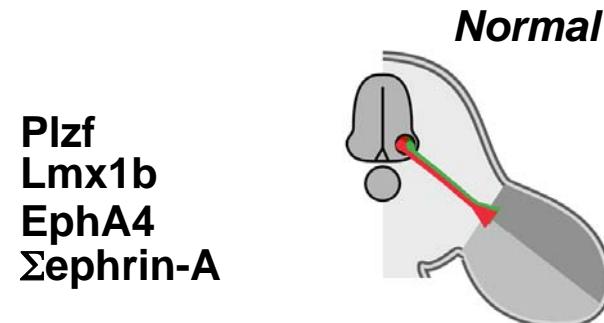
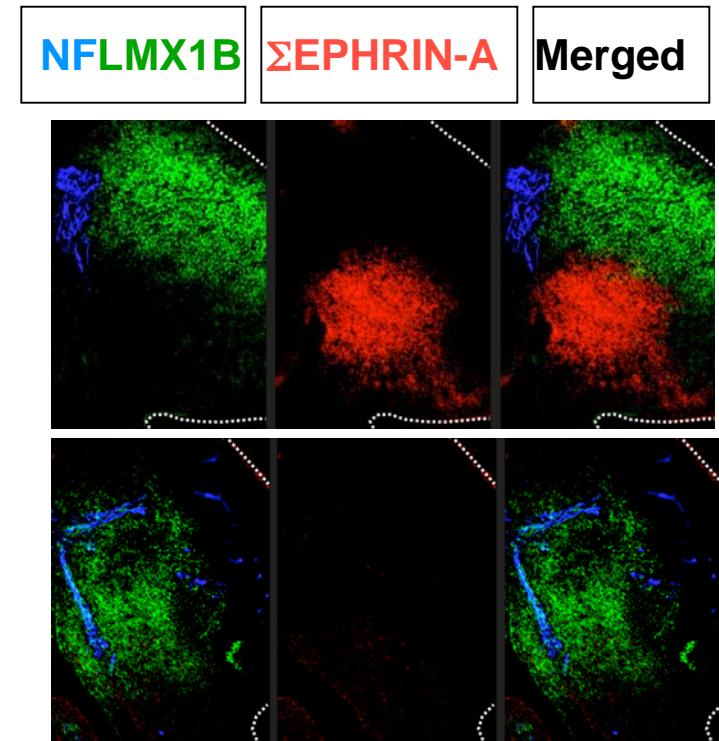
Bmp receptor Ia mouse conditional mutant
limbless chick mutant

Bmprla^{flox/-} hindlimbs are bidorsal

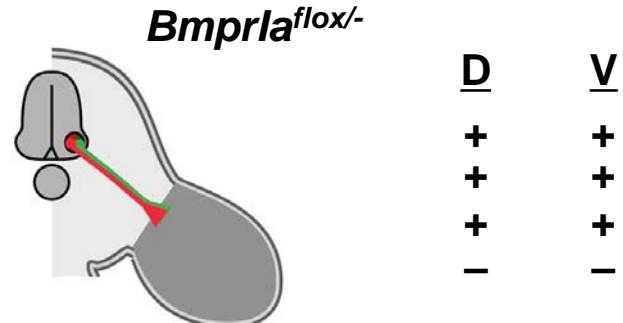
Dorsal markers are expanded



Ventral markers are absent

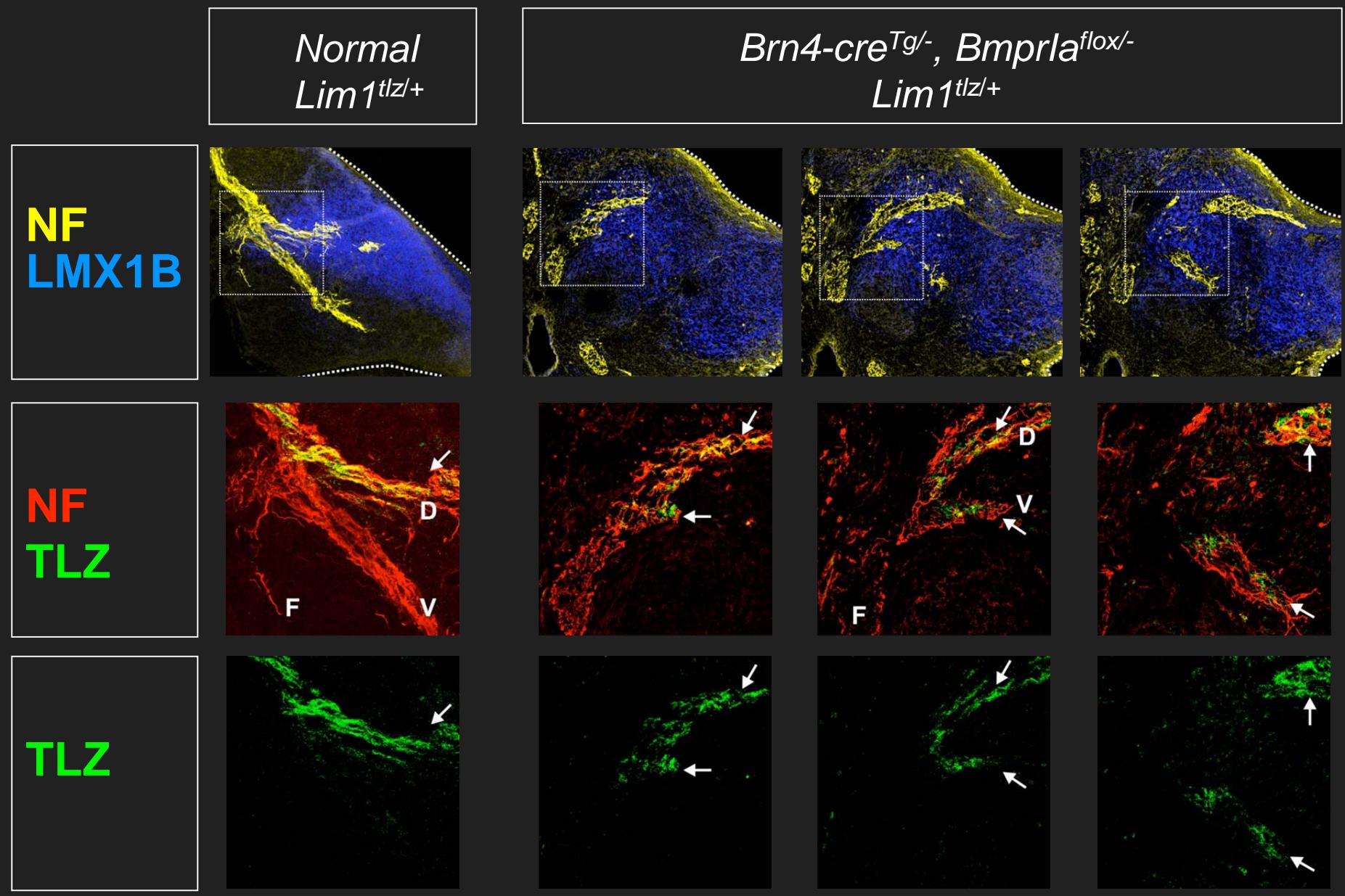


	D	V
-	+	+
+	-	-



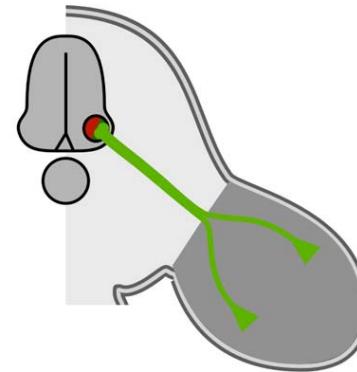
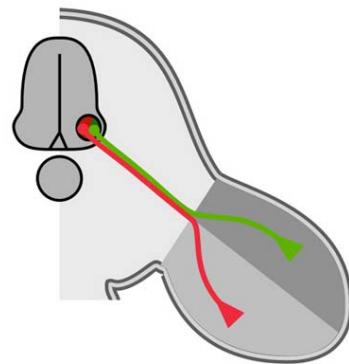
	D	V
-	+	+
+	-	+
-	-	-

LMCI populates both limb nerve branches in mutants



WT

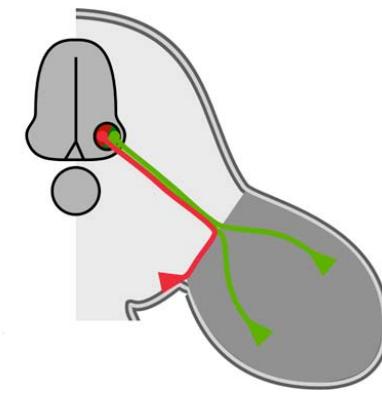
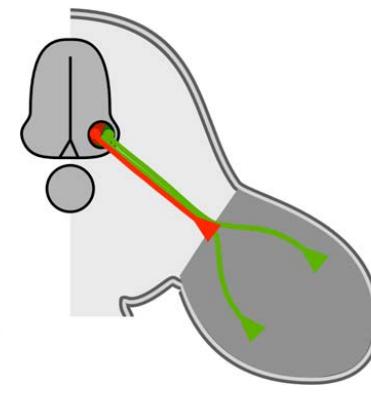
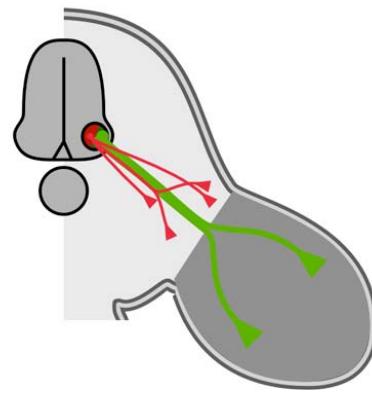
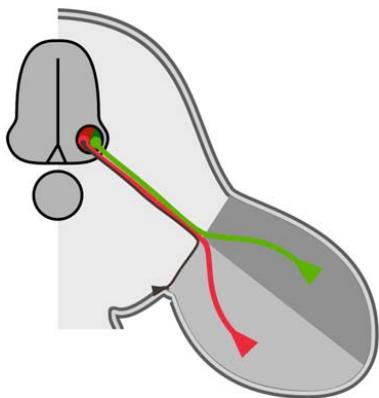
Brn4-cre^{Tg/-}, Bmprla^{flox/-}



*Where do the **LMCm** axons project?*

WT

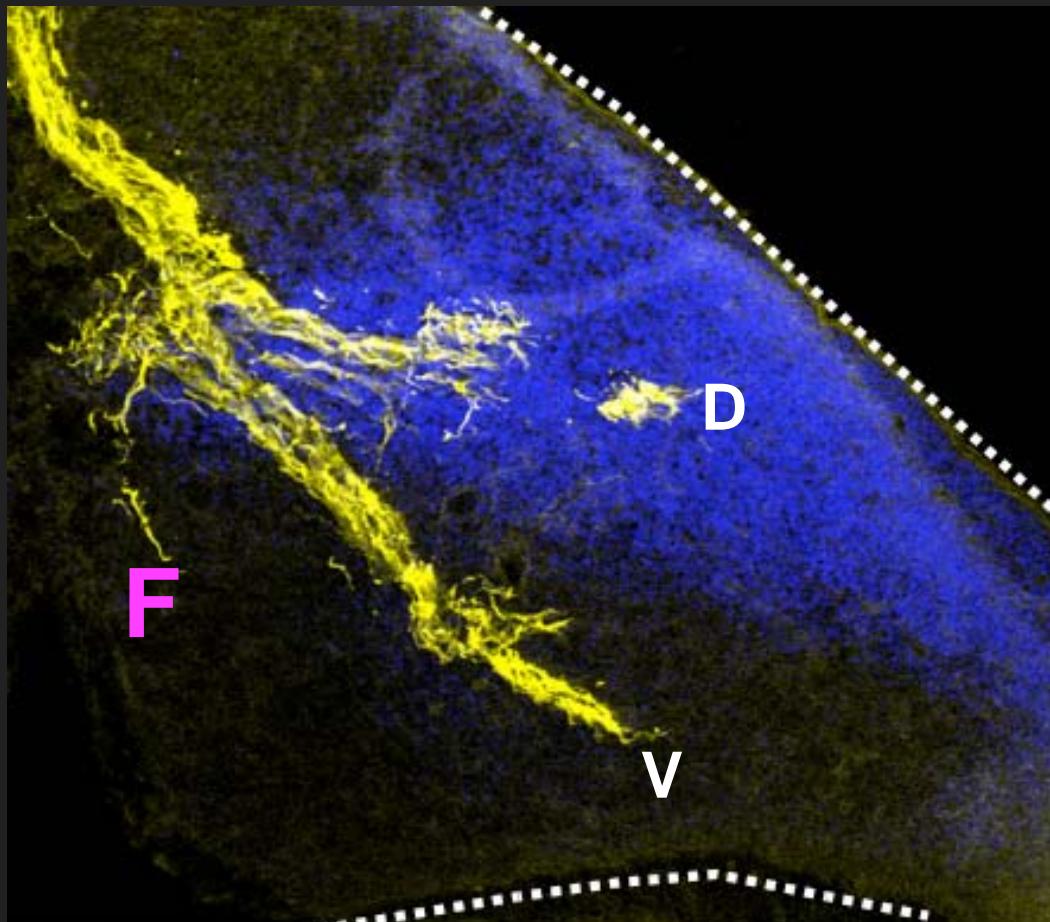
Brn4-cre^{Tg/-}, Bmprla^{flox/-}



DISORGANIZED? STALLED? REDIRECTED?

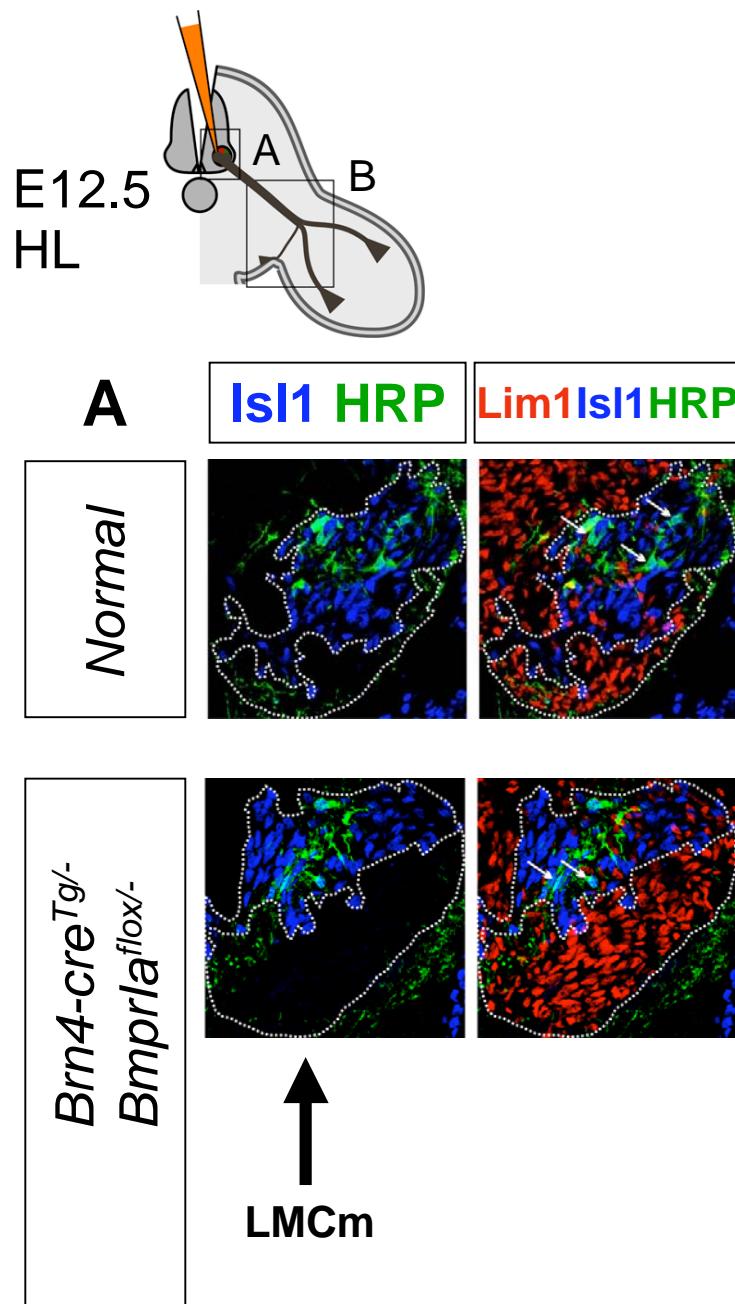
A third nerve branch innervates the flank

Nerves Dorsal limb

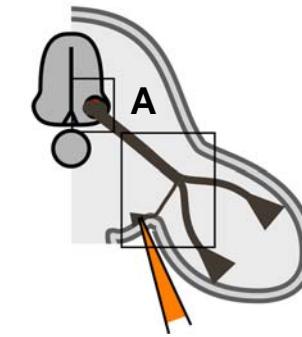


HL
E11.75

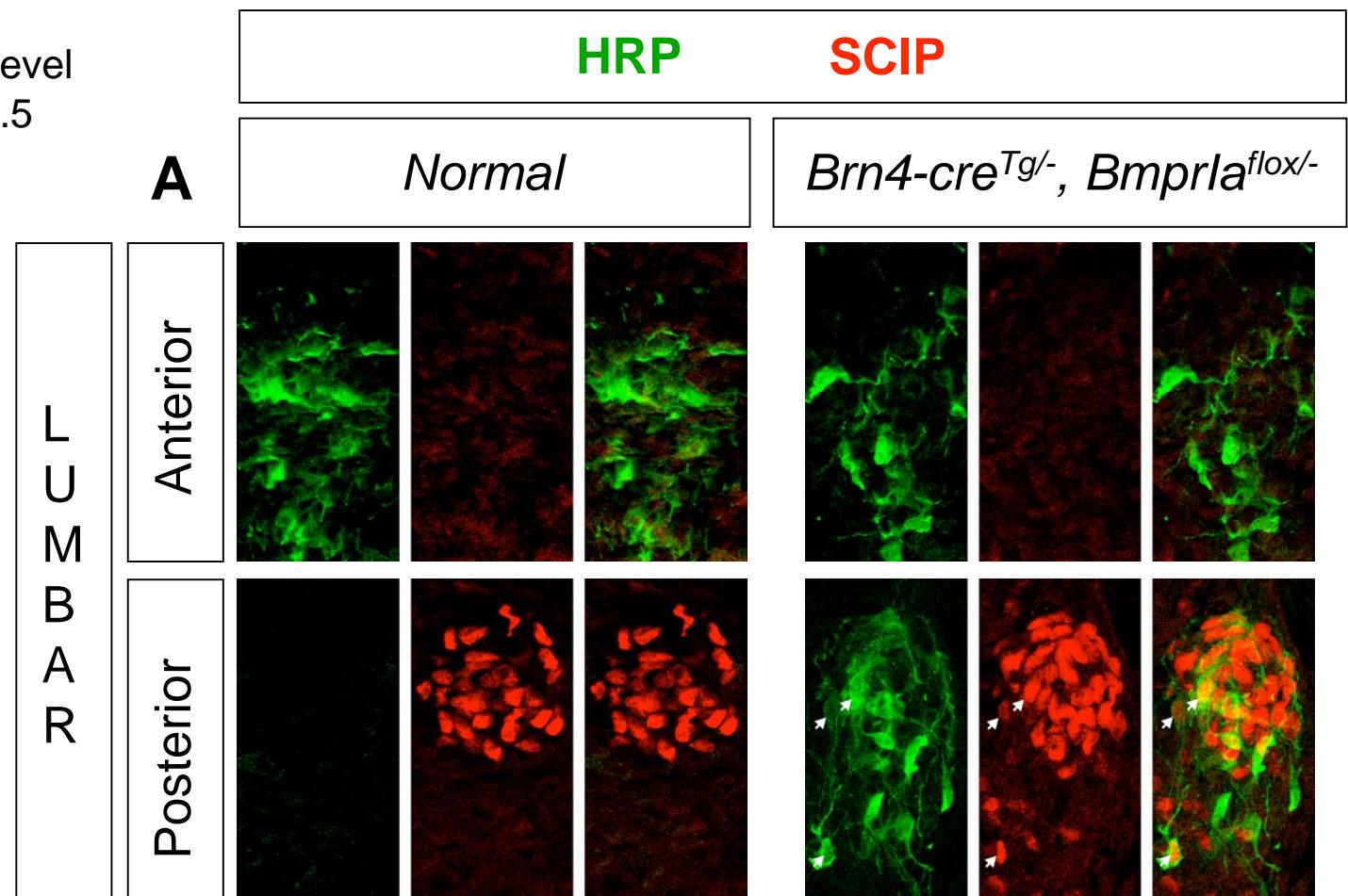
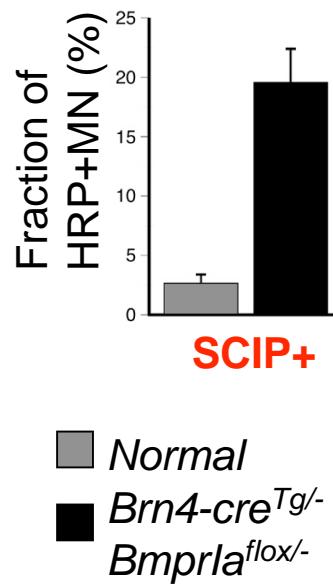
Mutant LMCm axons populate only the FLANK branch



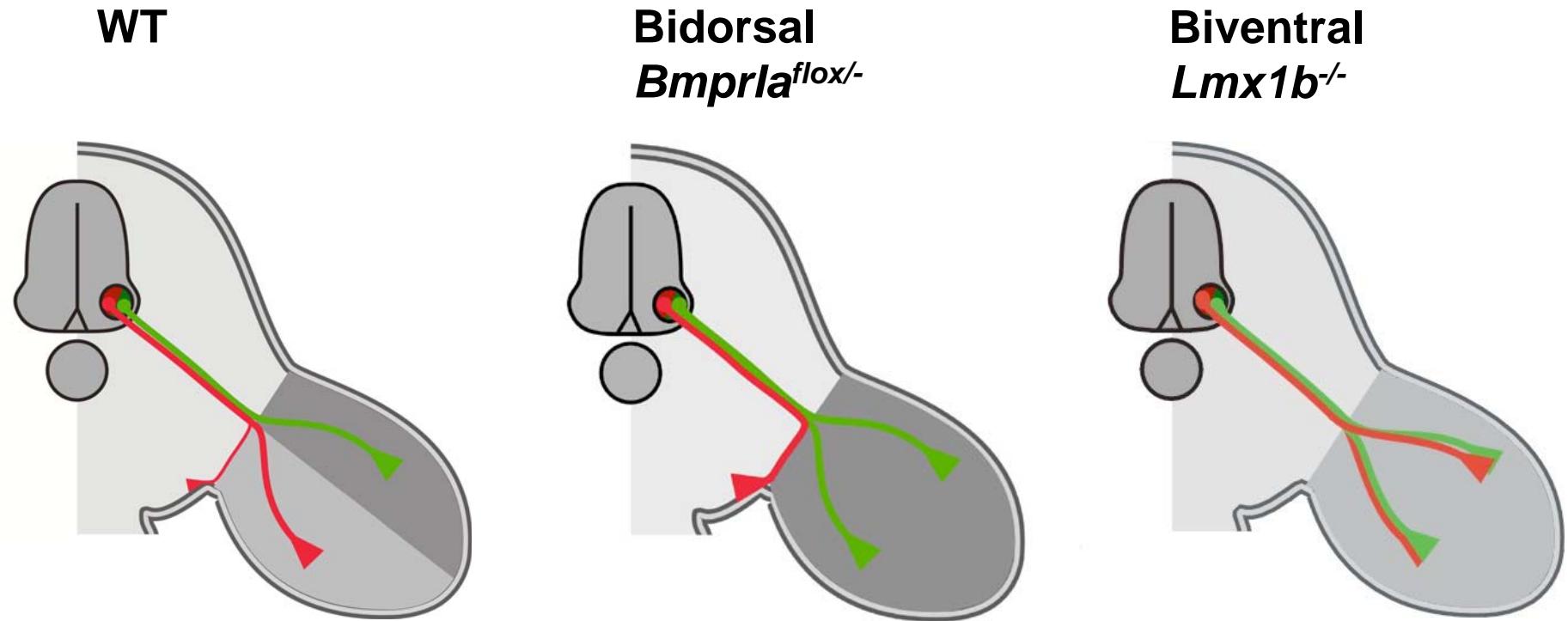
Limb-targeted LMCm axons are *redirected* to flank



HL level
E13.5

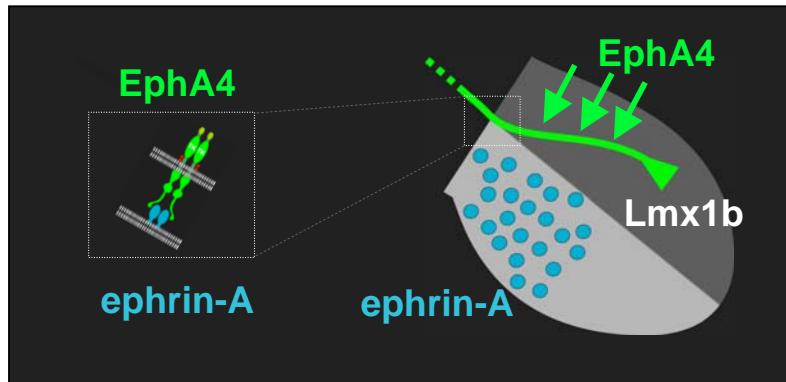


1. LMCm and LMCI have *non-symmetrical* preferences.



2. NEW TARGET: LMCm also innervate non-limb tissues.

II. What are the effector molecules that control LMCI trajectories?

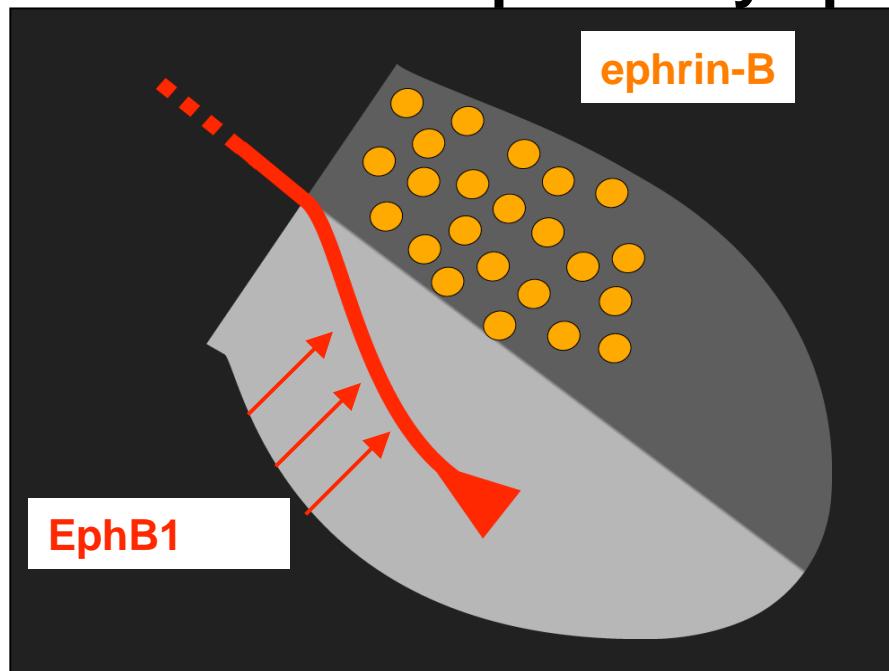


ephrin-A/EphA, GDNF/c-ret, Sema/Npn signaling influence LMCI trajectories

Kania et al., 2000
Helmbacher et al, 2000
Eberhart et al, 2002
Kania and Jessell, 2003
Huber et al., 2005
Kramer et al., 2006

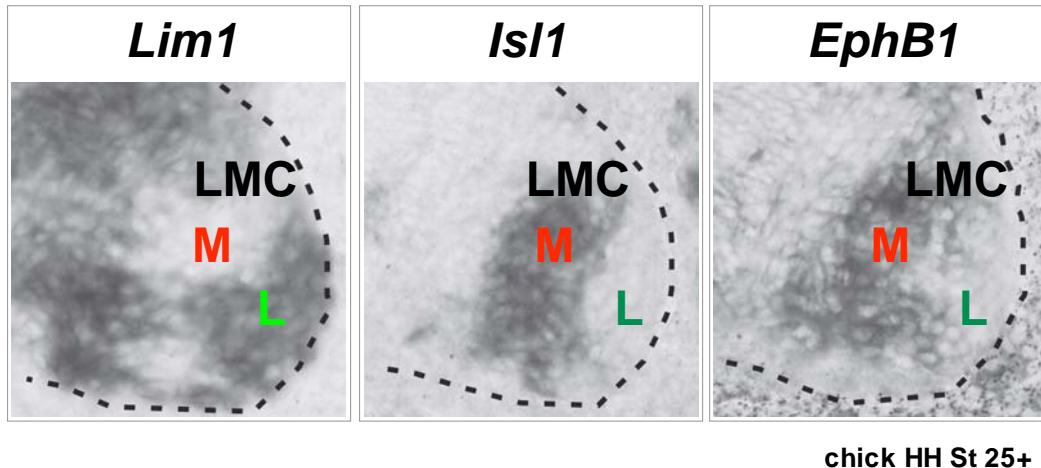
Hypothesis

EphB+ LMCM axons are repelled by ephrin-B+ dorsal limb



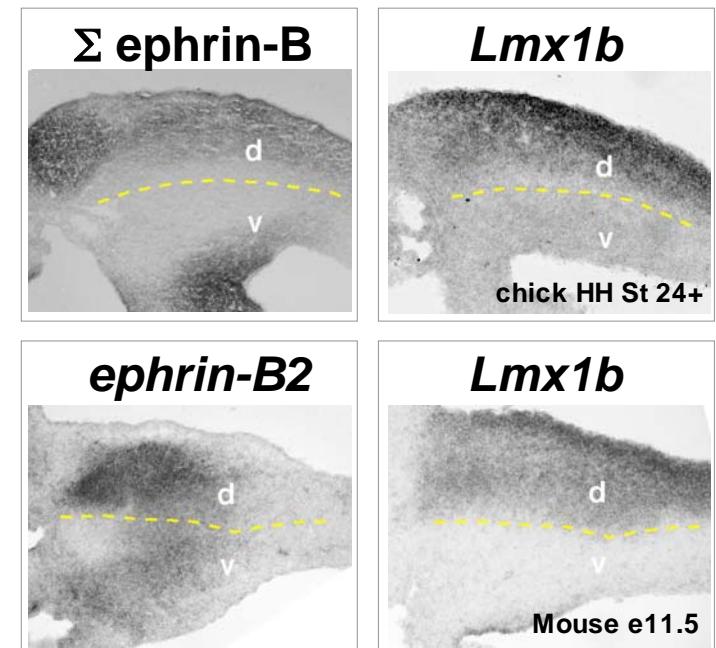
RECEPTORS

EphB1 in **medial LMC** neurons

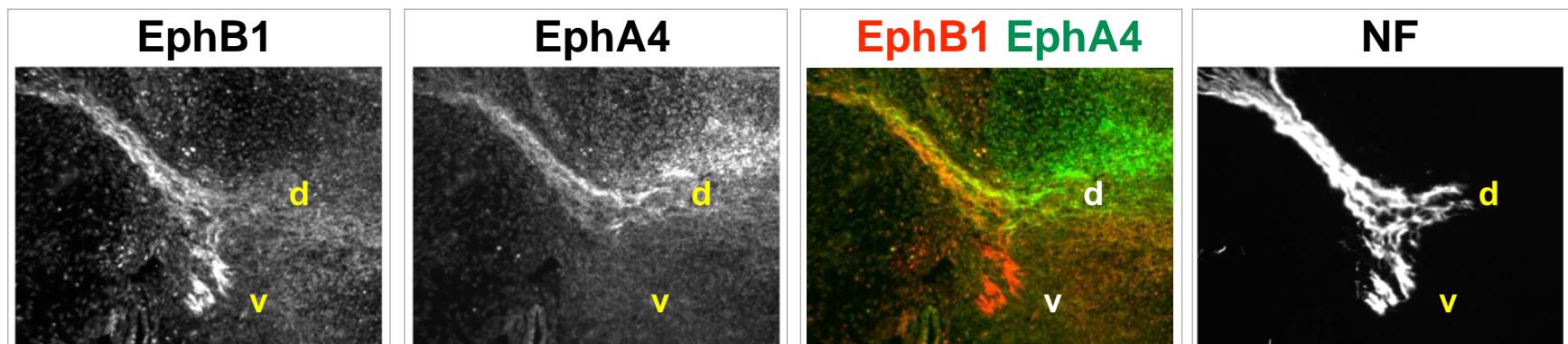


LIGANDS

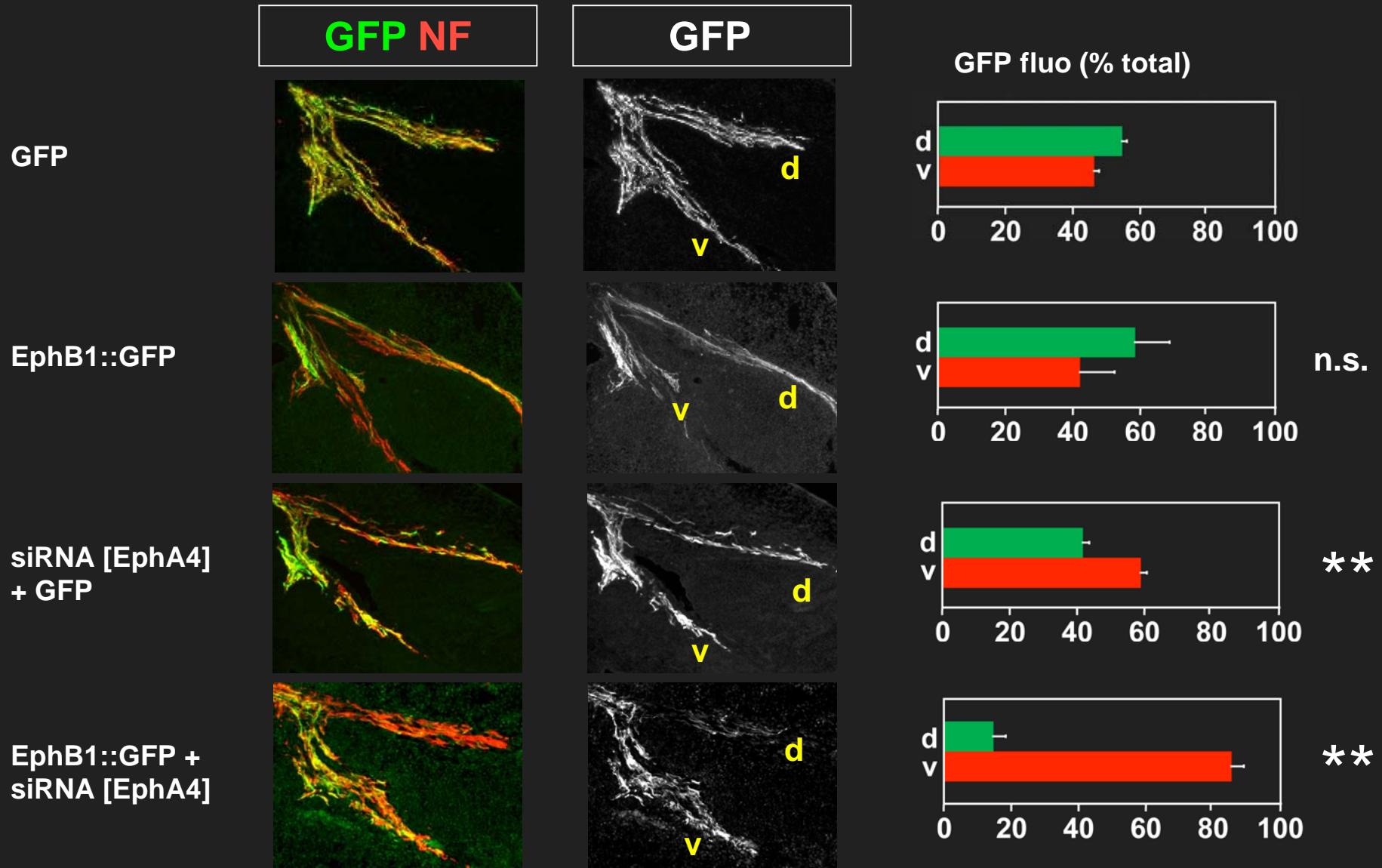
ephrin-B in dorsal limb



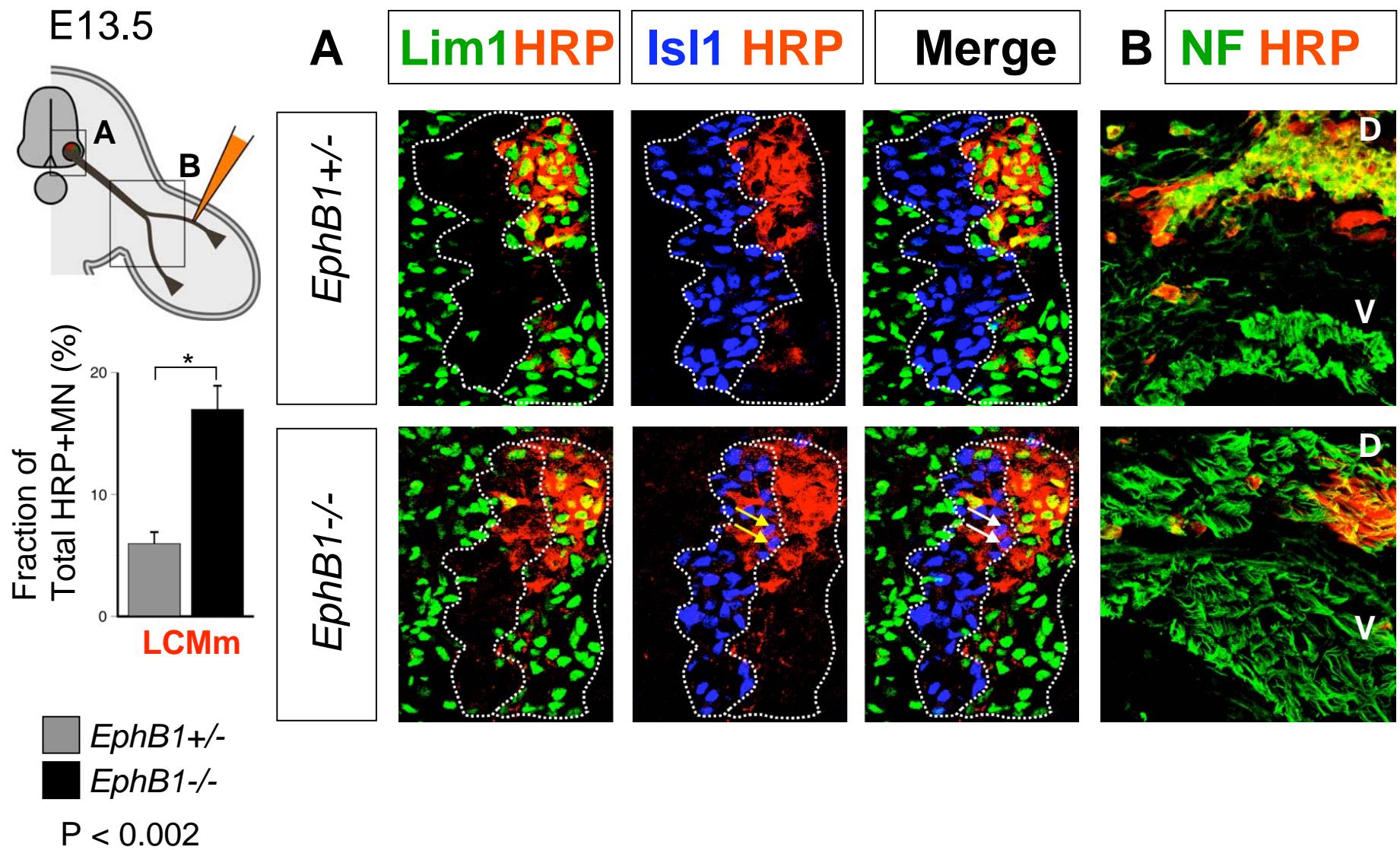
Limb nerve



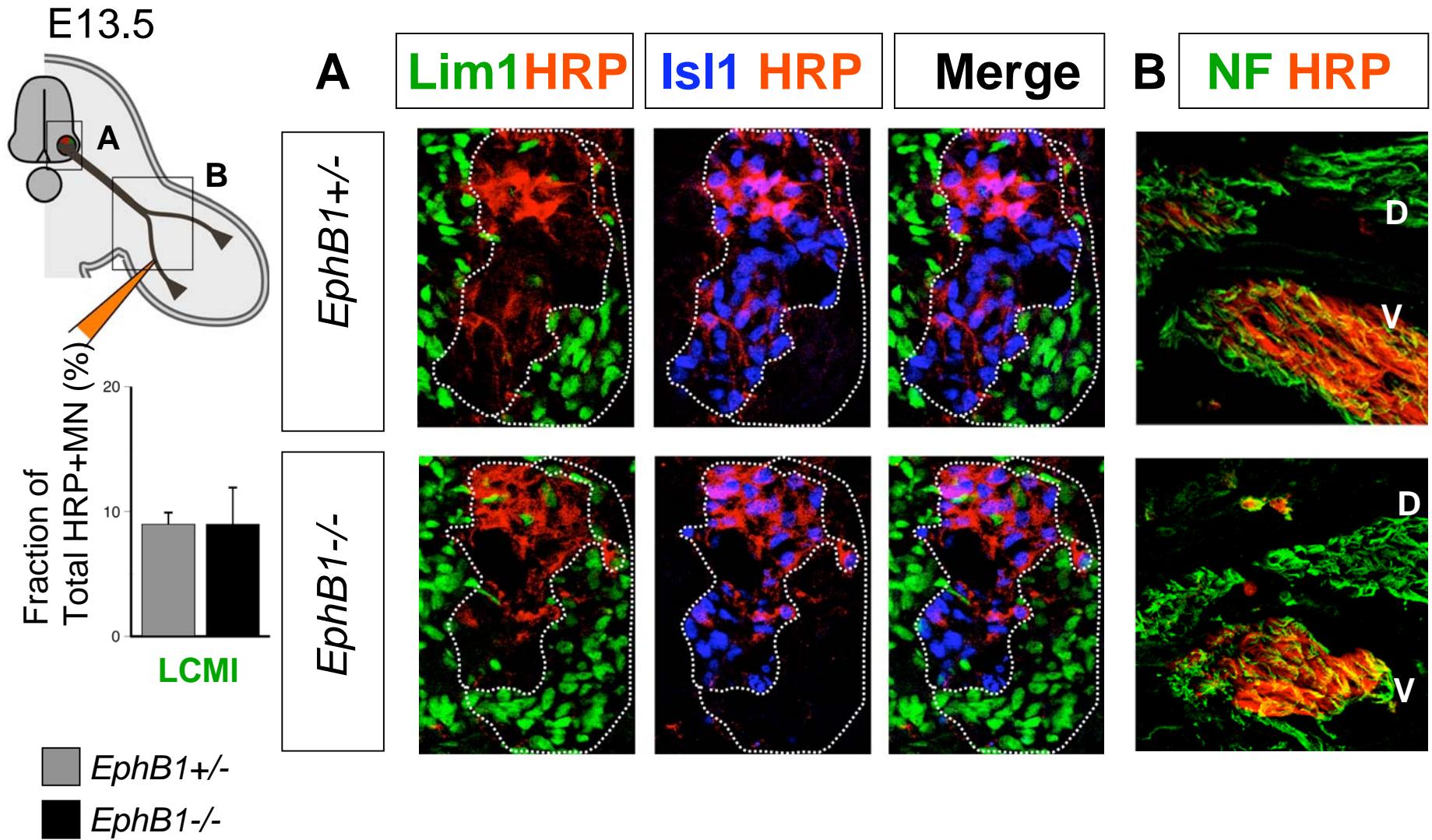
EphB1 expression can *redirect* LMC axons to the ventral limb



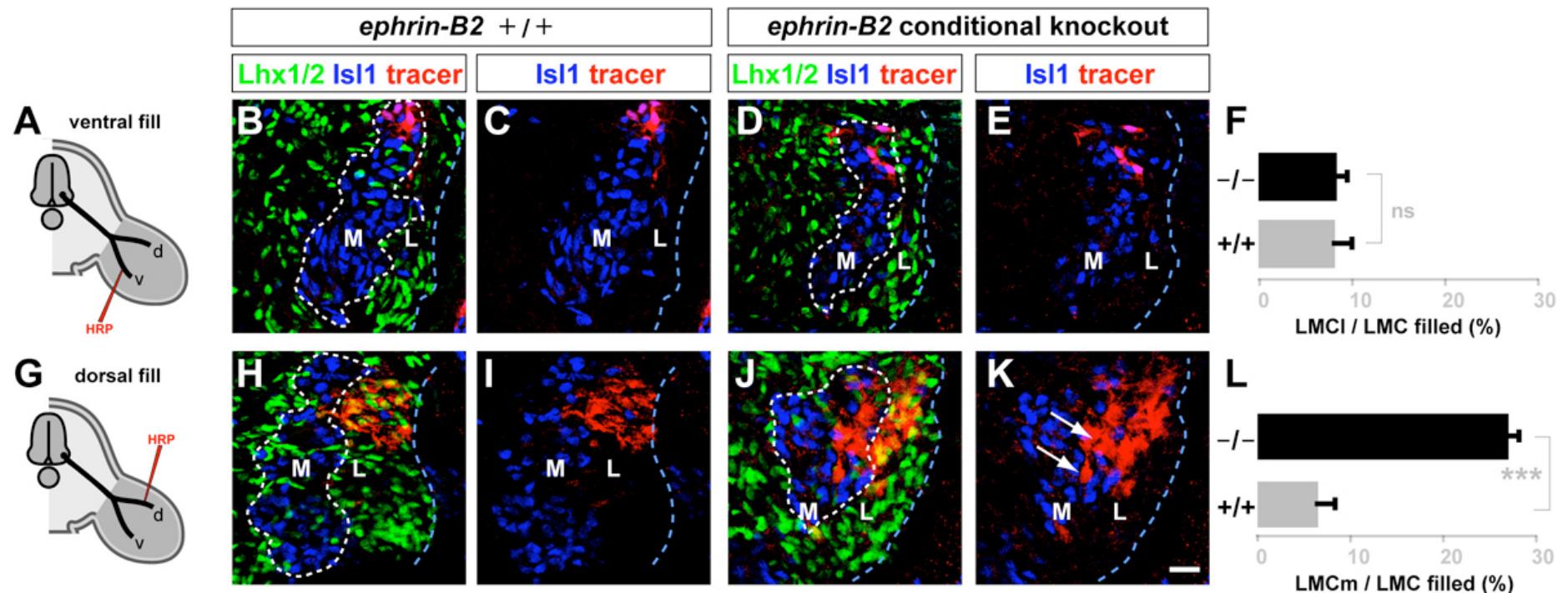
EphB1 is necessary for LMCm ventral targeting



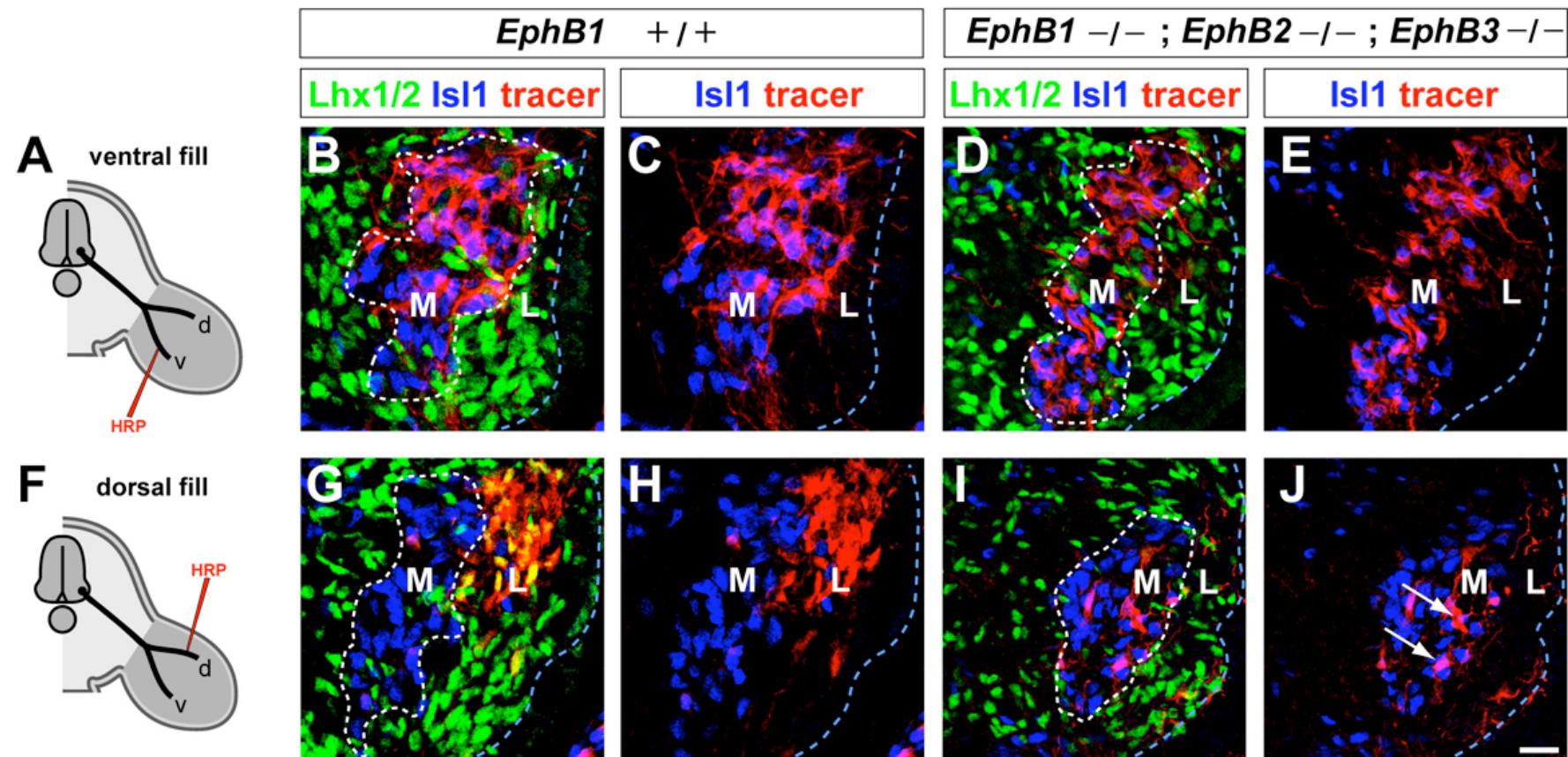
EphB1 does not influence LMCI projections



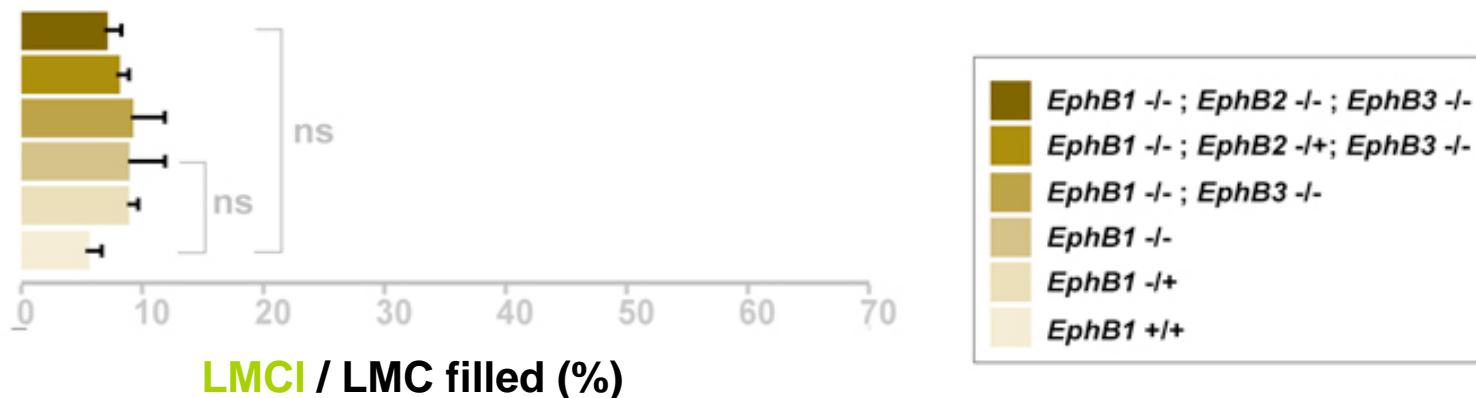
ephrin-B2 is necessary for LMCm ventral targeting



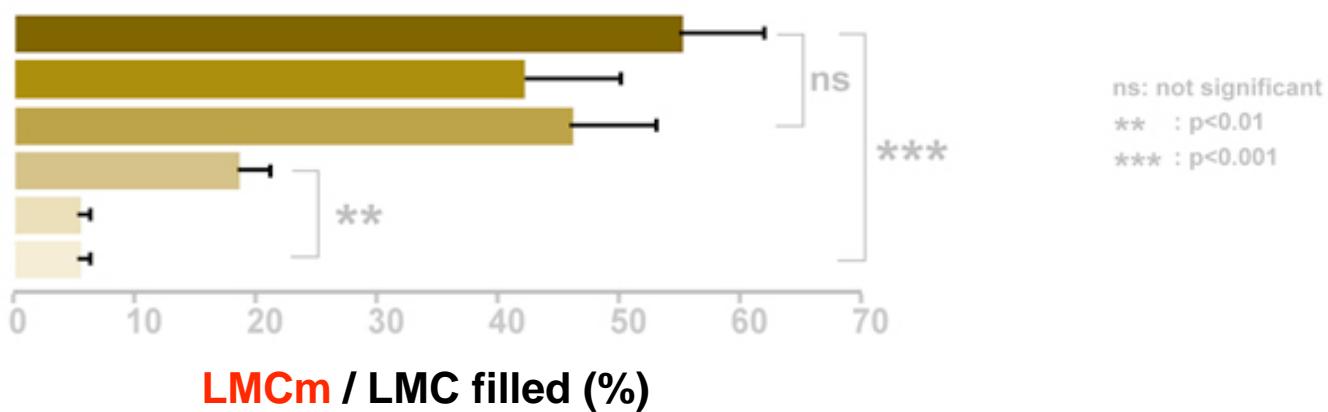
Three *EphB* receptors target LMCM axons ventrally



ventral fill



dorsal fill

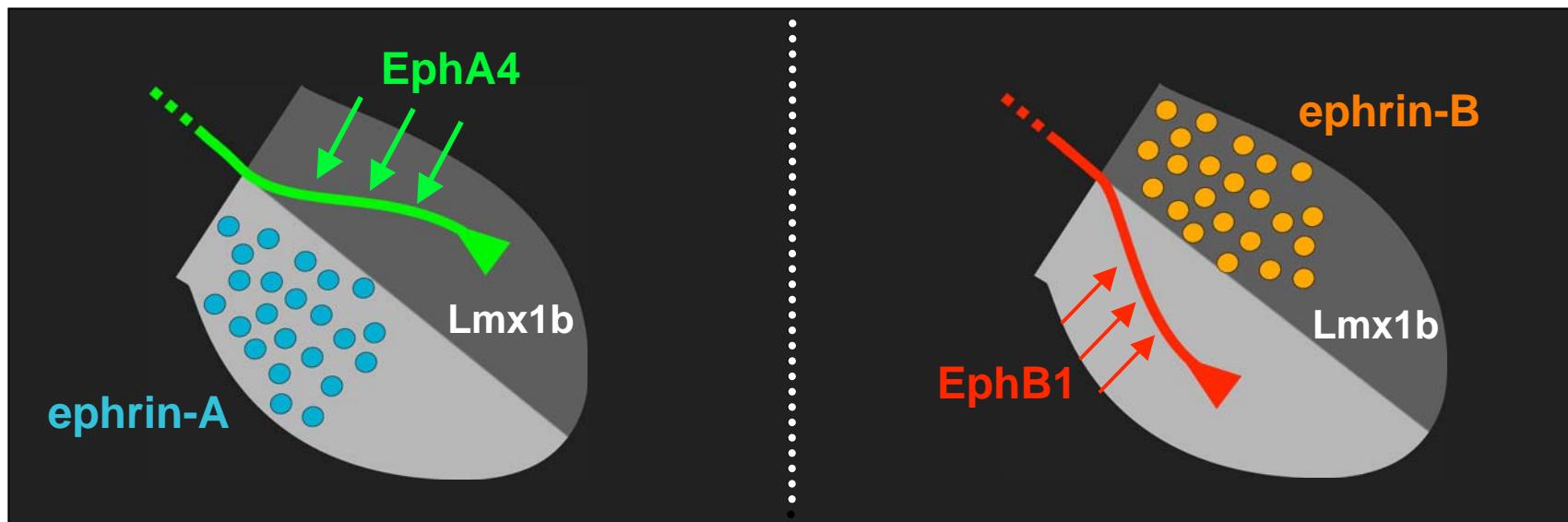


Mirror symmetry ephrin-Eph signaling controls motor axon trajectories to the limb

EXPRESSION

Dorsal limb	b	A
Ventral limb	a	B

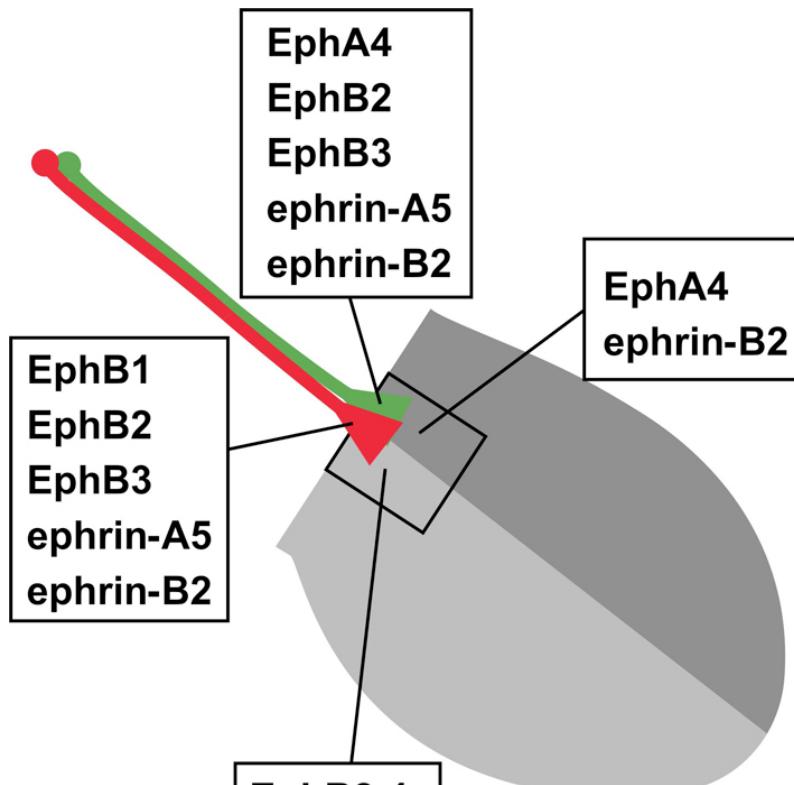
FUNCTIONAL



LMCI → EphA4 → Dorsal limb **LMCm → EphB1 → Ventral limb**

III. Quantitative models, experimental predictions and tests

Axons integrate multiple cues at limb entry

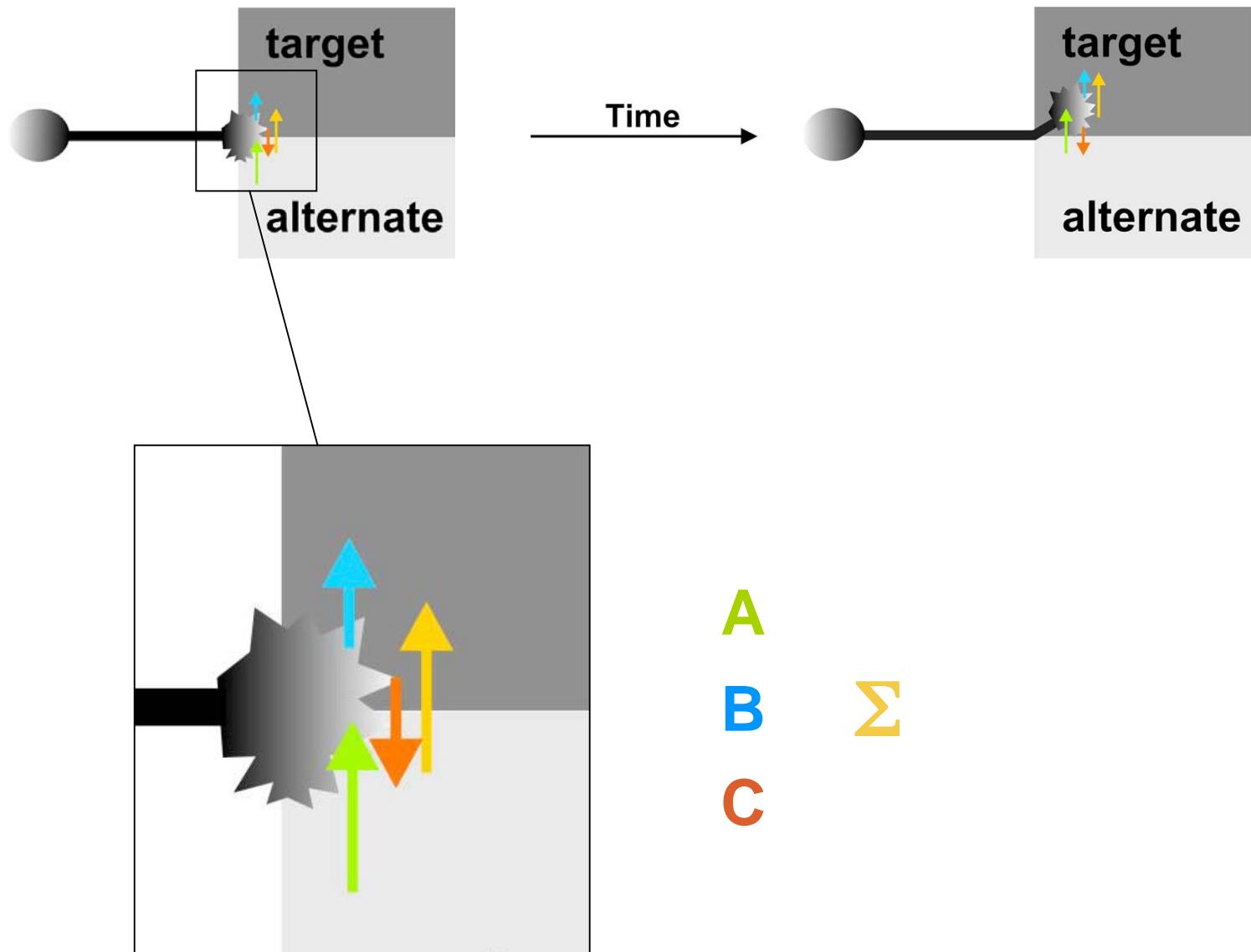


Axon	Interaction	Limb	Symbol
EphB1	>—<	ephrin-B2	B ₁ b ₂
EphB2	>—<	ephrin-B2	B ₂ b ₂
EphB3	>—<	ephrin-B2	B ₃ b ₂
EphB2	>—<	ephrin-As	B ₂ a
EphA4	>—<	ephrin-As	A ₄ a
EphA4	>—<	ephrin-B2	A ₄ b ₂
ephrin-B2	<—>	EphB3,4	b ₂ B ₃₄
ephrin-A5	<—>	EphA4	a ₅ A ₄
ephrin-B2	<—>	EphA4	b ₂ A ₄

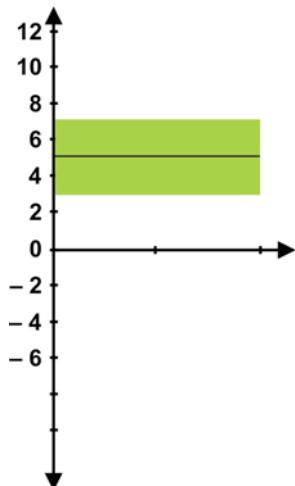
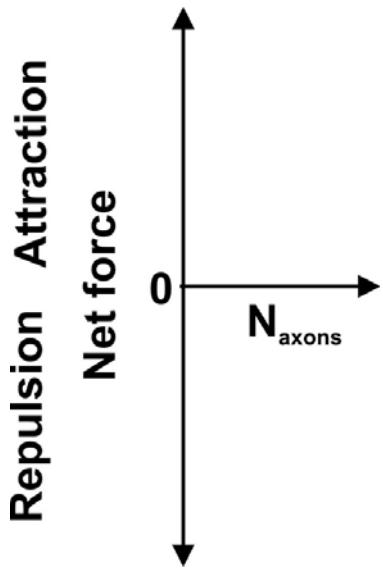
$$I-D = A_4a + B_2a + a_5A_4 + b_2A_4 - A_4b_2 - B_2b_2 - B_3b_2 - b_2B_{34} > 0$$

$$m-V = -B_1b_2 - B_2b_2 - B_3b_2 - b_2B_{34} + B_2a + a_5A_4 + b_2A_4 < 0$$

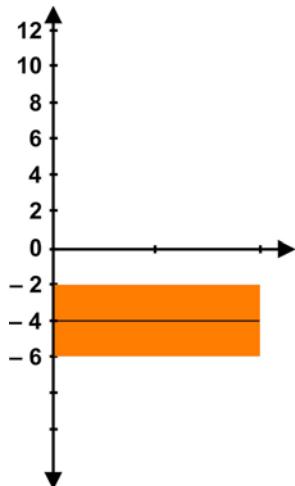
Axons integrate multiple cues at choice points



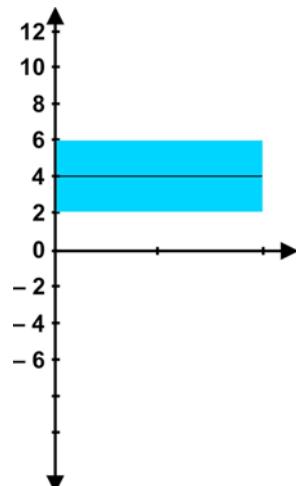
Cues are noisy



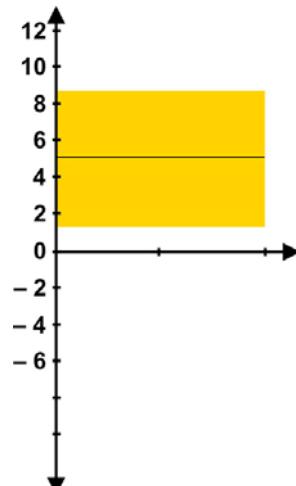
↑ A
 $A = 5 \pm 1$



↓ C
 $C = -4 \pm 1$



↑ B
 $B = 4 \pm 1$

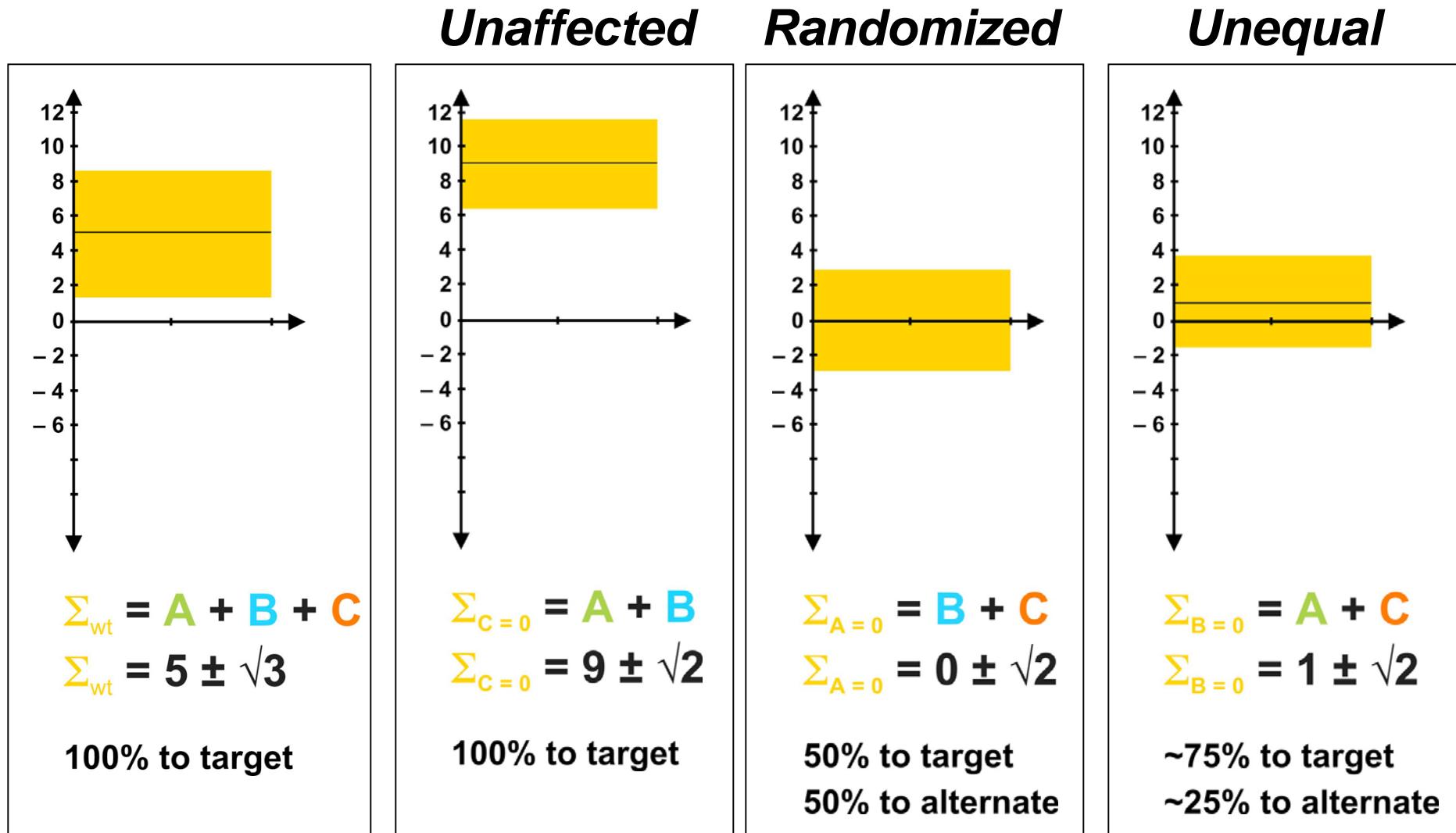


↑ $\Sigma_{\text{wt}} = A + B + C$
 $\Sigma_{\text{wt}} = 5 \pm \sqrt{3}$

Cue noise can explain guidance defects of mutants

WILD TYPE

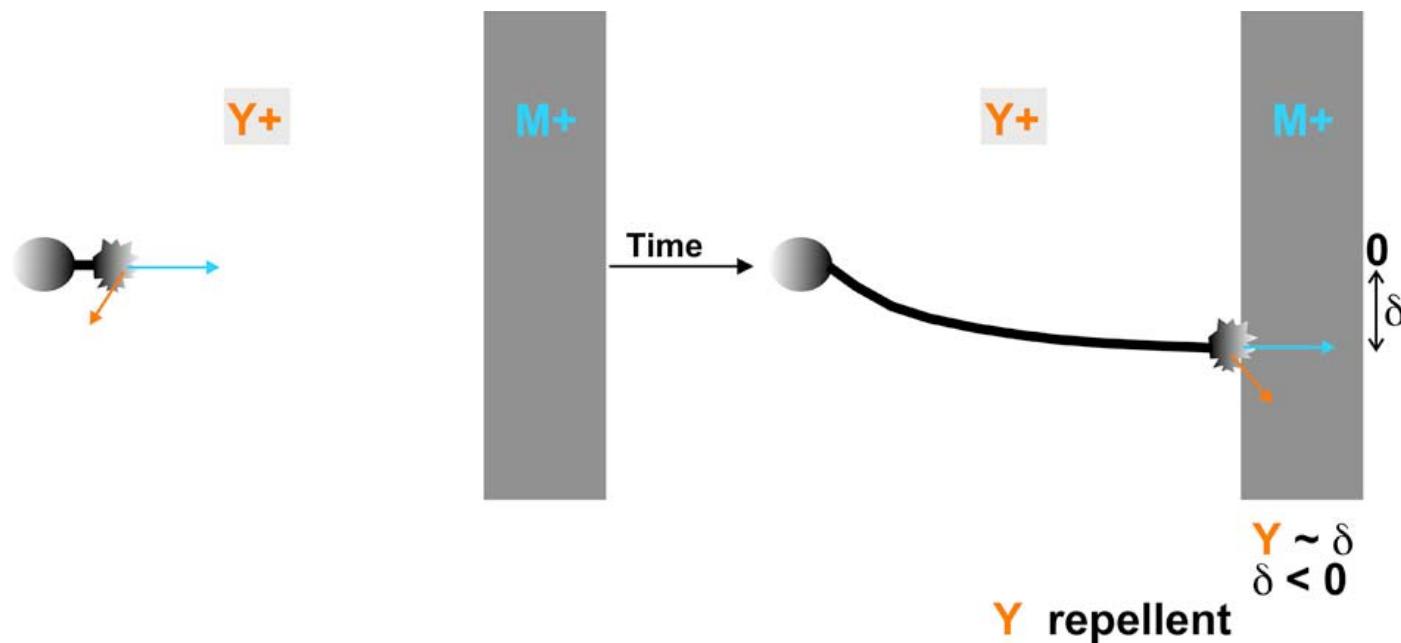
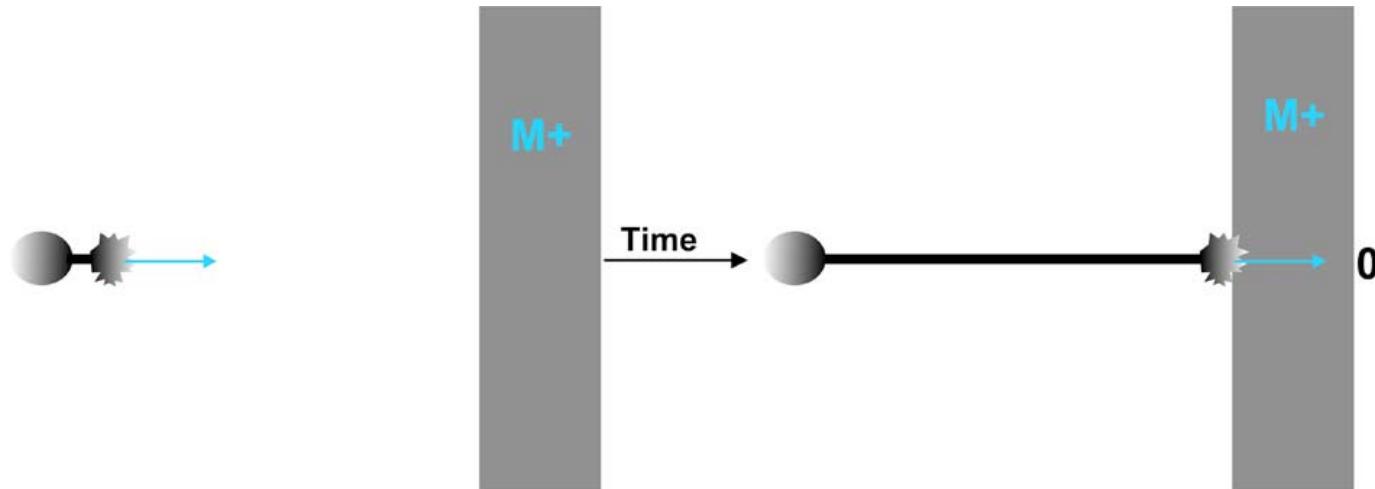
MUTANTS



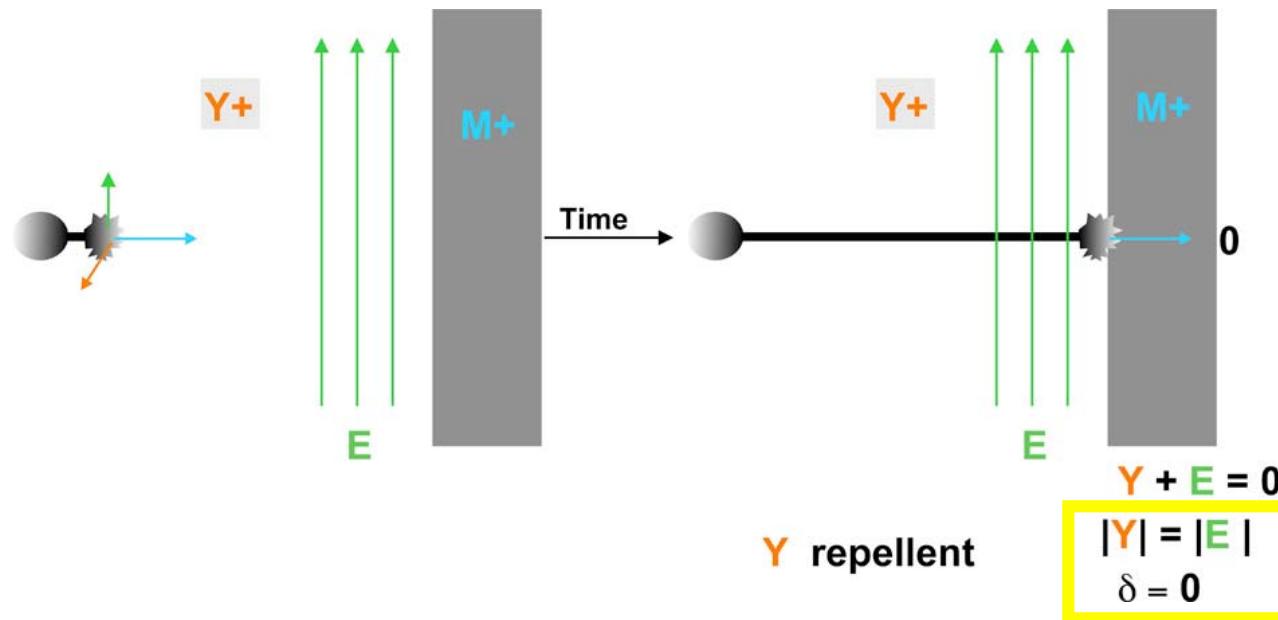
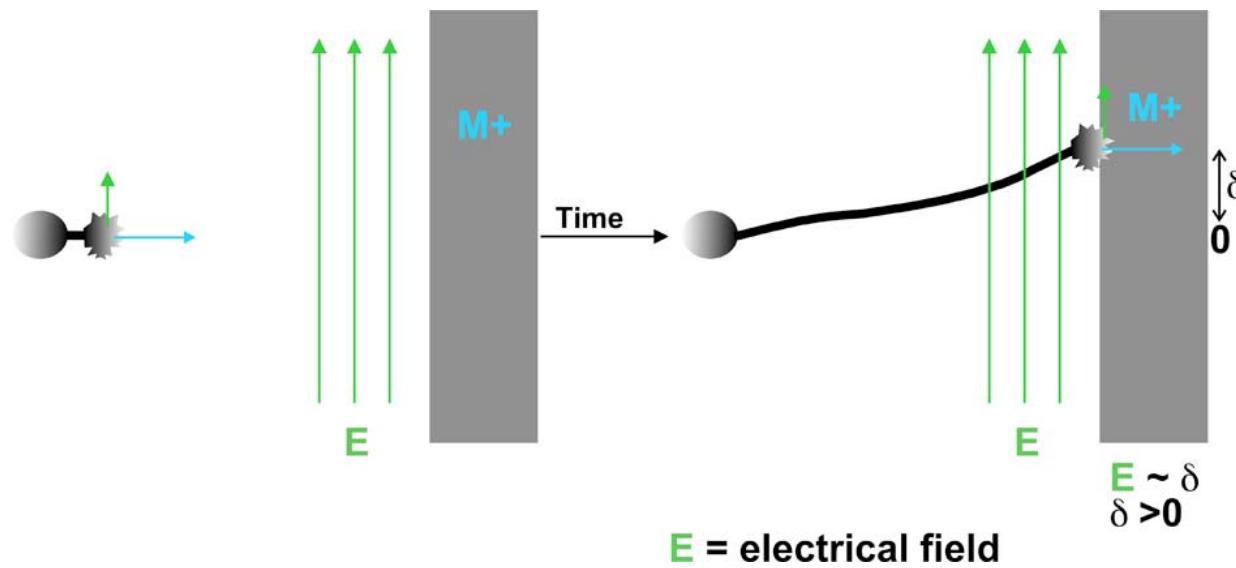
Model value

Explained	Predicted	Agnostic	New prediction
<i>Lmx1b</i>^{-/-} Kania et al., 2000	<i>EphB13</i>^{-/-}	<i>GDNF</i>^{-/-} Kramer et al., 2006	<i>EphA4 & B123</i>^{-/-} complete randomization of both LMCM and LMCI
<i>Bmprla</i>^{flox/-} Luria et al., 2007	<i>EphB123</i>^{-/-}	<i>ret</i>^{-/-} Kramer et al., 2006	
<i>EphA4</i>^{-/-} Helmbacher et al., 2000	<i>ephrin-B2</i>^{flox/-}	<i>Lhx3 Tg</i> Sharma et al., 2000	<i>ephrin-A235</i>^{-/-}
<i>Sema3F</i>^{-/-} Huber et al., 2005			> 2/3 LMCI mistargeted ventrally
<i>Npn2</i>^{-/-} Huber et al., 2005			LMCM normal
<i>EphB1</i>^{-/-} in optic chiasma Williams et al., 2005			
<i>EphB1</i>^{-/-}			

Trajectory changes reveal cue type



Cue strength: quantification using benchmarks

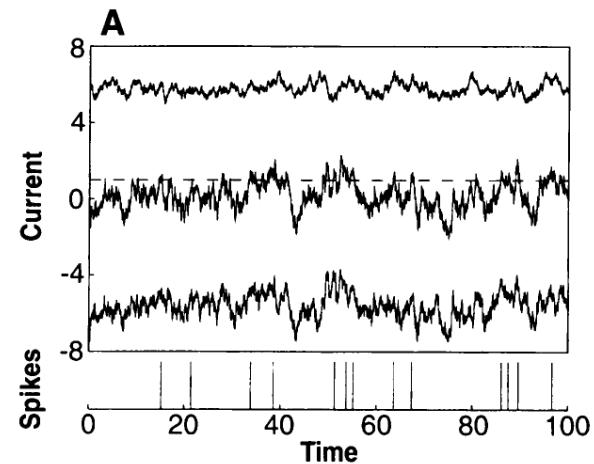


“Too much information” hypothesis: Noise limits the information used for any decision controlled by competing cues

1. Construction of neural circuits

2. Neuronal firing rates

Van Vreeswijk & Sompolinsky, 1996



3. Finance - the anti-portfolio effect

Vlad et al., 2007

4. Drosophila photoreceptor fields - 70% - 30% partitioning of the retina

Wernet et al., 2005

5. ES cells transient differentiation

Niiwa et al., 2008